

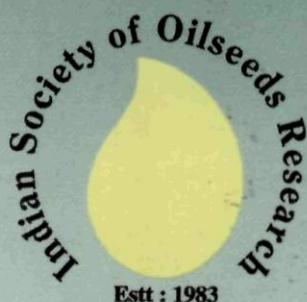
Journal of Oilseeds Research

Volume 26

Number 2

December, 2009

ISSN 0970-2776



**Indian Society of Oilseeds Research
Directorate of Oilseeds Research**

Rajendranagar, Hyderabad-500 030, India

THE INDIAN SOCIETY OF OILSEEDS RESEARCH

(Founded in 1983, Registration Number ISSN 0970-2776)

EXECUTIVE COUNCIL FOR 2008-09

President	:	Dr. M.V. Rao	
Vice-President	:	Dr. D.M. Hegde	
General Secretary	:	Dr. H. Basappa	
Joint Secretary	:	Dr. M.A. Shankar	
Treasurer	:	Dr. R.D. Prasad	
Councillors	:	Dr. Yashpal Yadav	(Northern Zone)
		Dr. M.M. Ansari	(Central Zone)
		Dr. H.V. Nanjappa	(Southern Zone)
		Dr. M.P. Deshmukh	(Western Zone)
		Dr. U.C. Kar	(Eastern Zone)

Editorial Board

Chief Editor	:	Dr. Harvir Singh
Editors	:	Dr. I.Y.L.N. Murthy
Members	:	Dr. S.S. Banga Dr. C.V. Reddy Dr. O.P. Joshi Dr. M.L. Lodha Dr. A. Bandopadhyay Dr. S.D. Kulkarni Dr. Arvind Kumar Dr. S.P. Tiwari Dr. S.J. Kolte Dr. D.R.C. Bakheta Dr. H. Basappa

MEMBERSHIP TARIFF

(w.e.f. 01.01.2007)

Life Membership	Annual Subscription	India	Abroad
Individual : Rs.2500/- + Admn. Fee Rs.50/-	Individual : Institutions : Students :	Rs. 300/- + Admn. Fee Rs.50/- Rs. 1500/- Rs. 200/- + Admn. Fee Rs.50/-	US\$ 100 Ordinary US\$ 150 Institutions

For subscription, please contact • The General Secretary, Indian Society of Oilseeds Research, Directorate of Oilseeds Research, Rajendranagar, Hyderabad-500 030, A.P., India

ADVERTISEMENT TARIFF

Location	Two issues (Rs)	One issue (Rs)
Back cover (in side)	8000/-	5000/-
Page facing back cover	3000/-	1500/-
Inside full page	2500/-	1500/-
Inside half page	1500/-	750/-
Overall size	23 cm height (max.) x 17 cm width (max.)	
1. Back cover & Full page	23 x 17 cm	
2. Half page	11 x 17 cm	

Indian Society of Oilseeds Research
thankfully acknowledges the financial
assistance received from INDIAN
COUNCIL OF AGRICULTURAL RESEARCH,
New Delhi for printing this Journal

Contents

Research Papers

- Heterosis in relation to genetic divergence in parental lines of sunflower, *Helianthus annuus* L.
K.G. Parameshwarappa, B.V. Anand Kumar and S. Neelima ... 89
- Heterosis and combining ability for seed yield, oil content and other quantitative traits in sunflower, *Helianthus annuus* L.
S. Neelima and K.G. Parameshwarappa ... 94
- Inheritance studies in castor, *Ricinus communis* L.
A. Ashok Kumar, P. Janila, M. Sujatha and V. Hemalatha ... 98
- Heterosis for yield and yield contributing traits in castor, *Ricinus communis* L.
Y.M. Barad, A.R. Pathak and B.N. Patel ... 102
- Studies on combining ability for seed yield and yield components in castor, *Ricinus communis* L.
Y.M. Barad, A.R. Pathak and B.N. Patel ... 105
- Combining ability studies for certain quantitative characters in linseed, *Linum usitatissimum* L.
Ram Jeet, P.K. Singh, S.D. Dubey and Amar Singh ... 109
- Productivity of summer groundnut, *Arachis hypogaea* L. as influenced by cumulative residual effect of crop residue incorporation and nitrogen management practices
C. Radhakumari and D. Srinivasulu Reddy ... 114
- Yield attributes, yield, quality and uptake of nutrients by summer groundnut, *Arachis hypogaea* L. as influenced by sources and levels of sulphur under varying irrigation schedules
G.N. Patel, T.P. Patel, P.H. Patel, D.M. Patel, D.K. Patel and R.M. Patel ... 119
- Effect of spacing and nitrogen levels on rabi castor, *Ricinus communis* Linn. grown under different cropping sequences in North Gujarat agro-climatic conditions
R.M. Patel, M.M. Patel and G.N. Patel ... 123
- Effect of potassium fertilization on cationic nutrient content and uptake in seeds of two castor, *Ricinus communis* L. cultivars
I.Y.L.N. Murthy and P. Padmavathi ... 126
- Nutrient requirements of kharif castor, *Ricinus communis* L. under irrigated conditions of Uttar Pradesh
S.K. Srivastava and D.R. Chandra ... 131
- Cultural and morphological variability in *Alternaria brassicae* isolates of Indian mustard, *Brassica juncea* L. Czern & Coss.
Dhiraj Singh, Rajender Singh, Harbinder Singh, Ram Chander Yadav, Neelam Yadav, Martin Barbetti and Phil Salisbury ... 134
- Influence of castor, *Ricinus communis* L. as relay crop in controlling root knot disease of groundnut caused by *Meloidogyne arenaria* (Neal) chitwood
C. Lukose, A.M. Moradia, I.U. Dhruj, R.S. Savalia, L.D. Vavadia and B.A. Kunadia ... 138
- Productivity potentials and profitability of non-monetary, low-cost and cost-effective oilseeds production technologies
R. Venkattakumar, S.V. Ramana Rao, M. Padmaiah and D.M. Hegde ... 140

Short communications

- Genetic divergence in groundnut, *Arachis hypogaea* L.
V.P. Korat, R.H. Kavani, M.S. Pithia, J.J. Savaliya and A.G. Pansuriya ... 148
- Phenotypic evaluation of RILs for rust and late leaf spot and their association on productivity traits in groundnut, *Arachis hypogaea* L.
Sarvamangala Cholin and M.V.C. Gowda ... 151
- Heterosis and genetic architecture for quality traits in sesame, *Sesamum indicum* L.
N.N. Prajapati, C.G. Patel, R.N. Patel, A.M. Patel and K.M. Patel ... 154

Evaluation of progenies selected from random mated population of safflower, <i>Carthamus tinctorius</i> L. using GMS lines <i>P.V. Mahajan, S.N. Deshmukh and R.D. Ratnaparkhi</i>	... 156
Genetic divergence in linseed, <i>Linum usitatissimum</i> L. under salt stress condition <i>R.L. Srivastava, H.C. Singh, Karam Husain, Y.P. Malik and Om Prakash</i>	... 159
Genetic diversity and variability in sesame, <i>Sesamum indicum</i> L. <i>S.R. Kumhar and Z.S. Solanki</i>	... 162
Rajasthan Til 346 : A high yielding white and bold seeded sesame, <i>Sesamum indicum</i> L. variety for National Zone-I of India <i>S.R. Kumhar, Z.S. Solanki, T.S. Rajpurohit, M.M. Sundria and M.S. Chandawat</i>	... 165
Effect of biofertilizer on sunflower, <i>Helianthus annuus</i> L. <i>Madhurendra, N. Prasad and R.K. Akhuri</i>	... 167
Inter-relay cropping of castor in greengram under irrigated condition <i>R.M. Patel, G.N. Patel and S.S. Solanki</i>	... 168
Study on intercropping of castor, <i>Ricinus communis</i> L. under irrigated condition <i>I. Singh</i>	... 170
Efficacy of new insecticides against mustard aphid, <i>Lipaphis erysimi</i> (Kalt.) <i>S.S. Dhaka, Gaje Singh, Y.P.S. Malik and A. Kumar</i>	... 172
Efficacy of bioagents and organic amendments against <i>Macrophomina phaseolina</i> (Tassi) causing root rot of sesame <i>S. Usha Rani, R. Udhayakumar and D. John Christopher</i>	... 173
Multiple resistance sources against major diseases and pests of safflower, <i>Carthamus tinctorius</i> L. <i>D.R. Murumkar, D.V. Indi, V.B. Akashe, A.J. Patil and M.A. Gud</i>	... 175
Seed mycoflora associated with castor, <i>Ricinus communis</i> L. and their effect on germination <i>O. Nagaraja, M. Krishnappa and A.M. Sathisha</i>	... 177
Screening of cold ethyl alcohol extract of <i>Pongamia pinnata</i> for insecticidal properties against <i>Spodoptera litura</i> Fabricius <i>Pratibhav Deshmukhe, Ashok A. Hooli and S.N. Holihosur</i>	... 181
Oil quality characteristics and fatty acid composition of castorbean, <i>Ricinus communis</i> L. cultivars <i>Kamal Kumar Verma, P.S. Kendurkar, Madhu Vajpeyi, Ashish Saini and Neeti Singh</i>	... 183
Development of a bullock drawn groundnut, <i>Arachis hypogaea</i> L. digger suitable for coastal Orissa <i>Jayanarayan Mishra</i>	... 187

Heterosis in relation to genetic divergence in parental lines of sunflower, *Helianthus annuus* L.*

K.G. Parameshwarappa, B.V. Anand Kumar and S. Neelima

Oilseeds Scheme, Main Agril. Research Station, University of Agricultural Sciences, Dharwad-580 005, Karnataka

(Received: July, 2008; Revised: November, 2009; Accepted: December, 2009)

Abstract

Evaluation of 60 hybrids derived from crossing six A lines and 10 restorers in a line x tester programme was carried out to know the extent of genetic divergence among parents and crosses and to relate with heterosis. The results revealed that the parents and hybrids were grouped in 12 clusters indicating substantial variability in the material studied. Almost all the A lines (corresponding B lines) were grouped in cluster I, whereas R lines in clusters II, IV, V, VI, VII, X and XII indicating that R lines are genetically more divergent than A lines. The D^2 values ranged from 97.93 (cluster IV) to 170.16 (cluster XI) supporting the existence of genetic variability in the material. Further, classification of parents and hybrids into four divergence classes revealed that crossing of parents with moderate genetic divergence from DC_2 and DC_3 classes resulted in higher frequency of hybrids with significant heterosis over mid parent of which (CMS-234A x NDOL-3) x RHA-857 (NB), CMS-234A x NDOL-3) x 6.D-1 (Br) and CSM-234A x DSF-2) x RHA-265 (NB) excelled. However, crossing of parents belonging to extreme genetic divergence classes DC_1 and DC_4 resulted in hybrids with non-significant heterosis.

Key words: Sunflower, heterosis, D^2 values, divergence

Introduction

Sunflower being a highly cross pollinated crop is ideal for exploitation of heterosis. The concerted efforts in this direction to develop and evaluate hybrids using different CMS and fertility restorer lines have been made. Plant breeders have extensively used and exploited heterosis to improve seed yield in many crops, more so in sunflower. The value of hybrids and importance of heterosis breeding was recognized very early in sunflower as they are generally more vigorous, uniform and self fertile with high productivity. However, genetic diversity of parents is equally important as pointed by Arunachalam *et al.* (1984) in groundnut and Behl *et al.* (1985) in sunflower. The above findings have established a close correspondence

between the genetic divergence and magnitude of heterosis. It is also being reported that hybrids from genetically diverse parents manifest greater heterosis than that of more closely related parents (Cheres *et al.*, 2000). This seems to be true not always as there are reports suggesting that requisite magnitude of heterosis can be realized when parents with moderate divergence are involved in crosses (Arunachalam *et al.*, 1984 and Behl *et al.*, 1985) and some time the expected heterosis may not be realized when one looks for highly divergent parents for crosses (Cress, 1966), as there may be existence of non correspondence between parents and hybrids on account of environmental factors.

With this background, the present investigation was designed to elucidate the kind of relationship that exists between heterosis and parental diversity in sunflower.

Materials and methods

Investigations were carried out using six CMS lines *viz.*, (VRF x NDOL-2), (851A x NDOL-2), (4546A x DSF-2), (234A x DSF-2), (234A x NDOL-3) and (VRF x DSF-2) in crosses with ten Restorers *viz.*, RHA-274 (Br), RHA-274 (NB), RHA-298 (NB), RHA-857 (NB), RHA-857 (BR), IX-11 (NB), X-13 (NB), V-20 (Br), RHA-265 (NB) and 6 D-1 (Br), DSF-2, NDOL-2 and NDOL-3 to produce 60 F_1 hybrids in a line x tester fashion at the Oilseeds Scheme, Main Agricultural Research Station, University of Agricultural Sciences, Dharwad during rainy season of 2007. These 60 hybrids along with six B lines of corresponding A lines and ten restorers were evaluated in a Randomized Block Design with two replications in summer 2007. Each genotype was provided plot size consisting of single row of 4 metre length with inter and intra row spacing of 60 cm and 30 cm, respectively. Two to three seeds treated with Imidacloprid (@ 5 g/kg of seeds were dibbled/hill, after applying half the recommended dose of N and entire P and K as basal. The remaining N was top dressed when the crop was around 35 days old followed by an inter culture. Thinning excess seedlings to maintain one healthy plant/hill was attended 15 days after complete emergence of the experimental crop. All the recommended agronomic practices were followed to raise a successful experimental crop. The observations were recorded from five random

* Part of Ph.D. (Agril.) Thesis submitted by the senior author to the Department of Genetics and Plant Breeding, UAS, Dharwad-580 005.

but competitive plants on 11 characters viz., days to 50% flowering, days to maturity, plant height (cm), head diameter (cm), number of leaves/plant, test weight (g), hull content (%), volume weight (g/100 cc), per cent seed filling, seed yield (g/plant) and oil content. After computing means, the data was subjected to D^2 analysis. The genotypes were grouped into different clusters according to Toucher's method and inter and intra cluster distances were calculated. Heterosis was estimated as the improvement of F_1 over better parent for each character. The method devised by Arunachalam *et al.* (1984) was used to delineate parental divergence in four divergent classes (DC). To take into account the variable magnitude of variation in parental divergence, the mean (M) and

standard deviation (S), the values of divergence were calculated. The minimum (X) and maximum (Y) values among D^2 values between parents being determined using 'M' and 'S' and the range of D^2 values is being divided into four divergence classes.

Results and discussion

The results of genetic diversity analysis (Table 1) revealed that 76 genotypes comprising both parents and their F_1 's were scattered in as many as 12 clusters. Thus number of clusters formed reflected on genetic divergence in the material. It is interesting to note that all six A lines appeared in cluster I, indicating narrow genetic divergence among them.

Table 1 Clusters formation and genotype composition of the clusters in sunflower

Cluster Number	Genotypes per cluster	Parents and hybrids clustered	Parents/ hybrid
I	08	(VRF x NDOL-2), 851A x NDOL-2), (4546A x DSF-2) (234A x DSF-2), (234A x NDOL-3), (VRF x DSF-2) (VRF x NDOL-2) x RHA-857(Br), (VRF x NDOL-2) x RHA-IX-11(NB)	A lines A lines Hybrids
II	08	(4546A x DSF-2) x RHA-V-20 (Br), (234A x NDOL-3) x RHA-X-13 (NB) (VRF x NDOL-2) x RHA 265(NB), (851A x NDOL-2) x RHA 265 (NB) (VRF x DSF-2) x RHA-X-13 (NB), (234A x DSF-2) x RHA-V-20 (Br) RHA 265 (NB), RHA- 298 (NB)	Hybrids Hybrids Hybrids Restorers
III	12	(VRF x NDOL-2) x RHA-274(NB), (851A x NDOL-2) x RHA-274(NB) (851A x NDOL-2) x RHA-857(Br), (4546A x DSF-2) x RHA-274(Br) (4546A x DSF-2) x RHA-857(Br), (234A x DSF-2) x RHA-857(Br) (234A x DSF-2) x RHA-274(Br), (234A x DSF-2) x 6D-1(Br) (234A x NDOL-3) x RHA-857(NB), (234A x NDOL-3) x 6D-1(Br) (VRF x DSF-2) x RHA- 298 (NB), (VRF x DSF-2) x RHA- 265 (NB)	Hybrids
IV	05	RHA-274(Br), RHA-X-13 (NB) (VRF x NDOL-2) x 6D-1(Br), (4546A x DSF-2) x RHA 265 (NB) (234A x NDOL-3) x RHA-V-20 (Br)	Restorers Hybrids Hybrid
V	13	RHA-V-20 (Br) (VRF x NDOL-2) x RHA-857(Br), (851A x NDOL-2) x RHA-274(Br) (851A x NDOL-2) x RHA- 298 (NB), (851A x NDOL-2) x RHA-V-20 (Br) (4546A x DSF-2) x RHA-X-11(NB), (4546A x DSF-2) x 6D-1(Br) (234A x DSF-2) x RHA-274(NB), (234A x DSF-2) x RHA-X-13 (NB) (234A x NDOL-3) x RHA- 265 (NB), (234A x NDOL-3) x RHA- 298 (NB) (VRF x DSF-2) x RHA- 274(Br), (VRF x DSF-2) x RHA-IX-11(NB)	Restorer Hybrids Hybrids Hybrids Hybrids Hybrids Hybrids
VI	08	RHA-274(NB) (VRF x NDOL-2) x RHA- 298 (NB), (851A x NDOL-2) x RHA-IX-11(NB) (4546A x DSF-2) x RHA-274(NB), (234A x DSF-2) x RHA-857(Br) (234A x NDOL-3) x RHA-274(Br), (VRF x DSF-2) x RHA-857(NB) (VRF x DSF-2) x 6D-1(Br)	Restorer Hybrids Hybrids Hybrid
VII	07	RHA-IX-11(NB) (VRF x NDOL-2) x RHA-V-20 (Br), (851A x NDOL-2) x RHA-X-13 (NB) (4546A x DSF-2) x RHA-X-13 (NB), (234A x DSF-2) x RHA-X-11(NB) (234A x NDOL-3) x RHA-X-11(NB), (VRF x DSF-2) x RHA-857(Br)	Restorer Hybrids Hybrids Hybrids
VIII	03	(851A x NDOL-2) x 6D-1(Br), (234A x DSF-2) x RHA- 265 (NB) (VRF x DSF-2) x RHA-V-20 (Br)	Hybrids
IX	06	(VRF x NDOL-2) x RHA-857(NB), (851A x NDOL-2) x RHA-857(NB) (4546A x DSF-2) x RHA- 298 (NB), (234A x DSF-2) x RHA- 298 (NB) (234A x NDOL-3) x RHA-274(NB), (VRF x DSF-2) x RHA-274(NB)	Hybrids
X	02	RHA-274(Br) (VRF x NDOL-2) x RHA-274(Br)	Restorer Hybrid
XI	02	(4546A x DSF-2) x RHA-857(NB), (234A x NDOL-3) x RHA- 298 (NB)	Hybrids
XII	02	RHA-857(NB), 6D-1(Br)	Restorers

Table 2 D² values and divergence class of parents related to heterosis over mid-parent and better parent in sixty crosses in sunflower

Crosses	Corresponding D ² values		Divergence class	Mid parent heterosis for seed yield/plant	Better parent heterosis for seed yield/plant
	between parents				
(VRF x NDOL-2) x RHA-274 (Br)	408.16		DC ₁	45.56 **	16.13
(VRF x NDOL-2) x RHA-274 (NB)	47.55		DC ₄	19.41	5.43
(VRF x NDOL-2) x RHA-298 (NB)	255.98		DC ₂	0.83	-2.65
(VRF x NDOL-2) x RHA-857 (NB)	174.62		DC ₃	-2.11	-6.28
(VRF x NDOL-2) x RHA-857 (Br)	206.47		DC ₃	18.14	-0.09
(VRF x NDOL-2) x RHAIX-11 (NB)	128.50		DC ₃	10.44	2.84
(VRF x NDOL-2)X-13 (NB)	143.43		DC ₃	3.82	-11.10
(VRF x NDOL-2) x RHAV20 (Br)	442.92		DC ₁	19.50	-4.97
(VRF x NDOL-2) x RHA-26 (Br)	144.23		DC ₃	15.57	13.07
(VRF x NDOL-2) x 6 D-1	305.14		DC ₂	39.48 **	12.37
(851A x NDOL-2) x RHA-274 (Br)	378.07		DC ₂	18.35	-2.66
(851A x NDOL-2) x RHA-274 (NB)	40.74		DC ₄	14.26	4.50
(851A x NDOL-2) x RHA-298 (NB)	288.01		DC ₂	21.78 *	13.24
(851A x NDOL-2) x RHA-857 (NB)	193.01		DC ₃	0.91	0.43
(851A x NDOL-2) x RHA-857 (Br)	186.77		DC ₃	16.07	1.48
(851A x NDOL-2)X-11 (NB)	151.60		DC ₃	3.95	-6.63
(851A x NDOL-2)X-13 (NB)	128.48		DC ₃	40.12 **	24.11
(851A x NDOL-2) x RHAV20 (Br)	432.04		DC ₁	40.22 **	14.92
(851A x NDOL-2) x RHA-265 (NB)	161.86		DC ₃	-3.50	-9.12
(851A x NDOL-2) x 6 D-1	316.18		DC ₂	49.15 **	23.93
(4546A x DSF-2) x RHA-274 (Br)	573.47		DC ₁	12.25	-16.43
(4546A x DSF-2) x RHA-274 (NB)	156.47		DC ₃	-0.96	-19.34 *
(4546A x DSF-2) x RHA-298 (NB)	215.17		DC ₃	-7.01	-12.50
(4546A x DSF-2) x RHA-857 (NB)	259.90		DC ₂	-19.42 *	-29.45 **
(4546A x DSF-2) x RHA-857 (Br)	99.52		DC ₄	6.15	-16.80
(4546A x DSF-2)X-11 (NB)	247.02		DC ₂	-0.04	-2.44
(4546A x DSF-2)X-13 (NB)	530.71		DC ₁	-16.40	-33.74 **
(4546A x DSF-2) x RHAV20 (Br)	211.22		DC ₃	7.68	-20.06 *
(4546A x DSF-2) x RHA-265 (NB)	407.19		DC ₁	-25.43 **	-30.72 **
(4546A x DSF-2) x 6 D-1	490.40		DC ₁	-7.36	-30.43 **
(234A x DSF-2) x RHA-274 (Br)	186.93		DC ₃	36.43 **	13.72
(234A x DSF-2) x RHA-274 (NB)	93.33		DC ₄	-5.46	-12.19
(234A x DSF-2) x RHA-298 (NB)	104.78		DC ₃	-12.82	-20.18
(234A x DSF-2) x RHA-857 (NB)	188.97		DC ₃	-2.60	-3.76
(234A x DSF-2) x RHA-857 (Br)	67.77		DC ₄	6.89	-5.18
(234A x DSF-2)X-11 (NB)	280.77		DC ₂	-3.86	-14.92
(234A x DSF-2)X-13 (NB)	280.78		DC ₂	13.11	1.68
(234A x DSF-2) x RHAV20 (Br)	351.38		DC ₂	9.91	-8.71
(234A x DSF-2) x RHA-265 (NB)	157.06		DC ₃	30.17 **	20.68
(234A x DSF-2) x 6 D-1	266.24		DC ₂	30.05 *	9.54
(234A x NDOL-3) x RHA-274 (Br)	207.70		DC ₃	-0.65	-18.02
(234A x NDOL-3) x RHA-274 (NB)	51.25		DC ₄	27.16 *	16.73
(234A x NDOL-3) x RHA-298 (NB)	317.36		DC ₂	0.06	-7.31
(234A x NDOL-3) x RHA-857 (NB)	139.99		DC ₃	38.90 **	38.81 **
(234A x NDOL-3) x RHA-857 (Br)	85.09		DC ₄	4.11	-8.66
(234A x NDOL-3)X-11 (NB)	202.41		DC ₃	-13.77	-22.83 *
(234A x NDOL-3)X-13 (NB)	108.81		DC ₃	-4.78	-15.36
(234A x NDOL-3) x RHAV20 (Br)	232.30		DC ₃	19.83	-1.47
(234A x NDOL-3) x RHA-265 (NB)	192.07		DC ₃	26.09 *	18.29
(234A x NDOL-3) x 6 D-1	187.97		DC ₃	61.70 **	34.81 **
(VRF x DSF-2) x RHA-274 (Br)	663.32		DC ₁	17.97	-7.73
(VRF x DSF-2) x RHA-274 (NB)	206.41		DC ₃	10.25	-4.88
(VRF x DSF-2) x RHA-298 (NB)	149.73		DC ₃	-1.62	-2.48
(VRF x DSF-2) x RHA857 (NB)	241.41		DC ₃	-17.36	-22.87 *
(VRF x DSF-2) x RHA-857 (Br)	300.76		DC ₂	-3.52	-20.17
(VRF x DSF-2) x RHAIX-11 (NB)	86.36		DC ₄	9.07	4.17
(VRF x DSF-2)X-13 (NB)	331.25		DC ₂	-13.72	-27.74 *
(VRF x DSF-2) x RHAV20 (Br)	551.18		DC ₁	-3.39	-24.68 *
(VRF x DSF-2) x RHA-265 (NB)	173.31		DC ₃	-34.86 **	-35.18 **
(VRF x DSF-2) x 6 D-1	359.78		DC ₂	1.26	-20.05

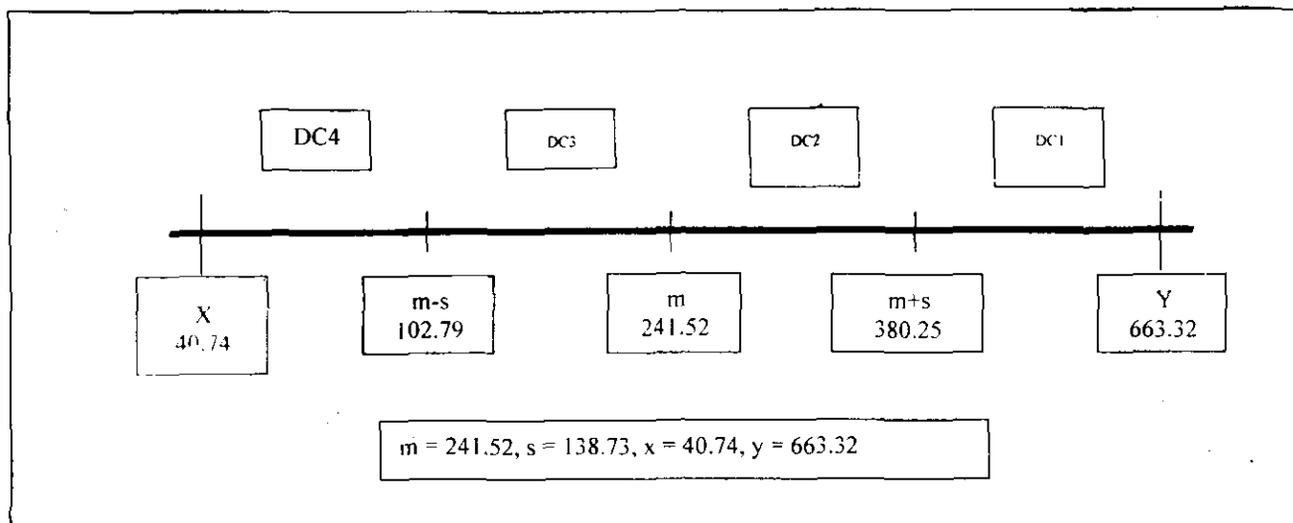


Fig. 1. Classification of parental divergence

This can be attributed to single source of cytoplasm currently employed in sunflower as well as genetically related inbreds in development of CMS lines. Such a narrow genetic variability among A lines appearing within the clusters has also been reported earlier (Teklewold *et al.*, 2000; Ramasubramanyam *et al.*, 2003 and Srinivas *et al.*, 2006). On the other hand, the R lines found to have been grouped in different clusters *viz.*, II, IV, V, VI, VII, X and XII indicating that they are genetically more divergent compared to A lines. Thus, the results clearly reveal that there is a need to diversify A lines for realizing significant standard heterosis. As regards to grouping of 60 hybrids, two clusters III and V formed the major clusters each containing 12 hybrids, which was followed by clusters VI with seven hybrids, clusters II, VII and IX with six hybrids each. The clusters IV had three hybrids, while I and XI two hybrids each. This suggests that the diversity existed even among hybrids in spite of involving genetically similar A lines, perhaps contributed from divergent R lines. The clustering pattern of hybrids is known to be influenced by their parents because of close affinity between the parents and their hybrids (Chaudhary and Singh, 1975).

The crosses were grouped into four divergence classes (Fig. 1) based on parental divergence (Arunachalam *et al.*, 1984), wherein the D^2 values between parental combinations ranged from 40.74 to 663.32. As for genetic divergence of parents in relation to heterosis is considered, the crosses irrespective of the traits appeared in greater frequency in DC₂ and DC₃ compared to DC₁ and DC₄, suggesting that moderate divergence prevailed among parents and crosses (Madrap and Makne, 1993; Lalitha Reddy *et al.*, 2000). As regards to number of hybrids exhibiting significant heterosis over mid parent in different classes it was observed that four hybrids appeared in DC₂, six in DC₃, two hybrids in DC₁ and only one in DC₄. Thus, DC₂ and DC₃ classes together

(moderate divergence) had ten hybrids when compared to three in DC₁ and DC₄ (extreme divergence). This indicates that parents showing moderate genetic divergence can produce higher frequency of heterotic hybrids, which is in agreement with the earlier studies of Arunachalam *et al.* (1984) and Behl *et al.* (1985). Out of 13 hybrids revealing significant heterosis over mid parent, three hybrids *viz.*, (CMS-234A x NDOL-3) x RHA-857 (NB), CMS-234A x NDOL-3) x 6 D1 (Br) and CSM-234A x DSF-2) x RHA-265 (NB), were superior in better parent heterosis (Table 2). So also, the commercial check hybrids KBSH-1, KBSH-44 and DSH-1 exhibited higher standard heterosis and the parents of these hybrids were grouped in moderate divergence classes. The above results clearly demonstrate that there is definite relationship between magnitude of genetic divergence of the parents involved and frequency of heterotic hybrids. On the other hand, the hybrids (CMS-851A x NDOL-2) x RHA-274 (NB) and (CMS-VRF x DSF-2) x RHA-274 (Br) were noticed in low and high divergence classes, respectively, as also their parents. However, relating heterosis with divergence is being contradicted by the reports of Cress (1966) as observed from non-correspondence between parental divergence and heterosis of hybrids in their studies. Although, heterosis is the function of genetic diversity, the positive relationship between heterosis and parental distance depends on several factors including availability of optimum environment for expression of heterosis and extent of internal balance of various components of heterosis. Thus, the present studies clearly suggest that the parents with moderate genetic divergence when crossed are expected to throw heterotic hybrids in higher number in sunflower. Further, the studies point towards diversification of A lines for realizing higher standard heterosis in the experimental hybrids.

References

- Arunachalam, V., Bandopadhyay, A., Nigam, S.N. and Gibbons, R.W. 1984. Heterosis in relation to genetic divergence and specific combining ability in groundnut. (*Arachis hypogaea* L.). *Euphytica*, **33** : 33-39.
- Behl, R.K., Singh, V.P. and Paroda R.S. 1985. Genetic divergence in relation to heterosis and specific combining ability in triticale. *Indian Journal of Genetics*, **45** : 308-375
- Choudhary, B.D. and Singh, V.P. 1975. Genetic divergence in some Indian and exotic barley varieties and their hybrids. *Indian Journal of Genetics*, **35** : 409-413.
- Cheres, M.T., Miller, J.F., Crane, J.M. and Knapp, S.J. 2000. Genetic distance as a predictor of heterosis and hybrid performance within and between heterotic group in sunflower. *Theory and Applied Genetics*, **100** (6) : 889-894.
- Cress, C.E. 1966. Heterosis of the hybrid related to gene frequency differences between two populations. *Genetics*, **53** : 269-274.
- Lalitha Reddy, S.S., Sheriff, R.A., Ramesh, S. and Mohan Rao, A. 2000. Exploring possible limits to parental divergence for the occurrence of heterosis in sesame (*Sesamum indicum* L.). *Crop Research*, **9** : 135-139.
- Madrap, I.A. and Makne, V.G. 1993. Heterosis in relation to combining ability effect and phenotypic stability in sunflower. *Indian Journal of Agricultural Sciences*, **63** (8) : 484-488.
- Ramasubramanyam, S.V., Kumar, S.S. and Raganatha, A.R.G. 2003. Genetic divergence for seed parameters in sunflower (*Helianthus annuus* L.). *Helia*, **26** : 73-80.
- Srinivas, B., Dangi, K.S., Drayaga, H. and Kumar, S.S. 2006. Genetic divergence studies in sunflower, *Heliantus annuus* L. germplasm lines. *Journal of Oilseeds Research*, **23** (1) : 41-51.
- Teklewold, A., Jayaramaiah, H. and Jayaramgowda. 2000. Genetic divergence study in sunflower (*Helianthus annuus* L.). *Helia*, **23** : 93-104.

Heterosis and combining ability for seed yield, oil content and other quantitative traits in sunflower, *Helianthus annuus* L.*

S. Neelima and K.G. Parameshwarappa

Oilseeds Scheme, Main Agril. Research Station, University of Agricultural Sciences, Dharwad-580 005, Karnataka

(Received: December, 2008; Revised: November, 2009; Accepted: December, 2009)

Abstract

The genetic analysis of line x tester crosses forming 30 hybrids from five CMS lines and six restorers revealed that SCA variances were predominant in the inheritance of seed yield, oil content and for most of the quantitative traits indicating the role of non-additive genes. The parents CMS 17 A and DCMS 51A found to be good general combiners for seed yield, hull content, volume weight, number of filled seeds, 100 seed weight, days to maturity, number of leaves/plant, head diameter, plant height, days to flowering and oil content, while CMS 234 A was a good general combiner for hull content, oil content and volume weight. The tester RHA-6D-1 exhibited high *gca* for 100-seed weight, per cent seed filling and oil content, while R-298 for number of filled seeds/head, volume weight and seed yield. The hybrid CMS 234A x RHA 95-C-1 had significant *sca* effects for seed yield and number of filled seeds/head. It also shown the highest better parental heterosis followed by CMS 234 A x RHA-6D-1, CMS 234 A x RHA-271, CMS 234 A x R-64 and CMS 234 A x R-298 for seed yield. All these hybrids found to have CMS 234 A as one of the parents and both parents in these crosses had low *gca* effects. The hybrids CMS 17 A x R-298, CMS 103 A x RHA-272, CMS 4546 A x RHA-271 and CMS 4546 A x RHA-272 exhibited significant better parental heterosis for oil content. However, none of the hybrids expressed positive and significant heterosis for both seed yield and oil content.

Key words: Line x tester, heterosis, combining ability

Introduction

Sunflower is an important oilseed crop which can be grown in varied climatic and soil conditions in any season of the year. A high degree of cross pollination coupled with availability of efficient cytoplasmic male sterile and restorer systems offers considerable scope for commercial exploitation of heterosis. The development and release of first ever sunflower hybrid BSH-1 (Seetharam *et al.*, 1980) using male sterility system gave

a fillip to large scale cultivation in India. The hybrids synthesized and released both by public and private for commercial cultivation are predominantly Single Crosses (SC), where uniformity is a distinct advantage. The knowledge on the combining ability of parents, nature of gene action governing seed yield and its component characters and magnitude of heterosis are important for commercial exploitation of heterosis (Rajanna *et al.*, 2001). An attempt has been made in the present investigations to know the extent of standard heterosis and combining ability of parents and crosses for important component traits.

Materials and methods

Five CMS lines *viz.*, CMS 234 A, CMS 4546 A, CMS 17 A, CMS 103 A and DCMS-51A were crossed with six restorer lines *viz.*, RHA-6D-1, RHA-95-C-1, R-298, R-64, RHA-271 and RHA-272 in L x T fashion to generate 30 single cross hybrids during *kharif* 2005. The hybrids were evaluated along with 11 parents during *kharif* 2006 at MARS, UAS, and Dharwad in a Randomized Complete Block Design with three replications. Each genotype was represented by three rows of 3 m length with inter and intra row spacing of 60 cm and 30cm, respectively. All the recommended agronomic practices were followed to raise the crop successfully. Observations were recorded on five random but competitive plants for 12 quantitative characters *viz.*, days to 50% flowering, plant height, head diameter, number of leaves/plant, days to maturity, 100 seed weight, per cent seed filling, number of filled seeds/head, volume weight, hull content, oil content and seed yield/plant. Mean values were subjected to line x tester analysis to estimate general combining ability (*gca*) and specific combining ability (*sca*) effects and their respective variances. The significance of heterosis was tested by comparing the estimates against standard errors using 't' test

Results and discussion

The analysis of variance for combining ability (Table 1) revealed that significant differences existed among CMS lines for all the characters, while among restorers for days to flowering, plant height, number of leaves/plant, days to maturity and volume weight indicating presence of

* Part of Ph.D. (Agril.) Thesis submitted by the senior author to the Department of Genetics and Plant Breeding, UAS, Dharwad-580 005.

considerable variability in the material. The line x tester interactions were highly significant for all the characters except days to 50% flowering and number of leaves/plant indicating that lines and testers have nick well ability in the hybrids synthesized and in turn in the expression of heterosis and combining ability. The variances due to specific combining ability (SCA) were higher than that of general combining ability (GCA) for most of the characters studied except days to flowering, which indicates that these characters are influenced by non-additive genes in their inheritance and suggest for exploitation through heterosis breeding. The predominant role of non-additive component of variance in the inheritance of seed yield, oil content and yield related traits has also been reported by Vara Prasad *et al.* (2006), Rukmini Devi *et al.* (2005) and Parameswari *et al.* (2004).

Among different CMS lines used in the development of hybrids, two lines CMS 17 A (parent of KBSH 44) and DCMS 51 A observed to be as good general combiners (Table 2) for number of traits such as seed yield/plant, hull content, volume weight, number of filled seeds, 100 seed weight, days to maturity, number of leaves/plant, head diameter and plant height. This amply suggests that these lines have been bred for good *gca* for more than one trait. However, no parent appeared to be a good general combiner for important traits like seed yield and oil content except DCMS 51A and leads to the opinion that it can be extensively used in crosses whenever the objective is to improve seed yield and oil content simultaneously. CMS 234 A, the parent of KBSH 1 appeared to be a good general combiner for hull content, oil content and volume weight but not for seed yield. It is also a good general combiner for days to 50% flowering, days to maturity and number of leaves/plant. The line CMS 103 A (parent of RSFH 1) is a good combiner for hull content, whereas

CMS 4546 A is a poor combiner in respect of majority of the traits studied, implying that they may not yield heterotic hybrids. The study clearly revealed that CMS 17 A, DCMS 51 A and CMS 234 A are the potential CMS lines for developing experimental with superior heterosis for seed yield and oil content. Among the restorers, RHA-6D-1 exhibited high *gca* for 100 seed weight, per cent seed filling and oil content, while R-298 for number of filled seeds/head, volume weight and seed yield and R-64 for days to 50% flowering, plant height and days to maturity and these findings are in accordance with the earlier reports by Shekhar *et al.* (2000) and Madhavi Latha *et al.* (2005). Irrespective of the CMS or R lines, the characters are known to be influenced by additive genes, which suggest that the new parental lines can be developed from the heterotic pools having good general combining ability.

Among various hybrids studied, CMS 234A x RHA 95-C-1 expressed significant *sca* effects (Table 2) for seed yield and number of filled seeds/head, however, the parents involved in the cross were poor general combiners. The possibility of getting crosses with significant *sca* involving poor general combiners for a given trait has been reported earlier (Radhika *et al.*, 2001; and Uttam *et al.*, 2005). The hybrids CMS 17A x R-298 and CMS 103 A x RHA 272 observed to be good specific combiners for oil content, while CMS 234 A x R-64 and CMS 4546A x RHA 95-C-1 for per cent seed filling and in all the crosses parents involved exhibited L x L *gca* effects. The hybrid CMS 234A x RHA 95-C-1 was a good specific combiner for seed yield, per cent seed filling and number of filled seeds. It is interesting to note that the parents with good *gca* for a given trait may not end up always in significant *sca* of the crosses.

Table 1 Estimates of general combining ability (*gca*) effects of parents for 12 characters in sunflower

Source	Days to 50% flowering	Plant height (cm)	Head diameter (cm)	No. of leaves/plant	Days to maturity	100 seed weight (g)	% seed filling	No. of filled seeds/head	Volume weight (g/100 cc)	Hull content (%)	Oil content (%)	Seed yield/plant (g)
Lines												
CMS 234A	-2.02 **	-1.71	-0.73 **	1.47 **	-2.67 **	-0.49 **	-0.94	-36.87 **	1.17 *	-2.27**	0.46	-0.61
CMS 4546A	-3.02 **	-10.61 **	-1.24 **	-3.05 **	-2.83 **	-0.20 *	-1.68	-36.34 **	-1.01	-0.42**	-0.01	-4.33 **
CMS 17A	2.59 **	16.52 **	0.69 **	1.90 **	3.78 **	0.86 **	-1.09	80.78 **	2.89 **	4.67**	-2.40 **	4.42 **
CMS 103A	0.26	-26.84 **	0.22	-1.64 **	-0.17	-0.60 **	-1.51	-51.92 **	-4.24 **	1.42**	0.05	-2.59
DCMS 51A	2.20 **	22.64 **	1.06 **	1.33 **	1.89 **	0.43 **	5.22 **	44.35 **	1.20 *	-3.4**	1.90 **	3.12 *
SE (Si)	0.271	1.4482	0.1735	0.4234	0.3846	0.0891	1.1343	9.4195	0.5372	0.1469	0.2518	1.3685
Testers												
RHA 6D-1	0.01	0.12	-0.32	-1.19 *	1.53 **	0.25 *	3.22 *	17.02	0.92	0.81**	1.32 **	0.03
RHA 95-C-1	0.74 *	11.27 **	0.09	-0.31	4.33 **	-0.10	1.54	-4.80	1.15	2.39**	0.10	0.15
R-298	-3.12 **	-7.85 **	0.11	0.14	-4.80 **	0.18	1.98	34.05 **	1.75 **	-0.75**	-1.06 **	4.23 **
R-64	4.88 **	7.14 **	0.34	-1.15 *	6.13 **	-0.10	-2.16	-32.99 **	-4.72 **	-0.46**	-0.77 **	-4.77 **
RHA-271	-0.92 **	-5.20 **	-0.13	1.64 **	-4.53 **	-0.34 **	-2.25	-15.26	0.29	-2.14**	0.39	1.35
RHA-272	-1.59 **	-5.48 **	-0.09	0.88	-2.67 **	0.10	-2.33	1.98	0.63	0.15	0.02	-1.01
SE (Sj)	0.2968	1.5864	0.1901	0.4638	0.4213	0.0976	1.2426	10.31866	0.5884	0.1609	0.2758	1.4991

**, * = Significant at 5% and 1% level, respectively

Table 2 Selected crosses with superior heterosis over better parent, *sca* and *gca* effects and mean performance

Character	Crosses with high heterotic performance over better parent	Heterosis over better parent	<i>sca</i> effects	<i>gca</i> status		<i>Per se</i> performance
				Female	Male	
Seed yield/ plant (g)	CMS 234 A x RHA-95-C-1	104.01**	7.94**	L	L	70.89
	CMS 234 A x RHA-6D-1	86.06**	1.83	L	L	64.66
	CMS 234 A x RHA-271	85.15**	0.19	L	L	64.34
	CMS 234 A x R-64	82.22**	5.29	L	L	63.32
	CMS 234 A x R-298	80.49**	-2.85	L	L	64.18
	CMS 17 A x R-298	10.97**	2.47**	L	L	37.77
Oil content (%)	CMS 103 A x RHA-272	10.47**	1.96**	L	L	40.80
	CMS 4546 A x RHA-271	7.96**	0.19	L	L	39.33
	CMS 4546 A x RHA-272	7.69**	0.42	L	L	39.20
	CMS 234 A x RHA-272	-20.73**	-4.55**	H	L	24.17
Hull content (%)	CMS 234 A x R-64	-15.89**	-2.46**	H	H	25.65
	DCMS 51 A x R-298	-15.87**	-1.14**	H	H	25.54
	CMS 4546 A x RHA-271	-14.88**	-2.57**	H	H	25.70
Vol. wt (g/100 cc)	CMS 4546 A x R-298	14.96**	0.90	L	H	38.89
	CMS 234 A x RHA-95-C-1	30.68**	5.08	L	L	93.26
Per cent seed filling	CMS 234 A x R-64	21.79**	6.65*	L	L	91.13
	CMS 4546 A x RHA-95-C-1	20.97**	5.50*	L	L	92.94
	CMS 4546 A x RHA-6D-1	17.56**	1.21	L	H	90.32
	CMS 17 A x RHA-6D-1	101.74**	58.82*	H	L	496.98
	CMS 234 A x RHA-95-C-1	94.24**	56.46*	L	L	355.15
No. of filled seeds/head	CMS 17 A x R-298	82.40**	-5.86	H	H	449.34
	CMS 4546 A x RHA-272	78.08**	32.83	L	L	338.83
	CMS 17 A x RHA-6D-1	29.79**	0.18	H	H	6.63
	CMS 17 A x RHA-95-C-1	22.75**	0.17	H	L	6.27
100 seed weight (g)	CMS 17 A x RHA-272	22.00**	-0.07	H	L	6.23
	CMS 17 A x R-298	16.19**	-0.04	H	L	6.34
	CMS 103 A x RHA-271	-13.15**	-1.30	L	H	83.67
	CMS 103 A x R-298	-12.11**	-0.03	L	H	84.67
	DCMS 51 A x R-298	-11.97**	-3.42**	L	H	83.33
Days to maturity	CMS 103 A x RHA-272	-11.76**	-1.83	L	H	85.00
	CMS 4546 A x RHA-272	28.23**	0.95*	L	L	15.94
	CMS 4546 A x RHA-271	23.83**	0.45	L	L	15.40
	CMS 234 A x RHA-271	22.54**	0.47	L	L	15.93
Head diameter (cm)	CMS 234 A x RHA-272	21.67**	0.31	L	L	15.82
	CMS 234 A x RHA-271	32.01**	0.49	H	H	35.60
	CMS 234 A x RHA-95-C-1	31.55**	0.47	H	L	33.63
	CMS 17 A x RHA-271	30.34**	0.69	H	H	36.23
No. of leaves/plant	CMS 234 A x RHA-95-C-1	50.70**	-2.22	L	H	216.70
	CMS 234 A x R-64	49.75**	2.54	L	H	217.33
	CMS 4546 A x RHA-95-C-1	42.78**	1.77	L	H	211.78
	CMS 103 A x R-298	-15.92**	0.68	L	H	56.33
Days to flowering	CMS 17 A x R-298	-15.27**	-0.66	L	H	57.33
	CMS 103 A x RHA-271	-14.93**	-0.86	L	H	57.00

On the other hand the crosses with significant *sca* also arose as a result of crosses between low x low or high x low general combiners. Considering hybrids with significant heterosis over better parent, combining ability and *per se* performance, five crosses CMS 234 A x RHA-95-C-1, CMS 234 A x RHA-6D-1, CMS 234 A x RHA-271, CMS 234 A x R-64 and CMS 234 A x R-298 exhibited significant heterosis over better parent. Further, it is noticed that all these hybrids had CMS 234 A as one of the parents in cross combinations. This suggests that CMS 234 A is a potential CMS line in realizing superior heterosis (104.0 %) for seed yield as seen from cross combination of CMS 234 A x RHA 95C-1. The prevalence of substantial magnitude of heterosis over better parent for seed yield has also been reported by Singh and Singh (2003) and Rajeswari *et al.* (2005). The hybrids CMS 17 A x R-298, CMS 103 A x RHA-272, CMS 4546 A x RHA-271 and CMS 4546 A x RHA-272 exhibited significant better parental heterosis for oil content (Nehru *et al.*, 2000). However, none of the hybrids expressed positive and significant heterosis for both seed yield and oil content. This suggests that there is a need to look for parents (A and R lines) with good *gca* for both yield and oil content and crosses produced from such parents have tendency to show correlated response. Nevertheless, manifestation of heterosis can mainly be attributed to the role of non-additive gene actions as seen from predominant proportion of SCA variance for the traits. As regards to heterosis for component traits, CMS 17 A in cross combination with RHA-6D-1 and RHA-95-C-1 has thrown out positive and significant heterosis for hull content, which involved parents mostly with high x high *gca* effects. Involving low x low general combiners in four crosses CMS 234A x RHA- 271, CMS 234A x RHA- 272, 4546 A x RHA-271 and 4546 A x RHA- 271 for head diameter and three crosses CMS 234A x RHA 95C-1, CMS 234A x R-64 and 4546 A x RHA 6D-1 for per cent seed filling resulted in positive and significant heterosis. The heterotic hybrids are also realized in respects of 100 seed weight, plant height and days to maturity as a result of crosses between L x H or H x L general combiners. Therefore, the pattern of *gca* of parents in cross combinations enables to predict heterosis during trait based improvements. The present investigation revealed the highest magnitude of heterosis for seed yield/plant followed by number of filled seeds/head and the least heterosis for oil content. The highest heterosis to the extent of 104% for seed yield over better parent in the cross CMS 234 A x RHA-95-C-1 is attributed to the contribution of heterosis from component characters such as hull content, per cent seed filling, number of filled seeds/head and earliness (Radhika *et al.*, 2001). From the present study it could be concluded that CMS 234 A, DCMS 51 A and CMS 17 A among the CMS lines and R-298 and RHA-6D-1 among the restorers are

the potential parental lines for developing superior experimental hybrids.

References

- Madhavi Latha, K., Reddy, A. V. and Reddy, G.L. 2005. Studies on genetic nature of yield and its components in sunflower (*Helianthus annuus* L.). *Journal of Oilseeds Research*, **21** (2): 252-256.
- Nehru, S.D., Eswarappa, G., Rangaiah, S. and Kulkarni, R.S. 2000. Studies on heterosis for seed yield and oil content to develop hybrids with high oil yield in sunflower (*Helianthus annuus* L.), *Mysore Journal of Agricultural Sciences*, **34** (1): 1-5.
- Parameswari, C., Muralidharan, V., Subbalakshmi, B. and Manivannan, M. 2004. Genetic analysis of yield and important traits in sunflower (*Helianthus annuus* L.) hybrids. *Journal of Oilseeds Research*, **21** (1): 168-170.
- Radhika, P., Jagadeshwar, K. and Khan, H.A. 2001. Heterosis and combining ability through line x tester analysis in sunflower (*Helianthus annuus* L.). *Journal of Research ANGRAU*, **29** (2): 35-43.
- Rajanna, M.P., Seetharam, A., Virupakshappa, K. and Ramesh, S. 2001. Heterosis in top-cross hybrids of diverse cytotsterile sources of sunflower (*Helianthus annuus* L.). *Helia*, **24** (34): 25-33.
- Rajeswari, U., Reddy, A.V., Rama Kumar, P.V. and Srinivasa Rao, V. 2005. Heterosis for seed yield and components in Sunflower (*Helianthus annuus* L.). *The Andhra Agricultural Journal*, **52** (1&2): 40-42.
- Rukmini Devi, K.R., Ranganatha, A.R.G. and Ganesh, M. 2005. Combining ability and heterosis for seed yield and its attributes in sunflower. *Agricultural Sciences Digest*, **25** (1): 11-14.
- Seetharam, A., Giriraj, K. and Kusuma Kumari, P. 1980. Phenotypic stability of seed yield in sunflower hybrids. *Indian Journal of Genetics and Plant Breeding*, **40**: 102-104.
- Shekhar, G.C., Jayaramgowda, Nehru, S.D., Halaswamy, B.H. and Ashok, 2000. Combining ability of CMS 234A x RHA 95-C-1 early maturing CMS lines and restorers in sunflower. *Mysore Journal of Agricultural Sciences*, **34**: 289-293.
- Singh, D.P. and Singh, S.B. 2003. Heterosis for seed yield and its components in sunflower (*Helianthus annuus* L.). *Journal of Oilseeds Research*, **20** (1): 40-41.
- Uttam, K.K., Sheoran, R.K. and Rakesh Kumar, 2005. Combining ability analysis in sunflower (*Helianthus annuus* L.). *National Journal of Plant Improvement*, **7** (1): 35-39.
- Vara Prasad, B.V., Reddy, A. V. and Nageswara Rao, T. 2006. Combining ability studies for seed yield and yield contributing characters in sunflower, (*Helianthus annuus* L.). *Journal of Oilseeds Research*, **23** (1): 86-88.

Inheritance studies in castor, *Ricinus communis* L.

A. Ashok Kumar, P. Janila, M. Sujatha and V. Hemalatha

Regional Agricultural Research Station, Acharya N.G. Ranga Agril. University, Palem-509 215, Mahaboobnagar Dist., AP

(Received: November, 2006; Revised: September, 2009; Accepted: October, 2009)

Abstract

In castor, *Ricinus communis* L. inheritance of anthocyanin pigment on stem (APS), bloom, capsule spiny and cup shape of leaves is monogenic. Capsule spiny is a co-dominant character and bloom is controlled by multiple alleles. Inheritance of papaya leaf lacination is digenic and shows dominant epistasis. Co-segregation studies revealed absence of linkage between the characters bloom, leaf shape, and APS.

Key words: Castor, inheritance, monogenic, multiple alleles, dominant epistasis and linkage

Introduction

Castor, *Ricinus communis* L. is an important oilseed crop of India, grown widely in the states of Gujarat and Andhra Pradesh. India is one of the centres of origin and large diversity can be seen in wild, semi-wild and cultivated forms. Extensive breeding efforts have resulted in release of some excellent varieties and hybrids. However, the genetics of the crop is poorly understood.

Though a large number of contrasting characters are available in castor, the knowledge about mode of inheritance and genetic linkage of characters is very scanty. In the present study, we attempted to understand inheritance of some morphological characters viz., anthocyanin pigment on stem (APS), bloom on aerial plant parts, spiny of capsules, cup shape of leaf and papaya leaf lacination. Co-segregation of the character pair's viz., bloom-leaf shape, APS-bloom and APS-leaf shape was also studied.

Materials and methods

Plant material and crossing: Nine strains of castor were used to make six crosses. The source and distinct morphological characters of the parents are given in Table 1. The parental strains were crossed in the combinations: Kranthi x PCS-122, Jyoti x PCS-122, VP-1 x Kiran, Haritha x PCS-122, DPC-9 x Haritha and LRES-17 x PCS-137. A total of 1790 F₂ plants belonging to six crosses were studied for present investigation. Crossing and selfing is accomplished manually by hand crossing and bagging of spikes. The study was conducted at Regional Agricultural Research Station, Palem during 2002 to 2004.

Description of traits and observations

Anthocyanin pigment on stem (APS): Anthocyanin pigments are plant pigments called flavonoids, which are classified as secondary metabolites and are ubiquitous among flowering plants. The anthocyanin pigment imparts colours like pink, red, blue, and purple to the plants. In Castor strains can be distinguished based on expression of anthocyanin pigment and absence of pigment. Observations were recorded on 45 days old plants. Anthocyanin pigment in general is found in stem, petiole, and young leaves. An extreme purple morphotype expressing intense anthocyanin pigment in all aerial plant parts was also reported (Anjani, 2005).

Bloom: Castor plants have white waxy coating on aerial parts, which is generally referred as bloom. It imparts certain level of tolerance to sucking pests, whereas in the zero bloom (absence of bloom), plants are highly susceptible to jassids and triple bloom genotypes are more tolerant to drought compared to zero and single bloom. Four distinct classes are identified based on the presence/absence of bloom on different parts, viz., triple (bloom on stem, petiole, lower and upper surface of leaf), double (on stem, petiole and lower surface of leaf), single (on stem only) and zero (absence of bloom). Observations are preferably taken in the early hours of the day, and to record bloom on upper surface of leaves, only young emerging leaves are observed.

Spiny of capsules: Spiny is a easily distinguishable morphological trait. Both types of capsules i.e., spiny and non-spiny are common in castor. Observations were recorded on developed capsules of the spike.

Leaf shape: The leaves of the strains VP-1 and LRES-17 have distinct cup shape, unlike the flat normal leaves. The 4th or 5th leaf from top is preferred for taking observations on leaf shape.

Leaf lacination: The strain PCS-122 has a distinct leaf lacination, popularly known as papaya leaf (deep lacination), which is distinguishable from normal (shallow) lacination. Similar to leaf shape, observations on leaf lacination were recorded on 4th or 5th leaf from top; young leaves were avoided.

The data collected were subjected to statistical analysis. Standard statistical procedures (Panse and Sukhatme, 1989) were used for fitting F₂ segregation ratios and

testing for goodness of fit using Chi-square test. Detection of linkage is done by calculating Chi-square due to linkage ($2L$) for 9:3:3:1 ratio (Mather, 1951).

Results and discussion

The dominance/recessive relation of the characters is revealed by F_1 phenotype (Table 2). The character APS present, flat shape of leaf, and normal lacination are dominant over their counterparts APS absent, cup shape of leaf, and papaya lacination, respectively. For bloom, triple is dominant over double and double is dominant over single and zero bloom. The F_1 phenotype for spiny nature of capsules is intermediate, i.e., sparsely spiny, indicating co-dominance nature of inheritance of these trait.

A total of 1790 plants belonging to six crosses were phenotyped to carry out inheritance and linkage studies. The F_2 segregation of the traits is given in Table 2. Segregation for APS is studied in 3 crosses viz., Kranthi x PCS-122, Jyoti x PCS-122 and VP-1 x Kiran. The F_2 segregation in these crosses showed goodness of fit for 3 APS present: 1 APS absent ratio though the range of probability varied among the crosses. Pooled data of these crosses showed a segregation of 785 APS present: 257 APS absent, which has a goodness of fit ($p=0.8-0.9$) for 3:1 ratio. APS present of castor plants is found to be under the genetic control of at least one pair of alleles such that A-results in the production of anthocyanin pigment (red stem). The recessive genotype (*aa*) lacks this pigment and normal green colour is exhibited.

Inheritance of stem colour was reported to be monogenic by Solanki and Joshi (2001), which is in agreement with our findings. However, we observed differences in the intensity of the pigment among the F_2 segregants. The pigmentation ranged from specks of red colour anthocyanin to intense pigment. More over, the pigmented F_2 plants could not be grouped into discrete classes with naked eye based on extent of pigmentation.

In plants like *Petunia* (Quattrocchio *et al.*, 1993) and rice (Reddy, 1996) polygenes governing the production of anthocyanin pigment was reported, further its accumulation is been directly correlated to exposure to bright sunshine (Moshkin, 1986). Thus in Castor too, the observed variation for accumulation of anthocyanin may also be attributed to polygenes and environmental influence and complete absence of pigment in some genotypes may be hypothesised to be due to loss of function of a single upstream regulatory element governing the expression of anthocyanin.

Spinyess of capsules is found to be co-dominant trait; the F_1 plants showed intermediate phenotype i.e., sparsely spiny capsules. The F_2 segregation of the cross VP-1 x Kiran showed goodness of fit for 1 spiny: 2 sparsely spiny: 1 non-spiny ($p=0.5-0.7$). The recessive character is governed by single recessive gene, thus the recessive

homozygotes *ss* are non-spiny; heterozygotes *Ss* produce sparsely spiny phenotype; and the dominant homozygotes *SS* results in spiny capsules. The inheritance of this character is in agreement with previous report. (Anjani, 1997). However, similar to the APS, large variation was observed in the expression of sparsely spiny character, nevertheless the trait can be distinguished from: the spiny capsules without ambiguity. It was also observed that the capsules of the same spike also showed variation for sparsely spinyess. However, discrete classes could not be made for the observed variation. Thus it may be concluded that the expression of the alleles in heterozygotes is either influenced by environment or show variable expressivity.

The segregation data of two crosses viz., VP-1 x Kiran and LRES-17 x PCS-137 indicates dominances of triple over double bloom and double bloom is governed by single recessive locus. The F_2 segregation studies on pooled data of the two crosses showed goodness of fit for 3:1 ratio ($p=0.5-0.7$). So also the individual F_2 segregation of the crosses VP-1 x Kiran and LRES-17 x PCS-137 showed good ness of fit for 3:1 ratio ($p=0.1-0.2$). Based on F_2 segregation data of 3 crosses viz., Kranthi x PCS-122, Jyoti x PCS-122 and Haritha x PCS-122 it is concluded that single bloom is governed by single recessive locus. In all the 3 crosses the F_2 segregation showed a goodness of fit for 3:1 ratio and the same is true for pooled data over three crosses ($p=0.7$). The F_2 segregation of the cross DPC-9 x Haritha also showed goodness of fit for 3:1 ratio ($p=0.5-0.7$), which suggests that the recessive trait zero bloom is also governed by single locus.

Solanki and Joshi studied inheritance of bloom by taking presence of bloom and absence of bloom as two variants and concluded monogenic inheritance (Solanki and Joshi 2001). In our study, we attempted a detailed study by making crosses involving variants within presence of bloom viz., single, double and triple. The results indicate that triple bloom is dominant over double bloom and both single and zero bloom are recessive to double bloom. Thus, we may hypothesise presence of at least four multiple alleles viz., *b1*, *b2*, *b3* and *b4* belonging to single locus for the character bloom.

Flat leaf is found to be dominant over cup shape leaf, and cup shape leaf is governed by recessive allele of the locus. The F_2 segregation for this character is studied in 2 crosses viz., VP-1 x Kiran and LRES-17 x PCS-137 and in both of them it showed a goodness of fit for 3:1 ratio ($p=0.3-0.5$). The pooled data also had a goodness of fit for 3:1 ratio ($p=0.2$).

Papaya leaf is recessive to normal leaf lacination and is governed by two loci. Digenic inheritance was confirmed by appearance of papaya leaf F_2 plants, which amount to 1/16 of total F_2 population. The F_1 phenotype is normal leaf lacination; while the F_2 plants expressed a third phenotype i.e., intermediate lacination in addition to parental types.

Inheritance studies in castor

The character segregation is studied in two crosses viz., Kranthi x PCS-122 and Jyoti x PCS-122 and in both the crosses F₂ segregation showed a goodness of fit to a ratio of 12 normal: 3 intermediate: 1 papaya ($p=0.5-0.7$ and $0.2-0.3$, respectively), indicating dominant epistasis. The genotypes *A-B-* and *A-bb* produce normal leaf; *aaB-* produces intermediate leaf and *aabb* produces papaya leaf. The locus *A* is epistatic over *B* locus, thus producing normal phenotype irrespective of alleles at *B* locus.

We studied co-segregation of three-character pairs viz., bloom - leaf shape, APS - bloom and APS - leaf shape (Table 3). The parents LRES-17 and VP-1 possess triple bloom and cup shaped leaves and their co-segregation was studied in two crosses viz., LRES-17 x PCS-137 and VP-1 x Kiran. In both the crosses the calculated Chi-square was non-significant indicating independent assortment of the characters triple bloom and cup shape leaves with good ness of fit to 9:3:3:1 ratio.

In the crosses Kranthi x PCS-122 and Jyoti x PCS-122, co-segregation of the characters presence of APS and double bloom was studied. The Chi-square was non-significant indicating goodness of fit for 9:3:3:1 ratio, thus it is concluded that the characters APS present and double bloom also assort independently. Similar were the results from the cross VP-1 x Kiran for the characters APS absent and triple bloom, where in the 2L was also non-significant. Independent assortment of the characters APS absent and cup shaped leaves was confirmed based on the F₂ co-segregation data of the cross VP-1 x Kiran.

Thus, co-segregation studies observed no linkages for the characters under study. However, more studies on inheritance and linkage analysis are required, to understand genetics of Castor and the role of environment on the expression of characters.

Table 1 Morphological characteristics of castor genotypes

Genotype	Source	Stem colour	Bloom (waxy coating)	Spinyess of capsules	Leaf lacination	Leaf shape
Kranthi	RARS, Palem	Red	Double	Spiny	Normal	Flat
Haritha (PCS-124)	-do-	Green	Double	Spiny	Normal	Flat
Kiran (PCS-136)	-do-	Red	Double	Non-Spiny	Normal	Flat
PCS-122	-do-	Green	Single	Spiny	Papaya (deep)	Flat
PCS-137	-do-	Green	Double	Spiny	Normal	Flat
DCS-9 (Jyoti)	DOR	Red	Double	Spiny	Normal	Flat
DPC-9	-do-	Green	Zero	Spiny	Normal	Flat
LRES-17	-do-	Green	Triple	Spiny	Normal	Cup
VP-1	SDAU, Gujarat	Green	Triple	Spiny	Normal	Cup

RARS - Regional Agril. Research Station; DOR - Directorate of Oilseeds Research, Hyderabad; SDAU - Sardarkrushinagar Dandiwada Agril. University

Table 2 Segregation pattern in F₂ generation for various characters in castor

Morphological character	Cross	F ₂ phenotype	No. of F ₂ plants with designated phenotype			Ratio	χ^2	Probability (%)				
Red stem colour Vs Green	Stem colour	Kranthi x PCS-122	Red stem	Green stem	Total	3:1	0.037	0.8-0.9				
			412	140	552							
			Jyoti x PCS-122	Red stem	66				262	3:1	0.02	0.8-0.9
			VP-1 x Kiran	Red stem	51				228	3:1	0.841	0.3-0.5
Triple Vs double bloom	Bloom	VP-1 x Kiran	Triple	double	Total	3:1	2.33	0.1-0.2				
			161	67	228							
			LRES-17 x PCS-137	Triple	17				91	3:1	2.09	0.1-0.2
Double Vs single bloom	Bloom	Kranthi x PCS-122	Double	Single	Total	3:1	0.34	0.5-0.7				
			420	132	552							
			Jyoti x PCS-122	Double	64				262	3:1	0.02	0.8-0.9
			Haritha x PCS-122	Double	134				530	3:1	0.04	0.8-0.9
			Double Vs Zero bloom	Bloom	DPC-9 x Haritha				Double	Zero	Total	3:1
97	30	127										
Flat Vs Cup leaf	Leaf shape	VP-1 x Kiran				Double	Cup	Total	3:1	0.584	0.3-0.5	
			166	62	228							
			LRES-17 x PCS-137	Double	26	91	3:1	0.52				0.3-0.5
Spiny Vs Non-spiny capsules	Capsule spinyess	VP-1 x Kiran	No. of F ₂ plants with designated phenotype			1:2:1	0.94	0.5-0.7				
			Spiny	Sparsely spiny	Non-spiny				Total			
			61	116	51				228			
			Normal (shallow) Vs Papaya leaf (deep)	Leaf lacination	Kranthi x PCS-122				Normal	Very shallow	Deep (Papaya)	12:3:1
406	114	32				552						
Jyoti x PCS-122	Normal	53				10	262	12:3:1	2.59	0.2-0.3		

Table 3 Joint segregation in F₂ generation of various characters in castor

Joint segregation of traits	Cross	No. of F ₂ plants with designated phenotype				Ratio	χ^2 L
Triple bloom – cup shape leaf	LRES-17 X PCS-137	Triple bloom & flat leaf	Triple bloom & cup leaf	Double bloom & flat leaf	Double bloom & cup leaf	Total	
	VP-1 x Kiran	57	17	8	9	91	9:3:3:1 4.846
Red stem colour Double bloom	Kranthi X PCS-122	Red stem & double bloom	Red stem & single bloom	Green stem & double bloom	Green stem & single bloom	552	9:3:3:1 2.898
	Jyoti X PCS-122	115	46	51	16	228	9:3:3:1 0.499
Green stem Triple bloom	VP-1 x Kiran	Red stem & triple bloom	Red stem & double bloom	Green stem & triple bloom	Green stem & double bloom	262	9:3:3:1 0.381
	VP-1 x Kiran	129	48	32	19	228	9:3:3:1 1.754
Green stem Deep cup leaf	VP-1 x Kiran	Red stem & flat leaf	Red stem & cup leaf	Green stem & flat leaf	Green stem & cup leaf	228	9:3:3:1 1.122

References

- Anjani, K. 1997. Inheritance of spinyess in castor (*Ricinus communis* L.). *Crop Improvement*, **24** (1): 113-114.
- Anjani, K. 2005. Purple-coloured castor (*Ricinus communis* L.) - A rare multiple resistant morphotype. *Current Science*, **88** (2): 215-216.
- Mather, K. 1951. Measurement of linkage. In: *Heredity*. Methuen and Co. Ltd., London.
- Moshkin, V.A. 1986. Genetics and breeding of castor. In: *Castor*. Oxonian Press Pvt. Ltd., New Delhi: 93-168.
- Panase, V.G. and Sukhatme, P.V. 1989. *Statistical Methods for Agricultural Workers*. Publication and Information Division, Indian Council of Agril. Research, New Delhi.
- Quattrocchio, F., Wing, J. F., Leppen, H.T.C., Mol, J.N.M. and Koes, R.E. 1993. Regulatory genes controlling anthocyanin pigmentation are functionally conserved among plant species and have distinct sets of target genes. *The Plant Cell*, **5**(11): 1497-1512.
- Reddy, A.R. 1996. International Rice Research Institute. In: *Rice Genetics III. Proceedings of the Third International Rice Genetics Symposium*. 16-20 October 2001, Manila, Philippines.
- Solanki, S.S. and Joshi, P. 2001. Inheritance study of some morphological traits in castor (*Ricinus communis* L.). *Indian Journal of Genetics and Plant Breeding*, **61** (2): 136-139.

Heterosis for yield and yield attributing traits in castor, *Ricinus communis* L.

Y.M. Barad, A.R. Pathak and B.N. Patel

Regional Research Station, Anand Agricultural University, Anand-388 110, Gujarat

(Received: August, 2009; Revised: November, 2009; Accepted: December, 2009)

Abstract

Heterobeltiosis and standard heterosis for seed yield and its attributes was studied in 40 hybrids developed through line x tester mating design (5 pistillate lines and 8 pollen parents) in castor. The significant positive heterosis over better parent (BH) and standard heterosis (SH) for seed yield ranged from -49.22% to 141.32% and -55.27% to 17.10%, respectively. The cross combinations ACP 1 x JI-35 (17.10) and JP 65 x VI-9 (13.82) exhibited significant positive standard heterosis over GCH 4 for seed yield/plant. These superior crosses also exhibited significant and positive heterosis for number of capsule/primary raceme, number of effective branches/plant and length of primary raceme. Hybrid vigour of superior crosses may be exploited commercially after verifying the stability performance across the environments over years.

Key words: Heterobeltiosis, standard heterosis, castor

Introduction

The exploitation of heterosis has been an important breeding tool in castor. With the availability of stable pistillate lines in castor, exploitation of heterosis on commercial scale has become feasible and economical (Gopani *et al.*, 1968). After release of first castor hybrid GCH-3, many high yielding hybrids have been released for commercial cultivation. Though per se performance of parental lines provides clues, reliable information on magnitude of heterosis and combining ability of parent for yield and its component characters involved in the inheritance in different characters are of more helpful in selecting appropriate parents and desirable cross combinations for commercial exploitation of hybrid vigour (Dangaria *et al.*, 1987). The present study was, therefore, undertaken to determine the extent of heterobeltiosis and standard heterosis in castor and to identify most heterotic hybrids.

Materials and methods

The experimental material comprised of five pistillate lines viz., VP 1, Geeta, JP 65, SKP 4 and ACP 1 and 8 male parents viz., I 21, EC 38538, JI 35, GC 2, EB 16, VI 9, SKI 283 and SPS 44-1. The parents were crossed in line x

tester mating design and resulting 40 hybrids along with 13 parents and commercial hybrid GCH 4 were evaluated in Randomized Block Design with three replication during *kharif* 2007-08 at Regional Research Station, AAU, Anand. Each entry was planted in a single row of 10 dibbles keeping 90 cm row to row and 60 cm plant to plant distance. Recommended package of practices was followed for raising the normal crop. Observations on five randomly selected competitive plants were recorded for 9 traits (Table 1). However, days to 50% flowering and days to maturity were recorded on plot basis. Each character was analyzed separately using analysis of variance technique suggested by Panse and Sukhatme (1969) and heterosis was calculated in F_1 hybrids over better parent and standard check hybrid GCH 4.

Results and discussion

The analysis of variance for parents and their hybrids for different traits revealed significant differences for all the traits suggesting the presence of considerable genetic variation in respect of various traits studied. Parents vs. hybrids comparisons were significant for all the characters except for 100-seed weight (Table 1). This indicated presence of overall heterosis for all the characters studied.

Earliness in emergence of primary raceme is highly desirable traits for the crop like castor. Hence, the crosses exhibiting heterosis in negative direction are of immense value for earliness. Hybrid VP 1 x GC 2 exhibited significant and maximum negative heterobeltiosis (-23.50) and standard heterosis (-19.90) for days to 50% flowering. For early maturity VP 1 x EB 16 (-14.89) and SKP 4 x GC 2 (-12.57) depicted significant heterobeltiosis and standard heterosis in desirable direction, respectively. VP 1 x EC 38538 exhibited significant and desirable heterobeltiosis (-71.79) and standard heterosis (-57.66) for plant height and for the number of nodes upto primary raceme JP 65 x SKI 283 having high heterotic value in desirable direction for heterobeltiosis (-33.93) and standard heterosis (-21.86) over standard check hybrid GCH 4, respectively.

JP 65 x JI 35 showed significant and maximum positive heterosis over BP (58.74) and SH (43.94) for length of primary raceme. The crosses, VP 1 x SKI 283 and JP 65 x SPS 44-1 depicted significant and highest positive heterosis over BP (72.72) and SH (51.46), respectively for number of capsules on primary raceme. Maximum

heterosis was observed in hybrid ACP 1 x JI 35 over BP (137.30) as well as SH (9.71) for number of effective branches/plant. The crosses, ACP 1 x EB 16 and SKP 4 x I 21 depicted significant and highest positive heterosis over BP (18.14) and SH (21.52) respectively for 100 seed weight, whereas, cross ACP 1 x EC 38538 (9.94) depicted highest heterosis for oil content over SH.

Comparison of top five crosses for yield (ACP 1 x JI 35, ACP 1 x I-21, JP 65 x VI 9, JP 65 x EB-16 and Geeta X

EB-16) based on *per se* performance, heterosis over BP and SH indicated that most superior hybrids were accompanied by significant and positive heterosis over BP and SH for yield attributes like length of primary raceme, number of capsules in primary raceme and number of effective branches/plant advocating that the high heterosis for seed yield resulted through contribution of its component traits (Table 2).

Table 1 Promising crosses for seed yield/plant with heterosis over standard check hybrid (GCH 4) and better parent (BP) in castor

Characters studied	Heterosis over	Range of heterosis		Best heterotic hybrids	No. of hybrids in desired direction
Days to 50% flowering	BP	-23.50	7.81	VP 1 x GC 2	29
	SH	-19.90	8.38	VP 1 x GC 2	21
Days to maturity of primary raceme	BP	-14.89	5.44	VP 1 x EB 16	33
	SH	-5.79	16.48	SKP 4 x GC 2	4
Plant height upto main raceme	BP	-71.79	85.45	VP 1 x EC 38538	21
	SH	-57.66	83.07	VP 1 x EC 38538	12
Nodes upto primary raceme	BP	-33.93	8.16	JP 65 x SKI 283	35
	SH	-21.86	28.83	JP 65 x SKI 283	16
Length of primary raceme	BP	-7.25	61.65	JP 65 x JI 35	27
	SH	-33.31	43.94	JP 65 x JI 35	10
No. of capsules on primary raceme	BP	-59.88	72.72	VP 1 x SKI 283	16
	SH	-57.50	51.46	JP 65 x SPS-44-1	11
No. of effective branches/plant	BP	-59.67	137.30	ACP 1 x JI-35	15
	SH	-73.59	9.71	ACP 1 x JI-35	4
No. of total branches/plant	BP	-55.43	108.77	JP 65 x VI-9	13
	SH	-58.31	16.35	ACP 1 x JI-35	7
100-seed weight	BP	-30.69	18.14	ACP 1 x EB 16	6
	SH	-13.22	21.52	SKP 4 x I 21	21
Oil content	BP	-9.41	12.40	JP 65 x EC 38538	10
	SH	-4.76	9.94	ACP 1 x EC 38538	12
Seed yield/plant (g)	BP	-49.22	141.32	JP 65 x VI-9	18
	SH	-55.27	17.10	ACP 1 x JI-35	2

BP = Better Parent

SH = Standard Hybrid (GCH 4)

On the whole, considerable heterobeltiosis and standard heterosis observed for seed yield and other associated characters suggested the presence of large genetic diversity among the males and the females and also the unidirectional distribution of allelic constitution contributing towards desirable heterosis in the present material. Low magnitude of desirable heterosis and heterobeltiosis was observed for some of the characters viz., oil content (%)

and 100-seed weight (g) indicated the narrow genetic base among the males and the females and also the ambidirectional distribution of allelic constitution contributing towards undesirable heterosis or may be due to mutual cancellation of effects of dominant alleles present in the material. Similar finding were also reported in castor by Chakrabarty (1997); Joshi *et al.* (2001) and Lavanya and Chandramohan (2003).

Heterosis for yield and yield attributing traits in castor

Table 2 Heterobeltiosis and standard heterosis of superior crosses for seed yield and other components in castor

Crosses	Mean seed yield/plant		Days To 50 % flowering	Days to maturity	Plant height (cm)	No of nodes/plant	Length of primary raceme (cm)	No of capsule on primary spike	No of effective branches	Total no of branches	100 seed Weight (gm)	Oil content (%)	Seed yield/plant (gm)
ACP 1 x JI 35	241.38	BP	-10.38**	-9.57**	-8.39**	-14.37**	48.12**	-6.16	137.30**	68.71**	2.40	-0.59	32.43**
		SH	-0.52	-1.11	26.37**	2.53	21.73**	11.95**	9.71**	16.35**	0.61	-0.52	17.10*
ACP 1 x I-21	232.42	BP	-8.96**	-11.60**	-9.44**	-10.05**	41.74**	-8.58	27.70**	33.50**	-0.80	4.11**	27.51**
		SH	1.05	-1.56	24.92**	7.71*	16.98**	9.07	-3.82*	7.32**	15.06**	5.69**	12.79
JP 65 x VI 9	234.54	BP	-17.59**	-11.36**	1.85	-21.79**	23.33**	4.10	130.85**	108.77**	-11.27**	-6.00**	141.32**
		SH	-14.14**	-4.45*	-11.58**	-7.07*	20.54**	8.89**	6.05**	6.59**	-2.36	-2.69*	13.82*
JP 65 x EB 16	222.67	BP	-8.00**	-11.07**	22.48**	-3.57	37.34**	-1.67	120.28**	40.65**	8.10**	-9.41**	129.11**
		SH	-3.66	-1.56	30.23**	14.04**	17.92**	4.17*	1.19	7.32**	18.95**	-4.35**	8.06
Geeta X EB 16	218.16	BP	-6.00*	-1.57	-6.85	-11.31**	39.33**	-8.25	58.52**	50.06**	1.40	-6.18**	38.96**
		SH	-3.38	8.24**	-0.96	4.33	10.69**	-2.81	6.05**	13.91**	3.69	0.93	5.87
S Ed.			3.19	6.11	5.16	0.94	2.89	3.39	0.74	0.76	0.88	0.53	13.13

*, ** Significant at 5 and 1%, respectively

References

Chakrabarty, S.K. 1997. Combining ability and heterosis studies in castor (*Ricinus communis* L. *Journal of Oilseeds Research*, 14 (2) : 182-188.

Dangaria, C.J., Dobarra, K.L., Fatteh, U.G. and Patel, V.J. 1987. Heterosis and combining ability analysis in castor. *Journal of Oilseeds Research*, 4 : 46-53.

Gopani, D.D., Kabaria, M.M. and Patel, R.H. 1968. Studies of hybrid vigour in castor. *Indian Journal of Agricultural Sciences*, 38 : 520-527.

Joshi, H.L., Mehta, D.R. and Jadon, B.S. 2001. Heterosis of yield and yield components in castor hybrids. *Journal of Oilseeds Research*, 18 (2) : 164-169.

Lavanya, C. and Chandramohan, Y. 2003. Combining ability and heterosis for seed yield and yield components in castor, *Ricinus communis* L. *Journal of Oilseeds Research*, 20 (2) : 220-224.

Panse, V.G. and Sukhatme, P.V. 1969. *Statistical Methods for Agricultural Workers*. Indian Council of Agricultural Research, New Delhi.

Studies on combining ability for seed yield and yield components in castor, *Ricinus communis* L.

Y.M. Barad, A.R. Pathak and B.N. Patel

Regional Research Station, Anand Agricultural University, Anand-388 110, Gujarat

(Received: August, 2009; Revised: November, 2009; Accepted: December, 2009)

Abstract

Combining ability for 11 characters was studied using line x tester mating design involving 5 diverse pistillate lines and 8 inbred lines in castor. Combining ability analysis revealed that *gca* variances were significant for days to 50% flowering, plant height, number of nodes up to primary raceme and 100-seed weight, whereas, *sca* variances were significant for all the characters, reiterating the preponderance of non-additive gene action in the inheritance of these characters. The study also revealed that JP 65 and VP 1 among females while JI 35, I 21 and VI 9 among males were superior and consistent general combiners for seed yield and majority its yield contributing characters. VP 1, GC 2 and EB 16 were found to be good general combiners for days to 50% flowering, days to 50% maturity of primary raceme, plant height and number of nodes upto primary raceme. Good general combiners for oil content were VP 1 among females and SKI 283 amongst the males. The *per se* performance of crosses is not correlated with the *sca* effects of the crosses. The study also suggested the involvement of at least one good general combiner as parent result in better yielding cross. A parallel behavior was observed between *per se* performance and general combining ability effects of parents. The best three hybrids recorded maximum significant positive *sca* effects for seed yield were ACP 1 x SPS-44-1, VP 1 x GC 2 and SKP 4 x EC 38538.

Key words: Castor, *Ricinus communis* L., combining ability, *gca*, *sca*

Introduction

With the advancement in biometrical genetics, several techniques are now available which permit quantitative genetic analysis and selection of promising parents and cross combinations for its further exploitation. The parents which produce good progenies on crossing are of immense value to the plant breeder. Combining ability analysis is a powerful tool to discriminate good as well as poor combiners and choose appropriate parental material in breeding programmes. At the same time, it also gives the information about the nature of gene effects involved

in the inheritance of various characters. Combining ability analysis was carried out in the present study to obtain information on selection of better parents and cross combinations for their further use in hybrid breeding programme.

Materials and methods

Five pistillate lines (Female) viz., VP 1, Geeta, JP 65, SKP 4 and ACP 1 were crossed with 8 male pollen parents viz., I 21, EC 38538, JI 35, GC 2, EB 16, VI 9, SKI 283 and SPS 44-1 in a line x tester mating design to obtain 40 hybrids during *kharif* 2006-07. The hybrids (F_1) along with their parents were grown in a Randomized Block Design with three replications during *kharif* 2007-08 at Regional Research Station, Anand Agricultural University, Anand. Each entry was planted in a single row of 10 dibbles keeping row to row 90 cm and plant to plant 60 cm distance. Recommended package of practices was followed for raising the normal crop. Observations on five randomly selected competitive plants were recorded for 9 traits (Table 1). The mean values of these traits were subjected to combining ability analysis as per methods suggested by Kempthorne (1957).

Results and discussion

Variance estimates due to *gca* (average) were significant for days to 50% flowering, plant height, number of nodes up to primary raceme and 100-seed weight indicating that additive gene action may play important role for these characters. While for the rest of the traits it was not significant indicating that additive gene action may not be playing much role and presence of non-additive gene action in the expression of these characters (Table 1). Variance estimates due to *sca* (average) were highly significant for all the characters. Significant estimates of σ^2_{sca} for all the characters suggested the involvement of non-additive gene action for the inheritance of all the traits studied.

Significant estimates of σ^2_{gca} for the characters mentioned above and the highly significant estimates of σ^2_{sca} for all the characters emphasized the importance of additive as well as non-additive gene actions. These results are akin to the reports of Gondaliya *et al.* (2001), Tank *et al.* (2003) and Solanki *et al.* (2004). However, from the σ^2_A/σ^2_D ratio, preponderance of non-additive gene

Studies on combining ability for seed yield and yield components in castor

action was evident for all the characters, except for plant height and 100-seed weight. These results are in conformity with the reports of Ramu *et al.* (2002) and Lavanya and Chandramohan (2003).

The results of GCA effects (Table 2) revealed that none of the parent was found to be good general combiner for all

the traits. However, an overall appraisal of general combining ability effects revealed that JP 65 and VP 1 were found to be a good and consistent general combiner for majority of the traits. Among males, good and consistent general combining ability effect was exhibited by the parents JI 35 and I 21 and VI 9.

Table 1 Analysis of variance and variance estimates for combining ability for seed and its yield in castor

Source	df	Days to 50 % flowering	Days to maturity	Plant height (cm)	No of nodes/plant	Length of primary raceme	No of capsule on primary raceme	No of effective branches	No of total branches	100 seed weight (gm)	Seed yield/plant (gm)	Oil content (%)
Replication	2	3.68	52.11	174.37 **	1.61 *	209.33 **	120.85 **	4.20 **	2.69	3.33	1279.73 **	0.23
Hybrid	39	63.56 **	184.26 **	951.13 **	8.92 **	257.82 **	737.07 **	55.79 **	57.38 **	25.91 **	3290.63 **	10.81 **
Females	4	175.84 **	300.88	1792.99 *	32.58 *	257.81	876.74	68.96	68.92	18.59	5471.29	22.41 *
Males	7	83.29	233.05	2653.64 **	5.85	490.06 *	1290.92	113.38 *	114.94 *	76.62 **	6018.48 *	14.62
F x M	28	42.59 **	155.40 **	405.24 **	6.31 **	199.75 **	578.66 **	39.52 **	41.34 **	14.28 **	2297.15 **	8.20 **
Error	78	4.36	17.43	10.10	0.31	14.35	17.05	0.75	0.94	1.13	244.80	0.34
$\sigma^2 F$		7.17	11.95	74.29	1.34	10.22	35.81	2.84	2.84	0.73	217.20	0.92
$\sigma^2 M$		5.29	14.59	176.24	0.37	31.83	84.91	7.50	7.61	5.03	384.01	0.95
$\sigma^2 gca$		6.45	12.97	113.50	0.97	18.53	54.70	4.63	4.67	2.38	281.36	0.93
$\sigma^2 sca$		12.91	47.09	131.72	1.99	62.40	187.14	12.90	13.50	4.37	679.57	2.59
$\sigma^2 A$		12.89	25.93	227.00	1.94	37.06	109.39	9.27	9.34	4.76	562.71	1.86
$\sigma^2 D$		12.91	47.09	131.72	1.99	62.40	187.14	12.90	13.50	4.37	679.57	2.59
$\sigma^2 A/\sigma^2 D$		0.99	0.55	1.72	0.97	0.59	0.59	0.72	0.69	1.09	0.83	0.72
Degree of dominance		1.00	1.35	0.76	1.01	1.30	1.31	1.18	1.20	0.96	1.10	1.18

Table 2 Estimates of general combining ability (gca) effects for yield and its components

Parents / hybrids	Days to 50 % flowering	Days to maturity	Plant height (cm)	No of nodes/plant	Length of primary raceme	No of capsules on primary raceme	No of effective branches	total no of branches	100 seed weight (gm)	Seed yield/plant (gm)	Oil content (%)
Female parents											
VP 1	-3.87 **	-1.58 **	-8.72 **	-1.53 **	0.08	-7.76 **	0.70 **	0.61 **	-0.70 **	-14.85 **	1.15 **
Geeta	1.84 **	6.05 *	14.12 **	1.56 **	-0.56 **	3.07 **	2.63 **	2.55 **	-1.00 **	6.32	-0.42 **
JP 65	-0.91 **	-2.66 **	0.12	-0.65 **	5.22 **	7.99 **	-0.60 **	-0.19 **	0.48 **	20.01 **	-0.96 **
SKP 4	-0.24 **	-1.99 **	-0.75 **	0.43 **	-3.81 **	-3.56 **	-1.45 **	-1.74 **	1.17 **	3.88	-0.69 **
ACP 1	3.18 **	0.18	-4.77 **	0.19 **	-0.93 **	0.26	-1.27 **	-1.24 **	0.04	-15.36 **	0.92 **
SEd±	0.40	0.77	0.65	0.12	0.72	0.85	0.18	0.19	0.22	3.28	0.13
CD (P=0.05)	0.80	1.53	1.29	0.24	1.44	1.69	0.37	0.38	0.44	6.53	0.13
Male parents											
I-21	1.06 *	-0.24 **	12.34 **	1.01 **	3.49 **	-3.26 **	0.92 **	1.29 **	3.22 **	19.91 **	0.12 *
EC-38538	2.33 **	7.09 **	-23.82 **	-0.30 **	-7.71 **	-8.55 **	-4.33 **	-4.53 **	0.68 **	-36.29 **	0.74 **
JI-35	0.46	-2.78 **	19.98 **	0.70 **	10.05 **	12.92 **	3.19 **	3.07 **	-1.06 **	25.45 **	-1.53 **
GC-2	-4.54 **	-5.78 **	-1.23 **	-0.70 **	2.19 *	-6.04 **	-0.51 **	-0.65 **	-3.07 **	-15.49 **	-0.09 **
EB-16	-0.14 **	-0.44 **	-7.00 **	-0.24 **	-6.91 **	11.90 **	2.10 **	1.97 **	1.44 **	7.72	-1.25 **
VI-9	-1.41 **	-2.51 **	-4.80 **	-0.50 **	-1.65 **	-0.12 **	1.83 **	1.98 **	-3.04 **	4.54	-0.16 **
SKI-283	2.99 **	2.16	-1.96 **	-0.41 **	-0.36 **	4.57 *	-3.84 **	-3.67 **	1.77 **	4.53	1.15 **
SPS-44-1	-0.74 **	2.49	6.49 **	0.45 **	0.90	12.37 **	0.64 **	0.53 **	0.08	-10.37 **	1.00 **
SEd±	0.51	0.97	0.82	0.15	0.92	1.07	0.23	0.24	0.29	4.15	0.17
CD (P=0.05)	1.01	1.93	1.63	0.30	1.82	2.13	0.46	0.48	0.55	8.26	0.34

It was also observed that the *per se* performance for different characters in general agreement with the *gca* effects. However, this can not be taken as a rule because parents or genotypes with high *per se* performance need not always be good general combiners e.g. VP 1, JP 65 and JI 35 was good general combiner for seed yield/plant but these parents were found poor on the basis of *per se* performance. This could be attributed due to the intra

and/or inter-allelic interaction of genes concerned with the character modified by environmental factors.

The perusal of data (Table 3) revealed that only few hybrids exhibited significant and desirable *sca* effects for 1 or 2 traits. Majority of the crosses with significant *sca* effects involved at least one good combiner parent indicating the significance of non-additive gene action.

Table 3 Estimates of specific combining ability (*sca*) effects for yield and its components

Hybrid No.	Hybrids	Days to 50% flowering	Days to maturity	Plant height (cm)	No. of nodes/plant	Length of primary raceme	No. of capsules on primary spike	No. of effective branches	No. of total 100 seed branches	100 seed weight (g)	Seed yield/plant (g)	Oil content (%)
H1	VP1 x I-21	1.73	-5.09*	2.14	-0.73*	-1.11	-6.32*	-1.89**	-1.44**	-1.48*	-12.95	-1.67**
H2	VP1 x EC-38538	2.47*	1.24	-7.04**	-0.68*	0.43	-8.06**	-0.51	-0.15	3.10**	-7.49	-0.44
H3	VP1 x JI-35	4.67**	10.11**	-15.51**	0.26	12.13**	-15.79**	1.77**	2.12**	-0.63	11.28	-0.87*
H4	VP1 x GC-2	-1.67	2.78	17.77**	-0.30	8.60**	17.89**	0.66	0.56	1.94**	46.62**	0.34
H5	VP1 x EB-16	-4.73**	-9.23**	1.74	-0.96**	3.16	3.16	4.80**	3.94**	-2.34**	-1.97	-0.20
H6	VP1 x VI-9	2.87**	4.51*	5.34**	0.72*	0.63	1.31	-3.27**	-3.93**	-2.59**	0.17	0.96*
H7	VP1 x SKI-283	-1.20	-0.16	1.70	0.81*	7.47**	21.29**	1.80**	1.99**	-3.09**	17.84	0.50
H8	VP1 x SPS-44-1	-4.13**	-4.16	-6.15**	0.89**	-7.04**	-13.47**	-3.35**	-3.08**	5.09**	-53.49**	1.35**
H9	Geeta x I-21	3.03**	6.95**	0.70	2.23**	0.20	-2.22	0.24	-0.12	0.86	-1.33	-0.20
H10	Geeta x EC-38538	3.43**	-4.72*	10.58**	-1.26**	-1.87	-4.96*	-2.24**	-2.09**	-1.43*	-33.06**	1.28**
H11	Geeta x JI-35	-1.71	-7.18**	13.85**	0.93**	3.30	13.04**	-2.23**	-2.43**	-0.62	-4.21	-0.25
H12	Geeta x GC-2	-0.71	-6.18**	-4.80*	0.93**	0.30	-8.74**	1.66**	1.49**	-1.42*	9.61	0.16
H13	Geeta x EB-16	-0.12	4.15	11.43**	-0.39	-6.55**	7.40**	2.13**	2.43**	-1.26*	23.74*	1.02**
H14	Geeta x VI-9	-1.52	8.88**	14.36**	-0.93**	-6.67**	-7.49**	-0.20	-0.27	1.38*	-27.15**	0.33
H15	Geeta x SKI-283	1.76	4.22	14.14**	-0.74*	-2.70	-9.51**	-3.20**	-2.75**	0.89	14.53	-1.43**
H16	Geeta x SPS-44-1	-4.18**	-6.12**	19.61**	-0.78*	13.99**	12.46**	3.85**	3.74**	1.60*	17.87	-0.88*
H17	JP 65 x I-21	-1.56	2.33	-5.96**	-0.93**	-6.78**	6.77**	-0.06	0.28	0.41	-14.50	0.29
H18	JP 65 x EC-38538	-3.16**	-4.68*	-8.68**	-1.90**	-13.25**	-11.97**	-2.28**	-2.36**	-3.25**	32.16**	-2.23**
H19	JP 65 x JI-35	-2.96*	0.86	0.39	-0.46	10.46**	0.49	-3.93**	-3.82**	1.32*	-20.65*	1.24**
H20	JP 65 x GC-2	3.04**	5.19*	-3.53	0.48	-2.55	11.11**	-1.50**	-1.51**	-0.50	-0.91	-1.25**
H21	JP 65 x EB-16	1.31	-1.81	21.97**	3.36**	12.81**	10.91**	1.76**	2.14**	2.22**	8.73	-0.09
H22	JP 65 x VI-9	-4.09**	-4.08	-6.22**	0.21	9.02**	-18.34**	2.83**	3.93**	-0.24	12.61	-0.38
H23	JP 65 x SKI-283	2.51*	-1.74	-7.40**	-2.15**	-11.87**	-20.09**	-2.30**	-3.02**	2.40**	-20.77*	2.21**
H24	JP 65 x SPS-44-1	4.91**	3.93	10.42**	1.39**	2.15	21.11**	5.48**	4.91**	-2.36**	3.32	0.21
H25	SKP 4 x I-21	-2.23	0.66	-1.10	-0.84*	-0.35	-9.69**	-3.15**	-4.18**	0.59	-2.87	0.07
H26	SKP 4 x EC-38538	-4.16**	-6.66**	7.46**	2.27**	11.65**	19.84**	3.51**	3.90**	-0.01	33.66**	-1.55**
H27	SKP 4 x JI-35	1.38	-2.14	3.79*	-0.51	-5.77**	6.67**	1.12*	1.07*	0.74	-14.92	-0.28
H28	SKP 4 x GC-2	-0.63	-3.48	-4.40*	-0.67*	-1.08	-6.31*	-0.25	-0.68	-0.19	-28.88**	-0.82*
H29	SKP 4 x EB-16	2.64*	6.86**	-8.16**	-0.12	-6.92**	-16.49**	-5.45**	-5.17**	-0.77	0.22	0.44
H30	SKP 4 x VI-9	-1.43	-2.74	4.38*	-0.99**	4.45*	8.44**	5.08**	4.96**	1.91**	12.30	-0.65
H31	SKP 4 x SKI-283	-4.49**	-7.41**	16.47**	2.21**	4.36*	7.52**	4.75**	5.14**	1.12	17.62	-0.81*
H32	SKP 4 x SPS-44-1	8.91**	14.93**	-18.45**	-1.35**	-6.35**	-9.98**	-5.60**	-5.06**	-3.38**	-17.13	3.59**
H33	ACP 1 x I-21	-0.98	-4.84*	4.23*	0.27	8.04**	11.50**	4.87**	6.00**	-0.38	31.64**	1.51**
H34	ACP 1 x EC-38538	1.43	14.83**	-2.32	1.57**	3.04	5.15*	1.52**	0.70	1.59*	-25.27**	2.94**
H35	ACP 1 x JI-35	-1.38	-1.64	-2.52	-0.21	4.14*	-4.41	3.27**	3.06**	-0.80	28.50**	0.16
H36	ACP 1 x GC-2	-0.04	1.69	-5.04**	-0.44	-5.26*	-13.96**	-0.57	0.14	0.17	-26.44**	1.57**
H37	ACP 1 x EB-16	0.89	0.03	-4.14*	-1.89**	-2.50	-4.99*	-3.24**	-3.35**	2.16**	-30.72**	-1.17**
H38	ACP 1 x VI-9	4.16**	-6.58**	10.87**	0.99**	-7.43**	16.09**	-4.44**	-4.69**	-0.46	2.07	-0.26
H39	ACP 1 x SKI-283	1.43	5.09*	4.36*	-0.13	2.74	0.80	-1.04	-1.37*	-1.32*	-29.22**	-0.47
H40	ACP 1 x SPS-44-1	-5.51**	8.58**	-5.43**	-0.17	-2.76	-10.13**	-0.38	-0.51	-0.95	49.43**	-4.27**
	SEd	1.14	2.17	1.83	0.34	2.05	2.40	0.52	0.53	0.62	9.28	0.38
	CD (P=0.05)	2.26	4.32	3.65	0.67	4.07	4.77	1.04	1.06	1.24	18.48	0.75

The estimates of sca effects revealed that none of the hybrids was consistently superior for all the traits. The best three hybrids on the basis of significant positive sca effects for seed yield/plant were ACP 1 x SPS 44-1, VP 1 x GC 2 and SKP 4 x EC 38538. It also depicted positive sca effects for majority of its yield attributing traits.

A perusal of data showed a good agreement between best general combining parents and best performing parents for most of the traits. This suggested that while selecting the parents for hybridization programme, *per se* performance of parents should be given due consideration. Such parallel behavior of *per se* performance and general combining ability was also reported by Dangaria *et al.* (1987); Pathak *et al.* (1989); Lavanya and Chandramohan (2003) and Thakker *et al.* (2005). The best three hybrids with maximum significant positive sca effects for seed yield/plant viz., ACP 1 x JI35 (poor x good), JP 65 x VI 9 (good x average) and JP 65 x EB 16 (good x average) had also significant desired heterotic response over betters parent and standard check hybrid. High yielding hybrids had high sca effects and high heterosis for most of its yield contributing characters. This appeared appropriate as yield being a complex character depends on a number of its component traits.

References

- Dangaria, C.J., Dobarra, K.L., Fattah, U.G. and Patel, V.J. 1987. Heterosis and combining ability analysis in castor. *Journal of Oilseeds Research*, **4** : 46-53.
- Gondaliya, A.B., Dangaria, C.L., Kavani, R.H. and Golakia, P.R. 2001. Genetic architecture for yield and its components in castor. *Journal of Oilseeds Research*, **18** (2) : 150-153.
- Kemphorne, O. 1957. *An introduction to genetic statistics*. John Willey and Sons. Ind., New York. Pp. 468-470.
- Lavanya, C. and Chandramohan, Y. 2003. Combining ability and heterosis for seed yield and yield components in castor. *Ricinus communis* L. *Journal of Oilseeds Research*, **20** (2) : 220-224.
- Pathak, H.C., Dixit, S.K. and Patel, P.G. 1989. Line x tester analysis for seed yield and its components in castor (*Ricinus communis* L.). *Indian Journal of Genetics*, **49** (1) : 125-129.
- Ramu, R., Sreedhar, N., Lavanya, C. and Ramesh, T. 2002. Combining ability studies in castor (*Ricinus communis* L.). *Journal of Oilseeds Research*, **19** (2) : 229-230.
- Solanki, S.S., Deora, V.S., Singh, D.P. 2004. Combining ability of new castor, *Ricinus communis* L. pistillate line: MCP-1-1. *Journal of Oilseeds Research*, **21**(2) : 274-276.
- Tank, C.J., Jaimini, S.N. and Ravindrababu, Y. 2003. Combining ability analysis over environments in castor. *Crop Research*, **26** (1) : 119-125.
- Thakker, D.A., Jadon, B.S., Patel, K.M. and Patel, C.J. 2005. Heterosis over environments for seed yield and other attributes in castor, *Ricinus communis* L. *Journal of Oilseeds Research*, **22** (2): 324-326.

Combining ability studies for certain quantitative characters in linseed, *Linum usitatissimum* L.

Ram Jeet, P.K. Singh, S.D. Dubey and Amar Singh

Department of Genetics and Plant Breeding, C.S. Azad University of Agriculture and Technology, Kanpur-208 002, UP

(Received: May, 2009; Revised: September, 2009; Accepted: October, 2009)

Abstract

Combining ability for 10 characters was studied using 7 lines viz., BAU-610A, EC-384154 JRF-3, JRF-4, Polf-34, H-25 and H-22 as females and 4 testers Padmini, Parvati, Shubhra and LCK-3003 as male crossed in line x tester mating design. Analysis of combining ability revealed the existence of highly significant variation among crosses for all characters in F_1 and F_2 generations, respectively. The lines BAU-610A was found to be good general combiner for days to maturity, plant height, number of primary branches/plant, 1000-seeds weight, seed yield/plant, oil content, days to 50% flowering and incidence of *Alternaria* blight in F_1 generation. However, BAU-610A had significantly positive *gca* effects for days to 50% flowering and incidence of *Alternaria* blight and poor general combiner in seed yield/plant and oil content in F_2 generations. The variance due to line x tester were significant for all the characters, suggesting the presence of non-additive gene effect for these traits. The crosses BAU 610-A x Shubhra, JRF-3x Padmini, JRF-4xPadmini in F_1 generation and EC-384154 x Shubhra, JRF-3 x Parvati in F_2 generation were the most promising for improvement of seed yield and one or more of its component traits.

Key words: Line x tester, *gca*, *sca*, linseed

Introduction

In any breeding programme the appropriate selection of parents based on their combining ability not only provides necessary information regarding the choice of parents but also simultaneously illustrate the nature and magnitude of gene action involved. The line x tester mating design helps in realising the objective to estimate the combining ability of parents and by there selecting superior parents as well as cross combinations. Accordingly, the present investigation was undertaken to have the knowledge about the yield and other yield attributing characters of nature of combining ability for some newly developed male and female lines of linseed. *Linum usitatissimum* L. and gene action for yield and other yield attributing characters.

Materials and methods

The experimental materials used in present investigation consisted of 28 crosses derived through line x tester design were grown during *rabi* 2004-05 for advancement of generation and promising 14 F_1 s were back crossed to their respective parents for inheritance studies and selfed F_2 seeds were also harvested. The experiment comprising of 67 genotypes (28 F_1 s + 28 F_2 s + 11 parents) was conducted in a Randomized Block Design with three replications during *rabi* 2005-06 at Oilseeds Research Farm, Kalyanpur of Chandra Shekhar Azad University of Agriculture and Technology, Kanpur. These entries were sown in single row of 3m with 30 cm row to row and 5 cm plant to plant spacing. The observations were recorded on 10 characters (Table 1).

Components of variance estimates of phenotypic and genotypic coefficient of variation (PCV and GCV) were evaluated as per Burton (1952). Heritability in the broad sense was calculated following Hanson (1963) and expected genetic advance was estimated as per Johnson *et al.* (1955) and Kempthorne (1957).

Results and discussion

The variances among the lines in respect of their general combining ability were highly significant for all of the characters in F_1 and F_2 generations (Table 1). The differences among the testers arising primarily due to their *gca* were highly significant for the characters in F_1 and F_2 except days to maturity, number of seeds/capsule, 1000-seeds weight and incidence of *Alternaria* blight in F_2 . The variances among the crosses due to line x tester interaction component indicating their *sca* were highly significant for all the characters in F_1 and F_2 except number of seeds/capsule and 1000-seeds weight in F_2 and incidence of *Alternaria* blight in both generations.

Estimates of *gca* effects showed that it was not possible to pick up a good general combiner for all characters because the combining ability effects of parents were not consistent for all the yield components. The lines BAU-610A was found to be good general combiner for days to maturity, plant height, number of primary branches/plant, number of capsules/plant, 1000-seeds weight, seed yield/plant and oil content days to 50%

Part of Ph.D. Thesis submitted to C.S. Azad University of Agriculture and Technology, Kanpur-208 002, UP.

flowering and least incidence of *Alternaria* blight in F₁ generation. However, BAU 610A had significantly positive *gca* effects for days to 50% flowering and incidence of *Alternaria* blight and poor general combiner in seed yield/plant and oil content in F₂ generation. JRF-4 was found to be a good general combiner for plant height, oil content and more incidence of *Alternaria* blight in both F₁ and F₂ generations. H-25 and H-22 were identified as poor general combiner for days to 50% flowering and number of capsules/plant in F₁ and F₂ generations. The lines EC-384154 was a good general combiner for days to 50% flowering and days to maturity in respect of late maturity duration in both F₁ and F₂ generations. Among the testers Parvati proved to be a good general combiner for delayed maturity and tallness in both F₁ and F₂ generations whereas, Padmini was best general combiner for earliness and dwarfness, seed yield, oil content and least incidence of *Alternaria* blight in F₁ generation. Shubhra was identified as average general combiner for days to 50% flowering and number of primary branches/plant and number of capsules/plant in both F₁ and F₂ generations. H-22 was adjudged to be good general combiner for least incidence of *Alternaria* blight whereas, JRF-3 and LCK-3003 for seed yield/plant in both F₁ and F₂ generations.

In the present investigation, the crosses exhibiting higher estimates of desirable *sca* effects common in both the generations were BAU 610A x Parvati for days to 50% flowering and EC-384154 x Padmini for number of capsules/plant. These common crosses over the generations indicate additive x additive type of gene action for the expression of concerned traits.

The cross combinations H-25 x Parvati, BAU 610A x Padmini, H-25 x Shubhra and EC-384154 x Parvati, Polf-34 x Padmini, JRF-4 x LCK-3003 for days to maturity; JRF-4 x Parvati, EC-384154 x Parvati, BAU 610A x LCK-3003 and Polf-34 x Parvati, H-25 x Parvati, H-22 x LCK-3003 for plant height; Polf 34 x Parvati, JRF-4 x Padmini, BAU 610A x LCK-3003 and EC-384154 x Padmini for number of primary branches/plant; H-25 x

LCK-3003, EC 384154 x Padmini, BAU-610A x LCK-3003 and JRF-4 x Parvati, EC384154 x Padmini, EC-384154 x Shubhra for capsules/plant; H-25 x Parvati, BAU 610A x Shubhra, JRF-4 x Shubhra and H-22 x LCK-3003 for number of seeds/capsules; BAU 610A x Shubhra, JRF-3 x Padmini, H-22 x Parvati and EC384154 x Shubhra for 1000-seeds weight; BAU 610A x Shubhra, JRF-3 x Padmini, JRF-4 x Padmini and EC-384154 x Shubhra, JRF-3 x Parvati, H-25 x Parvati for seed yield/plant; Polf-34 x Parvati, H-22 x LCK-3003, BAU 610A x Shubhra and EC-384154 x Padmini, JRF-3 x Parvati, Polf-34 x LCK-3003 for oil content; JRF4 x Shubhra, Polf-34 x LCK-3003, EC-384154 x Padmini and H-22 x Padmini, EC-384154 x Parvati, EC-384154 x Shubhra for incidence of *Alternaria* blight were the good specific combinations for F₁ and F₂ generations, respectively.

In general, maximum crosses showing significant *sca* effects, were invariably associated with better *per se* performance for respective traits (Table 3). The results are in agreement with Patil and Chopda (1983) who concluded that mean performance of the crosses were closely associated with *sca* effects and hence, selection of crosses on the basis of heterotic response should prove effective.

The good specific combiners for different characters involved parents with low x low, low x average, average x average, average x high and high x high general combinations but may also occur in other types of parent combinations. However, in majority of the cases the crosses exhibiting high *sca* effects were found to have both or one of the parents as good general combiners for the characters under reference. Kushalkar *et al.* (2002), Tewari *et al.* (2004) and Ratanparkhi *et al.* (2005) also observed that most cross combinations with high *sca* effects involved at least one high general combining parent. Most interesting crosses are those that involve both parents with high *gca* and posses high *sca* effects. The major parts of such variance would be fixable in later generations.

Table 1 Analysis of variance for combining ability for 10 characters in linseed (mean square)

Sources of variation	d.f.	Gen.	Days to 50% flowering	Days to maturity	Plant height (cm)	No. of Primary branches/plant	No. of capsules/plant	No. of seeds/capsule	1000-seeds weight (g)	Seed yield/plant (g)	Oil content (%)	Incidence of <i>A. blight</i> (0-5 scale)
Females (Lines)	6	F ₁	35.05**	130.14**	231.99**	2.29**	273.15**	0.46**	2.79**	8.56**	3.90**	2.70**
	6	F ₂	235.21**	113.33**	216.72**	7.76**	260.21**	1.26**	0.37**	8.75**	7.01**	2.14**
Males (Testers)	3	F ₁	39.88**	55.65**	1150.73**	0.91**	109.70**	2.17**	2.15**	5.23**	2.56**	1.65*
	3	F ₂	194.91**	6.62	2746.67**	0.78**	42.77**	0.46	0.27	7.20**	2.43**	0.33
Female x male	27	F ₁	14.38**	68.96**	247.39**	1.47**	143.05**	1.18**	2.16**	5.98**	2.84**	1.61*
	27	F ₂	25.21**	24.30**	217.45**	0.67**	111.22**	0.67*	0.28	4.76**	2.14**	0.31
Error	76	F ₁	3.20	5.33	3.15	0.21	21.42	0.12	0.24	0.44	0.18	0.36
	76	F ₂	5.08	5.48	4.24	0.27	9.49	0.37	0.16	0.19	0.13	0.06

* Significant at 5% probability level,

** Significant at 1% probability level

Table 2 Estimates of general combining ability effects for 10 characters in linseed

F₁ Generations

Crosses	Days to 50% flowering	Days to maturity	Plant height (cm)	No. of Primary branches/plant	No. of capsules/plant	No. of seeds/capsule	1000-seeds weight (g)	Seed yield/plant (g)	Oil content (%)	Incidence of <i>A. blight</i> (0-5 scale)
Lines										
BAU-610 A	-1.89**	-2.15**	1.68**	0.35*	8.10**	0.11	0.39**	1.06**	0.40**	-0.57**
EC-384154	1.52**	4.18**	-5.46**	0.01	-0.14	0.11	0.12	0.71**	0.04	-0.02
JRF-3	-0.06	-2.32**	0.50	-0.57**	4.18**	0.11	0.59**	0.43*	0.54**	0.03
JRF-4	1.02*	0-0.82	3.12**	0.10	-4.49**	0.19	-0.17	-0.12	0.42**	0.88**
Polf-34	2.44**	5.118**	7.05**	0.60**	0.51	-0.06	0.28*	0.06	0.08	-0.13
H-25	-1.56**	-2.99**	-3.35**	-0.57**	-2.99*	0.06	-0.48**	-0.79**	-1.06**	0.23
H-22	-1.48**	-1.07	-3.54**	4.10**	-4.90**	-0.39**	-0.73**	-1.35**	-0.43**	-0.42*
SE (sgi) ±	0.52	0.67	0.51	0.13	1.34	0.10	0.14	0.19	0.12	0.17
SE (sgi) ± sgj	0.73	0.94	0.72	0.19	1.89	0.14	0.20	0.27	0.17	0.25
Testers										
Padmini	-1.63**	0.89	-4.78**	0.12	-0.63	0.07	0.10	0.41**	0.49**	-0.36**
Parvati	1.70**	-1.39**	10.87**	0.12	-2.77**	0.07	0.40**	-0.16	-0.10	+0.19*
Shubhra	0.23	-1.35**	-1.51**	0.07	0.75	-0.45**	-0.34**	-0.64**	-0.33**	0.06*
LCK-3003	-0.30	1.85**	-4.59**	-0.31**	2.65*	0.39**	-0.15	0.39**	-0.07	-0.89
SE (sgi) ±	0.39	0.50	0.30	0.10	1.01	0.08	0.11	0.14	0.09	0.13
SE (sgi) ± sgj	0.55	0.71	0.55	0.14	1.43	0.11	0.15	0.20	0.13	0.19

F₂ Generations

Crosses	Days to 50% flowering	Days to maturity	Plant height (cm)	No. of Primary branches/plant	No. of capsules/plant	No. of seeds/capsule	1000-seeds weight (g)	Seed yield/plant(g)	Oil content (%)	Incidence of <i>A. blight</i> (0-5 scale)
Lines										
BAU-610 A	3.07**	0.40	-1.17*	0.43**	0.76	-0.26	0.00	-0.35**	-0.95**	0.26**
EC-384154	5.49**	2.07**	1.09	-0.65**	0.18	-0.26	0.11	-1.29**	-0.37**	0.55**
JRF-3	4.40**	4.74**	-5.33**	-1.07**	-4.24**	-0.01	0.27*	0.96**	0.52**	0.22**
JRF-4	-0.01	-1.93**	4.72**	-0.15	8.93**	-0.18	0.01	-0.85**	0.38**	0.17*
Polf-34	-5.51**	-5.10**	-5.05**	1.43**	1.76*	0.57**	-0.01	0.84**	1.09**	-0.19*
H-25	-3.18**	-0.26	5.44**	-0.15	-4.57**	0.32	-0.12	0.23	0.24*	-0.69**
H-22	-4.26**	0.07	0.30	0.18	-2.82**	-0.18	0.27*	0.45**	-0.91**	-0.33**
SE (sgi) ±	0.65	0.68	0.59	0.15	0.89	0.18	0.12	0.13	0.10	0.07
SE (sgi) ± sgj	0.92	0.96	0.84	0.21	1.26	0.25	0.16	0.18	0.15	0.10
Testers										
Padmini	-4.25**	-0.05	-9.63**	-0.12	-2.10**	0.01	0.15	-0.24*	0.37**	0.04
Parvati	2.04**	-0.38	16.58**	-0.17	0.98	0.20	-0.07	-0.79**	-0.13	0.14*
Shubhra	-0.15	-0.38	-4.07**	0.02	0.29	-0.08	0.03	0.56**	-0.41**	-0.01
LCK-3003	2.34**	0.81	-2.88**	0.26*	0.90	-0.13	-0.11	0.39**	0.16*	-0.16**
SE (sgi) ±	0.49	0.51	0.45	0.11	0.67	0.13	0.09	0.10	0.08	0.06
SE (sgi) ± sgj	0.70	0.72	0.64	0.16	0.95	0.19	0.12	0.13	0.11	0.08

* Significant at 5% probability level,

** Significant at 1% probability level

Table 3 Best specific combinations along with sca performance and gca effects of the parents for different characters in linseed

Characters	Best specific combinations	F ₁ per se performance	Gca effects of the parents	Best specific combinations	F ₂ per se performance	Gca effects of the parents
Days to 50% flowering	JRF-4xPadmini	-4.45**	HxL	JRF-4xLCK-3003	-3.70	AxH
	H-22xLCK-3003	-2.29*	LxA	EC-384154xParvati	-3.54	HxH
	BAU-610AxParvati	-2.20	LxH	BAU-610AxParvati	-2.79*	HxH
Days to maturity	H-25xParvati	-7.77**	LxL	EC-384154xParvati	-5.12**	HxA
	BAU-610xPadmini	-5.56**	HxA	Polf-34xPadmini	-3.29*	HxA
	H-25xShubhra	-5.49**	LxL	JRF-4xLCK-3003	-2.98*	LxA
Plant height (cm)	JRF-4xParvati	-19.32**	HxH	Polf-34xParvati	-16.63**	LxH
	EC-384154xParvati	-10.93**	LxH	H-25xParvati	-11.80**	HxH
	BAU-610AxLCK-3003	-6.56**	HxL	H-22xLCK-3003	-7.08**	AxL
No. of primary branches/plant	Polf-34xParvati	1.88**	AxA			
	JRF-4xPadmini	0.71**	AxA	EC-384154xPadmini	0.74*	LxA
	BAU-610AxLCK-3003	0.56*	HxL			
No. of capsules/plant	H-25xLCK-3003	14.83**	LxH	JRF-4xParvati	7.93**	HxA
	EC-384154xPadmini	13.55**	AxA	EC-384154xPadmini	6.68**	AxL
	BAU-610AxLCK-3003	6.76*	HxH	EC-384154xShubhra	6.63**	AxA
Number of seeds/capsule	H-25xParvati	1.01**	AxA	H-22xLCK-3003	0.80*	AxA
	BAU-610AxShubhra	0.70**	AxL			
	JRF-4xShubhra	0.62**	AxL			
1000-seed weight (g)	BAU-610AxShubhra	1.32**	HxL			
	H-22xParvati	1.20**	LxH	EC-384154xShubhra	0.46*	AxA
	JRF-3xPadmini	0.77**	HxA			
Seed yield/plant (g)	BAU-610AxShubhra	2.27**	HxL	EC-384154xShubhra	1.95**	LxH
	JRF-3xPadmini	2.16**	HxH	JRF-3xParvati	1.50**	HxL
	JRF-4xPadmini	1.89**	AxH	H-25xParvati	1.14**	AxL
Oil content (%)	Polf-34xParvati	1.69**	AxA	EC-384154xPadmini	1.50**	LxH
	H-22xLCK-3003	1.45**	LxA	JRF-4xParvati	1.31**	HxL
	BAU-610AxShubhra	1.36**	HxL	Polf-34xLCK-3003	1.18**	HxH
Incidence of <i>Alternaria blight</i> (0-5 Scale)	JRF-4xShubhra	-1.22**	HxH	H-22xPadmini	-0.39**	LxA
	Polf-34xLCK-3003	-0.91*	LxA	EC-384154xParvati	-0.35*	HxH
	EC-384154xPadmini	-0.82*	AxL	EC-384154xShubhra	-0.33*	HxA

Where H= Good general combiner,

L= Poor general combiner,

A= Average general combiner

The analysis of variance for combining ability revealed highly significant differences among crosses for all the characters in F_1 and F_2 generations. The difference among lines (female) were significant for days to 50% flowering, days to maturity, plant height, number of primary branches/plant, number of seeds/capsule, 1000-seeds weight, seed yield/plant, oil content and incidence of *Alternaria* blight in F_1 and F_2 generations respectively. Whereas differences among testers (male) for all the characters in F_1 and F_2 generations were highly significant except days to maturity, number of seeds/capsule, 1000-seeds weight and incidence of *Alternaria* blight in F_2 generation.

The estimates of σ^2_s were much higher than σ^2_g for all the characters. The estimates of average degree of dominance indicated overdominance for all characters in F_1 and F_2 generations, except days to 50% flowering, number of primary branches/plant and plant height in F_2 generation which showed partial dominance.

On the basis of general combining ability effects, BAU 610-A, JRF-3, EC384154, Padmani and LCK-3003 were identified as most promising parents for seed yield and its contributing traits in F_1 , whereas, H-22 was identified as best general combiner for least incidence of *Alternaria* blight. in both F_1 and F_2 generations.

References

- Burton, G.W. 1952. Quantitative inheritance in grasses. Proc. 6th International Grassland Congress, 1 : 227-283
- Hanson, W.D. 1963. Heritability, Statistical Genetics and Plant Breeding NASNRC. Washington, pp.125 - 140.
- Johnson, H.W., Robinson, H.F. and Comstock, R.E. 1955. Estimates of genetic and environmental variability in soybean. *Agronomy Journal*, 47 :314-316.
- Kempthorne, O. 1957. *An Introduction to Genetical Statistics* (ed.) John Wiley and Sons, Inc., New York, U.S.A. pp. 458-471.
- Kushalkar, A.M., Patil, B.R., Thawari, S.B., and Khatod, J.P. 2002. Combining ability studies in linseed (*Linum usitatissimum* L.). *Annals of Plant Physiology*, 16 (2) : 133-137.
- Patil, V.D. and Chopde, R.R. 1981. Combining ability analysis over environment in diallel crosses on linseed (*Linum usitatissimum* L.). *Theory and Applied Genetics*, 60 (6) : 339-343.
- Ratanparkhi, R.D., Dudhe, M.Y., Gawande, N.D. and Bhongli, S.A. 2005. Combining ability study in linseed through line x tester analysis. *Annals of Plant Physiology*, 19 (1) : 99-102.
- Tewari, Nalini, Dixit, R.K. and Singh, H.C. 2004. Combining ability analysis for seed yield and its components in linseed. *Journal of Oilseeds Research*, 21(2) : 343-345.

Productivity of summer groundnut, *Arachis hypogaea* L. as influenced by cumulative residual effect of crop residue incorporation and nitrogen management practices

C. Radhakumari and D. Srinivasulu Reddy

Department of Agronomy, S.V. Agricultural College, Tirupati-517 502, AP

(Received: May, 2009; Revised: July, 2009; Accepted: August, 2009)

Abstract

Field experiments were conducted in the wetland farm of S.V. Agricultural College (Acharya N.G. Ranga Agricultural University), Andhra Pradesh for two consecutive years 2002-03 and 2003-04 to investigate the cumulative residual effect of incorporated crop residues and different nitrogen management practices applied to preceding lowland rice on the performance of succeeding groundnut and results revealed that cumulative residual effect of incorporation of fieldbean crop residues and supply of 100% N through FYM to rice has resulted in superior performance of groundnut crop compared to any other crop residue incorporation and nitrogen management practices adopted to preceding rice crop. However, similar magnitude of performance of groundnut was noticed with the combination of fieldbean crop residue incorporation and supply of 50% nitrogen each through fertilizer and FYM to preceding rice.

Key words: Groundnut, crop residue incorporation, nitrogen management practices, nematode, groundnut

Introduction

In recent years the emphasis has been shifted from individual crop to cropping system as a whole since the response in component crop of the cropping system are influenced by the preceding crops and the inputs applied to them. Legume crop residues incorporated in to the field after harvesting seed can contribute considerable quantity of nitrogen to succeeding crops (Rekhi and Meelu, 1983). About less than 30% of nitrogen and small fractions of phosphorus and potash in organic manure may become available to immediate crop and rest to subsequent crop (Gaur and Singh, 1982). Conjunctive use of nutrients partly through organics and inorganics to preceding rice exhibited significant residual effect on succeeding groundnut (Thimmegowda and Devakumar, 1994). Rice-groundnut is one of the important cropping system in the southern agroclimatic zone region and maintenance of optimum soil fertility is an important consideration for

obtaining higher and sustainable yield due to large turn over of nutrients in soil plant system. Since, the information on cumulative residual effect of crop residue incorporation and nitrogen management practices on groundnut grown after rice is lacking for southern agroclimatic zone of Andhra Pradesh, the present study was conducted to assess the effectiveness of cumulative residual effect of incorporation of crop residues, farm yard manure and fertilizer on dry matter production, nutrient uptake and pod yield of groundnut.

Materials and methods

Field investigations were conducted during 2002-03 and 2003-04 at wetland farm of S.V. Agricultural College, Tirupati (Andhra Pradesh). Soil analysis for physico-chemical properties was carried out initially, prior to the start of the experiment, by drawing soil samples at random from 0-30 cm depth of the experimental field. The results revealed that experimental field was sandy clay loam in texture, slightly alkaline in reaction, low in organic carbon and available nitrogen (160.8 kg/ha), medium in available phosphorus (25.6 kg/ha) and available potassium (175.4 kg/ha).

The experiment was laid out in a Randomized Block Design with five replications. There were four treatments comprising of preceding crops to rice raised during *kharif* season viz., C₁: Greengram, C₂: Clusterbean, C₃: Fieldbean, C₄: Cowpea whose crop residues (after picking the economic yield up to a common period of time of 75 days) are to be incorporated prior to transplanting of succeeding rice crop. Immediately after the last picking of the economic yield of respective crops up to 75 DAS, plants were uprooted from the entire plot area and weight of the four crop residues was recorded on fresh weight basis. At the time of termination of crops, they were at different post-flowering stages, possessing immature pods, flowers and even flower buds. Since, the crop residues have to be incorporated at a common point of time, all of them were removed without waiting for their full maturity. The crop residues thus obtained were chopped and incorporated in to their respective plots. Samples of all the crop residues were taken plot and replication wise, to estimate the nutrient content (Table 1) before incorporation. N, P and K contents of crop residues were

analysed by standard procedures outlined by Jackson (1973). The varieties of greengram, clusterbean, fieldbean, cowpea were LGG-407, Pusa Navabahar, HA-3 and CO-4, respectively.

Rice crop was raised during rabi season after harvest of preceding crops (raised during *kharif*) in the same layout, by sub-dividing each of the *kharif* treatments into four sub-plots, to which four nitrogen management practices were assigned. Rice was taken up in a Split Plot Design with incorporation of crop residues of four preceding crops to rice as main plot treatments viz., C₁: incorporation of greengram crop residues, C₂: incorporation of clusterbean crop residues, C₃: incorporation of fieldbean crop residues and C₄: incorporation of cowpea crop residues and four nitrogen management practices imposed on rabi rice as sub-plot treatments viz., N₁: No nitrogen, N₂: 100% recommended nitrogen through fertilizer, N₃: 50%

recommended nitrogen through fertilizer + 50% recommended nitrogen through farm yard manure, N₄: 100% recommended nitrogen through farm yard manure. The recommended dose of nutrients was 120 kg N, 80 kg P₂O₅ and 40 kg K₂O/ha. The N content in FYM (Table 1) was taken into consideration and the quantity of FYM required for N₃ and N₄ treatments was calculated and incorporated in to the plots 10 days before transplanting of rice. For the treatments N₂ and N₃, fertilizer nitrogen in the form of urea was applied in three split doses of 50% as basal, 25% at active tillering and 25% at panicle initiation stages. A uniform dose of 80 kg P₂O₅ and 40 kg K₂O/ha was applied basally to all the treatments except to N₁, in the form of single super phosphate and murate of potash, respectively, after duly taking into consideration of phosphorus and potassium content of FYM in the FYM involved treatments. Test variety of rice was NLR 33359.

Table 1 Quantity of crop residues and nutrient content (%) of crop residues and FYM, incorporated before planting of rice

Source	2002-03				2003-04			
	Crop residues incorporated* (kg/ha)	N	P ₂ O ₅	K ₂ O	Crop residues incorporated* (kg/ha)	N	P ₂ O ₅	K ₂ O
FYM	--	0.50	0.20	0.51	--	0.50	0.20	0.51
Greengram residue*	7230	0.81	0.20	0.62	6970	0.83	0.21	0.64
Clusterbean residue*	13820	0.52	0.12	0.49	13100	0.54	0.14	0.51
Fieldbean residue*	16900	0.66	0.15	0.45	17200	0.65	0.16	0.44
Cowpea residue*	15440	0.61	0.14	0.50	15200	0.60	0.15	0.49

Groundnut crop was raised during summer season after the harvest of *rabi* rice in the same undisturbed layout of Split Plot Design, to study the cumulative residual effect of incorporation of crop residues of preceding crops to rice and N management practices imposed on *rabi* rice crop on the performance of succeeding groundnut. The layout of *rabi* rice crop was undisturbed, in which groundnut crop was sown. Each plot of *rabi* rice was ploughed using power tiller without disturbing the layout and plots were levelled individually using spades. No treatments were imposed to groundnut crop and the treatments consisted of the same those of *rabi* rice. Treatments consisted of cumulative residual effect of incorporation of crop residues of four preceding crops to rice as main plot treatments viz., C₁: incorporation of greengram crop residues, C₂: incorporation of clusterbean crop residues, C₃: incorporation of fieldbean crop residues and C₄: incorporation of cowpea crop residues and residual effect of four nitrogen management practices imposed on *rabi* rice as sub-plot treatments viz., N₁: No nitrogen, N₂: 100% recommended nitrogen through fertilizer, N₃: 50% recommended nitrogen through fertilizer + 50% recommended nitrogen through farm yard manure, N₄: 100% recommended nitrogen through farm yard manure. Incorporation. Test variety of groundnut was K-134 a spanish bunch, suitable for cultivation in summer season. The seeds of groundnut were treated with mancozeb (3

g/kg seed) and used for sowing with a spacing of 22.5 x 10 cm between rows and plants. Sowing was done by hand dibbling. Intercultivation was done with star weeder at 20 and 40 days after sowing for controlling weeds in inter row spaces followed by hand hoeing for removal of intra row weeds. Plant samples collected for drymatter estimation at different growth stages of groundnut were oven dried, ground into fine powder and used for nutrient analysis. N, P and K contents of plant samples were analysed by the standard procedure out lined by Jackson (1973). The uptake of N, P and K in Kg/ha at different stages of crop growth was calculated by multiplying the nutrient content with the respective drymatter weights.

Results and discussion

Cumulative residual effect of crop residue incorporation and nitrogen management practices imposed on *rabi* rice has exerted marked influence on the growth (Table 2) and yield (Table 3) of succeeding groundnut. At all the crop growth stages of recording, plants of the tallest stature, highest LAI and drymatter production of groundnut were produced with the incorporation of crop residues of fieldbean (C₃) to preceding rice followed by cowpea, (C₄), clusterbean, (C₂) and greengram (C₁) residues, with significant disparity between any two of them and shortest plants were noticed with the incorporation of crop residues of greengram (C₁) to preceding rice, during both the years

Productivity of summer groundnut as influenced by cumulative residual effect of crop residue incorporation and nitrogen management practices

of study (Table 2). This might be due to substantial amount of residual nutrients left by fieldbean crop residues to extend the favourable carry over effect on succeeding groundnut crop. The tallest plants, highest LAI and drymatter production of groundnut were produced with the supply of 100 % nitrogen through FYM (N₄) to preceding rice, which were comparable with 50% nitrogen each through fertilizer and FYM (N₃), but significantly superior to 100% nitrogen through fertilizer (N₂) and no nitrogen (N₁), which in turn were comparable between them and the shortest plants were noticed with non-supply of nitrogen (N₁) to preceding rice, during both the years of study. This might be due to the residual effect of FYM either alone or in combination with fertilizer nitrogen, which was comparatively higher than that of the exclusive inorganic source of nitrogen applied to preceding rice crop.

The highest number of pods/plant, hundred pod weight, hundred kernel weight, pod yield and haulm yield were produced with the incorporation of crop residues of fieldbean to rice (C₃) followed by cowpea (C₄), clusterbean (C₂) and greengram (C₁), with significant disparity between any two of them and the lowest number of pods/plant 100 pod weight, 100 kernel weight, pod yield and haulm yield was noticed with the incorporation of crop residues of greengram (C₁) to preceding rice, during both the years of study (Table 3). This might be due to residual and cumulative effect with the incorporation of fieldbean crop residues, which was comparatively higher than that of the other crop residue incorporation, with in the crop residues incorporation, differential residual response with different crop residues added can be attributed to their pattern of mineralization and decomposition.

Table 2 Growth of groundnut as influenced by cumulative residual effect of crop residue incorporation and nitrogen management practices to preceding rice

Cropping system, 2003 -2004												
Treatment	Incorporation of crop residues											
	Plant height (cm)				Leaf area index				Dry matter production (kg /ha)			
	30 DAS	60 DAS	90 DAS	Harvest	30 DAS	60 DAS	90 DAS	Harvest	30 DAS	60 DAS	90 DAS	Harvest
C ₁ : incorporation of greengram crop residues	6.5	13.7	23.4	30.6	0.49	1.60	2.72	2.35	489	2170	3888	5195
C ₂ : incorporation of clusterbean crop residues	6.7	15.8	25.6	32.7	0.49	1.64	2.96	2.59	556	2426	4212	5608
C ₃ : incorporation of fieldbean crop residues	7.2	20.3	30.0	36.4	0.60	1.73	3.43	3.06	689	2923	4779	6517
C ₄ : incorporation of cowpea crop residues	6.9	18.0	27.8	34.6	0.54	1.69	3.20	2.82	626	2677	4552	6067
SEm ±	0.05	0.6	0.7	0.65	0.01	0.01	0.06	0.08	18.3	85.0	87.0	123.4
CD (P=0.05)	0.1	1.5	1.8	1.6	0.04	0.03	0.14	0.21	45	208	215	302
Nitrogen management practices												
N ₁ : No nitrogen	6.5	13.7	22.2	27.6	0.44	1.59	2.74	2.29	506	2119	3818	5200
N ₂ : 100% recommended nitrogen through fertilizer	6.6	15.3	23.8	29.2	0.48	1.62	2.87	2.52	545	2371	4086	5582
N ₃ : 50% recommended nitrogen through fertilizer + 50% recommended through farm yard manure	7.0	18.6	29.6	37.9	0.56	1.70	3.29	2.94	634	2730	4635	6119
N ₄ : 100% recommended nitrogen through farm yard manure	7.2	20.3	31.2	39.7	0.59	1.73	3.42	3.07	675	2970	4892	6488
SEm ±	0.07	0.9	1.04	0.92	0.02	0.02	0.08	0.12	26.0	120.2	124.2	174.5
CD (P=0.05)	0.2	1.8	2.1	1.9	0.05	0.04	0.17	0.25	54	248	275	360
Cropping system, 2004-2005												
Incorporation of crop residues												
C ₁ : incorporation of greengram crop residues	6.3	14.9	22.3	30.0	0.46	1.59	2.83	2.26	509	2231	4196	5073
C ₂ : incorporation of clusterbean crop residues	6.7	16.9	24.2	31.6	0.51	1.63	3.16	2.53	574	2447	4479	5501
C ₃ : incorporation of fieldbean crop residues	7.3	20.3	29.0	35.3	0.61	1.71	3.53	3.02	574	2895	4971	6223
C ₄ : incorporation of cowpea crop residues	7.0	18.4	26.4	33.5	0.56	1.68	3.35	2.78	641	2671	4738	5930
SEm ±	0.06	0.6	0.7	0.61	0.02	0.008	0.06	0.09	23.7	80.1	100.5	113.6
CD(P=0.05)	0.1	1.4	1.7	1.5	0.04	0.02	0.17	0.24	58	196	236	278
Nitrogen management practices												
N ₁ : No nitrogen	6.5	14.7	21.7	27.1	0.46	1.59	2.83	2.18	523	2167	4095	5022
N ₂ : 100% recommended nitrogen through fertilizer	6.6	16.2	22.7	28.5	0.49	1.61	3.00	2.45	572	2388	4346	5346
N ₃ : 50% recommended nitrogen through fertilizer + 50% recommended through farm yard manure	7.1	19.2	27.8	36.6	0.57	1.69	3.43	2.88	648	2748	4854	6019
N ₄ : 100% recommended nitrogen through farm yard manure	7.2	20.5	29.6	38.2	0.61	1.72	3.62	3.07	689	2941	5089	6342
SEm ±	0.08	0.8	0.9	0.86	0.02	0.01	0.09	0.14	34.1	113.2	142.1	174.5
CD (P=0.05)	0.2	1.7	2.0	1.8	0.05	0.03	0.2	0.28	70	234	293	360

Table 3 Yield attributes, pod yield and economics of groundnut as influenced by cumulative residual effect of crop residue incorporation and nitrogen management practices to preceding rice

Cropping system, 2003-2004										
Incorporation of crop residues										
Treatments	Number of pods/ plant	100 pod weight (g)	100 kernel weight (g)	Pod yield (kg /ha)	Haulm yield (kg/ ha)	Shelling percentage	Harvest index*	Gross returns (Rs/ ha)	Net returns (Rs/ha-)	Benefit-cost ratio
C ₁ : incorporation of greengram crop residues	10.0	74.1	30.0	1898	3604	72.8	34.23	25034	14384	2.35
C ₂ : incorporation of clusterbean crop residues	10.7	76.6	31.8	2063	3956	72.9	34.08	27215	16565	2.56
C ₃ : incorporation of fieldbean crop residues	12.2	81.4	35.5	2393	4737	73.5	33.39	31583	20933	2.97
C ₄ : incorporation of cowpea crop residues	11.5	79.0	33.7	2228	4388	73.2	33.47	29403	18753	2.76
SEm±	0.24	0.86	75.6	75.6	97.2	0.94	—	887	0842	0.08
CD (P=0.05)	0.6	2.1	185	185	238	NS	---	2172	1864	0.19
Nitrogen management practices										
N ₁ : No nitrogen	9.7	73.9	29.5	1696	3745	72.2	31.19	22429	11779	2.11
N ₂ : 100% recommended nitrogen through fertilizer	10.1	75.6	30.8	1911	3920	72.6	32.84	25242	14592	2.37
N ₃ : 50% recommended nitrogen through fertilizer + 50% recommended through farm yard manure	12.0	80.0	34.7	2379	4384	73.4	35.23	31372	20722	2.95
N ₄ : 100% recommended nitrogen through farm yard manure	12.6	81.7	35.9	2594	4634	74.3	35.92	34192	23542	3.21
SEm±	0.35	1.21	106.9	106.9	137.5	1.17	---	1688	1642	0.15
CD (P=0.05)	0.7	2.5	220	220	284	NS	---	2854	2672	0.27
Cropping system, 2004-2005										
Incorporation of crop residues										
C ₁ : incorporation of greengram crop residues	9.1	72.9	29.1	1743	3395	73.2	33.85	23005	12355	2.16
C ₂ : incorporation of clusterbean crop residues	9.9	75.7	30.9	1938	3762	73.3	33.79	25577	14927	2.40
C ₃ : incorporation of fieldbean crop residues	11.5	80.6	34.6	2328	4540	73.6	33.77	30725	20075	2.88
C ₄ : incorporation of cowpea crop residues	10.6	78.4	32.7	2133	4184	73.5	33.59	28154	17504	2.64
SEm±	0.16	0.78	0.65	78.4	88.2	0.82	---	937	902	0.09
CD (P=0.05)	0.4	1.9	1.6	192	216	NS	---	2294	2014	0.23
Nitrogen management practices										
N ₁ : No nitrogen	8.9	72.9	28.5	1633	3546	72.6	31.52	21590	10940	2.03
N ₂ : 100% recommended nitrogen through fertilizer	9.3	74.7	29.7	1798	3723	73.1	32.76	23753	13103	2.23
N ₃ : 50% recommended nitrogen through fertilizer + 50% recommended through farm yard manure	11.2	79.2	33.7	2243	4187	73.7	34.90	29584	18934	2.78
N ₄ : 100% recommended nitrogen through farm yard manure	11.6	81.0	35.3	2468	4426	74.3	35.82	32533	21883	3.05
SEm±	0.23	1.10	0.92	110.9	124.8	1.02	---	1814	1794	0.18
CD (P=0.05)	0.5	2.3	1.9	229	258	NS	---	3140	2942	0.29

Supply of 100% nitrogen through FYM (N₄) to preceding rice resulted in the production of highest number of pods/plant, 100-pod weight, 100-kernel weight as well as pod yield and haulm yield, which was comparable with 50% nitrogen each through fertilizer and FYM (N₃), but significantly superior to 100% nitrogen through fertilizer (N₂) and no nitrogen (N₁), which were comparable between them and the lowest number of pods/plant, 10 pod weight, 100 kernel weight, pod yield and haulm yield were noticed with non-supply of nitrogen (N₁) to preceding rice, during both the years of study. This might be due to the residual effect of FYM either alone or in combination with fertilizer nitrogen, which was comparatively higher than that of the exclusive inorganic source of nitrogen applied to preceding rice crop. During both the years of study, shelling percentage of groundnut did not show any significant variation due to residual and cumulative effect of either

incorporation of different crop residues or varied nitrogen management practices tried on preceding rice. Harvest index of groundnut was not altered to a statistically noticeable extent either by incorporation of different crop residues or nitrogen management practices to preceding rice. However, the highest value of harvest index was recorded with incorporation of fieldbean crop residues (C₃) in combination with the supply of 100% nitrogen through FYM (N₄) to preceding rice, while it was the lowest with incorporation of greengram crop residues (C₁) without any nitrogen supply (N₁). In the present study, the residual effect of organic source at higher proportion was evident from higher leaf area index, drymatter accrual, number of pods/plant, 100 pod weight, 100 kernel weight, pod and haulm yield. This clearly indicates that organic source at higher proportion can sustain the nutrient status of soil to produce reasonable residual effect. Organic source of

Productivity of summer groundnut as influenced by cumulative residual effect of crop residue incorporation and nitrogen management practices

nitrogen, besides supplying nutrients to the current crop, quite often leave substantial residual effect on succeeding crops in the cropping system. These results are in accordance with the findings of Maskina and Meelu (1984). Significant carry over effect due to substitution of nitrogen with higher proportions of organic sources to rice crop on the succeeding crops was also reported by Thimmegowda and Devakumar (1994) and Paulraj and Velayudham (1995). Residual effect of fertilizer nitrogen applied to rice was not traceable on the succeeding groundnut crop as exhibited in the present study corroborates the findings of Ramaseshaiah *et al.* (1985).

The highest gross returns, net returns and benefit-cost ratio of groundnut were realized with cumulative residual effect of incorporation of fieldbean crop residues (C_3) to rice were due to higher pod and haulm yield realized by this treatment followed by cowpea (C_4) and clusterbean (C_2), with significant disparity between any two of them, while they were found to be the lowest with the residual and cumulative effect of incorporation of greengram crop residues (C_1) to preceding rice, during both the years of study (Table 3). The highest gross returns, net returns and benefit-cost ratio of groundnut were realised with the residual and cumulative effect of 100% nitrogen through FYM (N_4) applied to preceding rice, which were comparable with residual and cumulative effect of 50% nitrogen each fertilizer and FYM (N_3) but significantly superior to 100% nitrogen through fertilizer (N_2) and no nitrogen (N_1), which were comparable between them and the lowest gross returns, net returns and benefit-cost ratio were recorded with the residual and cumulative effect of non-supply of nitrogen (N_1) applied to preceding rice and also since groundnut crop raised as residual crop, the cost of cultivation did not differ among the treatments.

Based on the out come of the investigation, it could be inferred that by raising a reasonably short duration leguminous crop (either a pulse crop or vegetable crop depending up on the farming situation) preceding to rice and incorporation of the crop residues after picking the economic yield and supply of 50% recommended dose of nitrogen each through fertilizer and FYM to rice followed by raising groundnut as residual crop, to utilize the residual fertility was found the best integrated nitrogen management package for rice-groundnut cropping system to achieve higher growth, productivity and economic returns of succeeding groundnut.

References

- Gaur, A.C. and Singh, R. 1982. Integrated nutrient supply system. *Fertiliser News*, 27(2): 87-98.
- Jackson, M.L. 1973. *Soil chemical analysis*. Prentice Hall of India Pvt. Ltd., new Delhi, pp.134-204.
- Maskina, M.S. and Meelu, O.P. 1984. Farm yard manure (FYM) in a rice-wheat rotation. *International Rice Research Newsletter*, 9(5): 27.
- Paulraj, N.J. and Velayudham, K. 1995. Direct and residual effect of Mussori rock phosphate, organic manures and phosphobacteria in rice-blackgram system. *Madras Agricultural Journal*, 82(3): 220-221.
- Ramaseshaiah, K. Yogeswara Rao, Y. and Bheemaiah, G. 1985. Fertilizer requirements of groundnut grown in sequence with rice. *The Indian Journal of Agricultural Sciences*, 55(1): 25-27.
- Rekhi, R.S. and Meelu, O.P. 1983. Effect of complimentary use of mung straw and inorganic fertilizer nitrogen on the nitrogen availability and yield of rice. *Oryza*, 20: 125-129.
- Thimmegowda, S. and Devakumar, 1994. Effect of residual fertility on yield of groundnut grown in rice fallows. *Indian Agriculturist*, 38: 287-290.

Yield attributes, yield, quality and uptake of nutrients by summer groundnut, *Arachis hypogaea* L. as influenced by sources and levels of sulphur under varying irrigation schedules

G.N. Patel, P.T. Patel, P.H. Patel, D.M. Patel, D.K. Patel and R.M. Patel

C.P. College of Agriculture, Sardarkrushinagar Dantiwada Agril. University, Sardarkrushinagar-385 506, Dantiwada, Gujarat

(Received: June, 2008; Revised: November, 2009; Accepted: December, 2009)

Abstract

The response of summer groundnut Cv. GG-2 to sources and levels of sulphur under varying irrigation schedules was studied on loamy sand soils at Main Castor-Mustard Research Station, S.D.A.U., Sardarkrushinagar during 2003 and 2004. Pooled data indicated that irrigation scheduled at 40 mm CPE significantly improved the yield attributes, pod and haulm yield, quality of groundnut and uptake of nutrients (NPS). Maximum WUE was achieved with irrigation schedule of 50 mm CPE. Sources of sulphur had non-significant effect on yield, quality and uptake of nutrients (NPS) by groundnut.

Key words: Groundnut, irrigation schedules, yield attributes, quality, sulphur

Introduction

In Gujarat, groundnut is largely grown under rainfed condition and yield is influenced due to erratic monsoon, the scope of increasing area under *kharif* groundnut is limited. Due to favorable climatic condition in North Gujarat, summer groundnut in the loamy sand soil provides assured production than the rainy season crop. There is a vast scope to expand groundnut area during summer season in North Gujarat with the availability of Narmada irrigation water in near future. However, the yield potential of this crop grown during summer can be realized to a greater extent by adopting suitable water and nutrient management practices (Gosh *et al.*, 2001). Irrespective of crops, sulphur is now rightly called the fourth major plant nutrient, next only to N, P, and K but for oilseed crops it is as important as phosphorus. Patel *et al.* (1986) while reporting the available S status of the soils in Gujarat indicated the average per cent deficiency of 38.3 in South Gujarat, 42.0 in middle Gujarat, 44.3 in North Gujarat and 25.0 in Saurashtra and Kachchh. Summer groundnut is likely to find the place in the cropping system in light texture soils under assured irrigation facilities in the arid and semi-arid parts of Gujarat. Lack of information on sulphur and water management in summer groundnut led to plan this investigation.

Materials and methods

A field experiment was conducted at Main Castor-Mustard Research Station, Sardarkrushinagar Dantiwada Agricultural University, Sardarkrushinagar, Gujarat during summer seasons of 2003 and 2004. The soil of the experimental plot was foamy sand and slightly alkaline in reaction (pH 7.9), having low organic carbon, available N, high in available phosphorus, medium in available potassium, and low in sulphur (7 ppm). The experiment comprised of three main plot treatments of irrigation schedules viz., I₁: 40 mm CPE, I₂: 50 mm CPE and I₃: 60 mm CPE and seven sub-plot treatments in combination of sources (C₁: elemental sulphur and C₂: gypsum) and levels of sulphur (0, 20, 40 and 60 kg S/ha) was laid out in Split Plot Design and replicated four times. Three common irrigations were followed for proper germination and establishment of the plants. The seeds were drilled at 30cm apart during first half of February. Common dose of 25 kg N and 50 kg P₂O₅/ha was applied in the form of DAP and urea as basal. Elemental sulphur and gypsum were applied 21 days prior to sowing as per treatment as basal. Oil content in the kernel was estimated by Nuclear Magnetic Resonance Spectrophotometer (NMRS). Whereas N, P and S content from kernels, haulms and shells were determined employing standard methods.

Results and discussion

Irrigations scheduled at 40 mm CPE (I₁), showed its superiority for improving number of pods/plant, pod weight/plant, shelling percentage and 100 kernels weight as well as, pod and haulm yield of groundnut (Table 1). Application of frequent irrigations (40 mm CPE, I₁) led to higher pod yield due to improvement in yield attributes viz., number and weight of pods, kernel weight and shelling percentage. Moreover, accumulation of more photosynthates has helped better pod filling in summer groundnut. The results corroborate with the findings of Shaikh *et al.* (2004) and Raskher and Bhoi (2003) indicating that scheduling of irrigation at 75mm CPE improved yield parameters as compared to that of 100 and 125 mm CPE.

Yield attributes, yield, quality and uptake of nutrients by summer groundnut as influenced by sources and levels of sulphur under varying irrigation schedules

The sources of sulphur had no significant effect on the yield attributes as well as on pod and haulm yield of groundnut (Table 1). This is ascribed to similar response of both sources on growth and yield parameters of groundnut. Similar findings have been reported by Kumar *et al.* (2001) for sources of sulphur (gypsum, elemental sulphur and pyrite) in mustard.

Both levels of sulphur (40 and 60 kg) did not vary from each other in respect to production of number of pods, 100 kernel weight and weight of pods per plant showed superiority over lower levels. However, application of S irrespective of dose significantly increased shelling percentage over no sulphur application. A linear increase in pod and haulm yield was observed up to 40 kg S/ha. It increased pod yield over 0 and 20 kg/ha to the tune of 19.90 and 8.34% whereas for haulm yield the corresponding values were 13.49 and 6.45%, respectively. The increase in pod and haulm yield is mainly ascribed to improvement in yield attributes with S application in addition to its multiple role in metabolism as an essential constituent of amino acids and also improvement in vegetative structure, production of reproductive structures and assimilates thereby maintaining balance source sink. These results are in close conformity with the findings Nayak *et al.* (2004). The interactive effect of irrigation schedules and sulphur levels was found significant on pod yield. The treatment combination I₁S₂ standing at par with I₁S₃ (40mm CPE and 40 kg S/ha) registered significantly higher pod yield than rest of the treatment combinations. This indicated that supply of more than 40 kg did not respond under adequate irrigations. It sufficed the need of summer groundnut on S deficient soils.

Quality: Irrigation schedules had no significant effect on oil content of groundnut (Table 1). However, the oil yield

was positively improved with scheduling of irrigations is attributed to increased pod yield and 100 kernel weight. These results are in line with the findings reported by Dodia (1985). The protein content and protein yield were also significantly influenced due to irrigation schedules. The highest protein content was noticed with lower rate of irrigation (I₃) and the reverse was found true for frequent irrigations (I₁). This is mainly due to increase in nitrogen content in kernels produced with lower rate of irrigation (I₃).

Application of sulphur brought significant improvement in quality of groundnut. Significantly higher oil content and oil yield were found with 60 kg S/ha than lower levels of S but it was at par with 40 kg S/ha in case of oil yield. The increase in oil content might be due to S application as it is required for synthesis of S containing amino acids, protein and oils. The highest oil yield was recorded with higher rate of sulphur (60 kg/ha) is attributed to high oil content and pod yield (Table 1) by respective treatment.

The significant positive response of each level of sulphur was observed on protein content. Similar trend as that of oil yield was observed for protein yield with application of sulphur (Table 1). This is contributed to increase in protein content and pod yield (Table 1) as well as nitrogen uptake by groundnut crop (Table 2) with S application, as nitrogen is an integral part of chlorophyll and also a major content of protein. The interaction effect of irrigation schedules and sulphur levels was found significant in case of oil and protein yield. The treatment combination I₁S₂ (40 mm CPE and 40 kg S/ha) was found at par with I₁S₂ (40 mm CPE and 60 kg S/ha) and produced significantly higher oil as well as protein yield than rest of the treatment combinations.

Table 1 Effect of irrigation schedules, sources and levels of sulphur on yield attributes, yield and quality of groundnut (pooled)

Treatments	Pods/plant	Shelling percentage	100 kernel weight (g)	Weight of pods/plant (g)	Pod yield (kg/ha)	Haulm yield (kg/ha)	Oil content (%)	Oil yield (kg/ha)	Protein content (%)
Irrigation schedules									
I ₁ : 40mm CPE	32	75.48	37.54	23.5	3784	6040	48	1363	24
I ₂ : 50mm CPE	31	75.15	36.38	20.9	3475	5546	48	1263	24
I ₃ : 60mm CPE	30	74.79	35.00	18.9	3156	5187	49	1144	24
CD (P=0.05)	0.7	0.24	0.46	0.6	234	290	NS	19	0.06
Sulphur sources									
C ₁ : Elemental S	31	75.21	36.29	21.0	3462	5598	48	1249	24
C ₂ : Gypsum	31	75.07	36.33	21.1	3482	5583	49	1265	24
CD (P=0.05)	NS	NS	NS	NS	NS	NS	NS	NS	NS
Sulphur levels									
S ₀ : Control	27	74.29	33.05	17.2	3085	5161	47	1051	23
S ₁ : 20 kg/ha	31	75.29	35.98	20.9	3414	5502	48	1238	24
S ₂ : 40 kg/ha	33	75.57	38.17	23.1	3699	5857	49	1366	24
S ₃ : 60 kg/ha	33	75.43	38.04	23.1	3688	5843	49	1371	24
CD (P=0.05)	3.2	0.42	0.49	1.3	89	117	0.3	122	0.01

Table 2 Effect of irrigation schedules, sources and levels of sulphur on nutrient uptake, water use, water use efficiency (WUE) and economics of groundnut (pooled)

Treatments	Nutrient uptake (kg/ha)			WUE (kg/ha/mm)	Net return (Rs/ha)	Net ICBR
	N	P	S			
Irrigation schedules						
I ₁ : 40mm CPE	194	28	18	4.8	50282	1.5.56
I ₂ : 50mm CPE	181	27	17	4.7	47084	1:7.37
I ₃ : 60mm CPE	168	25	16	4.4	40687	-
CD (P=0.05)	9.9	1.4	0.8	-	-	-
Sulphur sources						
C ₁ : Elemental S	181	27	17	4.6	45174	-
C ₂ : Gypsum	181	27	17	4.6	45738	-
CD (P=0.05)	NS	NS	NS	-	-	-
Sulphur levels						
S ₀ : Control	155	23	14	4.0	38875	-
S ₁ : 20 kg/ha	177	26	16	4.6	44534	1.29.78
S ₂ : 40 kg/ha	195	29	18	4.9	49522	1:40.55
S ₃ : 60 kg/ha	197	29	19	4.9	49181	1:-2.38
CD (P=0.05)	15.1	1.7	1.4	-	-	-

Table 3 Interaction effect of irrigation schedules and sulphur levels on pod, oil and protein yield (pooled)

Treatments	Pod yield (kg/ha)				Oil yield (kg/ha)				Protein yield (kg/ha)			
	S ₀	S ₁	S ₂	S ₃	S ₀	S ₁	S ₂	S ₃	S ₀	S ₁	S ₂	S ₃
I ₁ : 40mm CPE	3339	3697	4073	4026	1094	1344	1512	1503	563	650	737	734
I ₂ : 50mm CPE	3082	3386	3717	3716	1069	1228	1372	1383	525	606	675	681
I ₃ : 60mm CPE	2834	3159	3308	3323	992	1143	1213	1228	493	565	603	606
CD (P=0.05)	80				74				50			

Irrigation schedule I₁ (40mm CPE) recorded significantly the highest value of N, P and S uptake by groundnut (Table 2). This is ascribed to higher yield (Table 1) with frequent irrigations as well as improvement of nutrient availability under conducive soil environment. Similar findings have been reported by Bharambe *et al.* (2004). With respect to removal of nitrogen, phosphorus and sulphur by groundnut, none of the sources of sulphur had pronounced effect. This is mainly due to even influence of sources on the concentration of N, P and S in kernels, haulm and shell as well as on pod and haulm yield. Successive increase in sulphur application up to 40 kg/ha realized remarkable improvement in N, P, and S uptake by the crop. This could be ascribed to higher nitrogen, phosphorus and sulphur content in kernels, haulm and shell, besides higher pod and haulm yield with higher level of sulphur. The results are in accordance with those reported by Nayak *et al.*

(2004). WUE was increased with decrease in irrigation schedules. The highest WUE (4.76 kg/ha.mm) was achieved with 40mm CPE (I₁). Similarly the highest WUE (4.92 kg/ha mm) being with 60 kg S/ha, was almost equal as that with 40 kg S/ha (Table 2). Net realization accrued from I₁ (40mm CPE) was much higher than 50 (I₂) and 60mm CPE (I₃). However, net ICBR value was highest with 50mm CPE. Similarly higher net return and net ICBR values were obtained with 40 kg S/ha. Application of sulphur through gypsum was found more advantageous. The findings of the investigation indicated that irrigations scheduled at 40 mm CPE (17 irrigations) with 50mm depth of water in summer groundnut supplying 40 kg S/ha through gypsum or elemental sulphur was found more effective and economically viable in loamy sand soils of semi-arid parts of North Gujarat region.

Yield attributes, yield, quality and uptake of nutrients by summer groundnut as influenced by sources and levels of sulphur under varying irrigation schedules

References

- Bharambe, P.R., Patil, V.V., Shelke, D.K., Oza, S.A. and Sondge, V.D. 2004.** Response of rabi groundnut to phosphorus levels under different land layouts and moisture regimes grown on Vertisol. *Journal of Indian Society of Soil Science*, **52** (3) : 262-265.
- Dodia, I.N. 1985.** Response of summer groundnut (*Arachis hypogaea* L.) to irrigation based on IW/CPE ratio. M.Sc. (Agri.) Thesis, Submitted to Gujarat Agricultural University, Sardarkrushinagar.
- Gosh, P.K., Bandopadhyay A., Nautiyal, P.C. and Mathur, P.K. 2001.** Technologies for rabi and summer groundnut cultivation. 12 pp. Technology bulletin, NRCG (ICAR), Junagadh, Gujarat.
- Kumar, S., Singh, B. and Rajput, A.L. 2001.** Response of Indian mustard (*Brassica juncea* L.) to source and levels of sulphur. *Indian Journal of Agronomy*, **46** (3) : 528-532.
- Nayak, S.C., Sahu, S.K., Sarangi, D. and Pradhan, K.C. 2004.** Evaluation of efficacy of source and levels of sulphur for groundnut in lateritic soil. *International Arachis Newsletter*, **24** : 48-49.
- Patel, M.S., Gundaliya, J.D. and Golakiya, B.A. 1986.** Sulphur status in Gujarat soils in relation to oilseeds and pulses. Paper presented in 24th meeting of Agronomy and Soil Science sub- committee of Research Council held at Navsari Campus of Gujarat Agricultural University, May 21-23.
- Raskher, B.S. and Bhoi, P.G. 2003.** Response of summer groundnut (*Arachis hypogaea* L.) to irrigation regimes and mulching. *Indian Journal of Agronomy*, **48** (3) : 210-213.
- Shaikh, A.A., Nimbalkar, C.A. and Jawale, S.M. 2004.** Effect of irrigation scheduling and mulching on yield and yield contributing characters of summer groundnut. *Journal of Maharashtra Agricultural University*, **29**(2): 163-165

Effect of spacing and nitrogen levels on *rabi* castor, *Ricinus communis* Linn. grown under different cropping sequences in North Gujarat agro-climatic conditions

R.M. Patel, M.M. Patel and G.N. Patel

Department of Agronomy, C.P. College of Agriculture, S.D. Agricultural University, Sardarkrushinagar-385 006, Gujarat

(Received: July, 2008; Revised: May, 2009; Accepted: August, 2009)

Abstract

Studies on the effect of spacing and nitrogen levels on *rabi* castor grown under different *rabi* castor based cropping sequences were conducted during 2003-04 and 2004-05. The results revealed that among different *kharif* crops, clusterbean grown as green manure recorded remarkably higher seed yield, stalk yield, net realization and BCR as compared to rest of the *kharif* crops. In case of spacings, 90 cm x 45 cm recorded significantly higher seed and stalk yields than 90 cm x 60 cm. Net realization was not affected due to different spacings. Application of 120 kg N/ha produced remarkably higher seed yield over 40 and 80 kg N/ha. Among different sequences, greengram - castor gave the highest net realization, BCR, system productivity and system profitability.

Key words: *Rabi* castor, system productivity, system profitability, cropping sequence

Introduction

Castor (*Ricinus communis* L.) is the most important non-edible oilseed crop. In Gujarat, it is grown over an area of 3.58 lakh ha producing about 7.08 lakh tonnes with an average productivity of 1978 kg/ha (2007-08). There is a greater scope of growing castor in *rabi* season. Therefore, there are possibilities to take any short duration crop in *kharif* season before growing of *rabi* castor. Development of dairy industries in North Gujarat has created the demand for forage to nourish milch animals. Double cropping system involving cultivation of *rabi* castor after suitable *kharif* crops can solve the food and fodder requirements of farmers and increase the overall income. Fodder sorghum and greengram are important short duration *kharif* crops of the region can successfully be cultivated before *rabi* castor. The beneficial effect of green manure to improve the yield of succeeding crop and soil nutrient status of the soil is well known (Rajaram *et al.*, 2003).

Materials and methods

A field experiments were conducted at the Agronomy Instructional Farm, C.P. College of Agriculture, S.D. Agricultural University, Sardarkrushinagar on the most

profitable *rabi* castor based cropping system and to study the effect of *kharif* crops on spacing and nitrogen requirement of *rabi* castor during *kharif* and *rabi* seasons 2003-04 and 2004-05. The soil of the experimental sites was loamy sand in texture having pH of 7.6, organic carbon 0.17, available nitrogen 179 kg/ha, 40 kg/ha available phosphorus and 184 kg/ha available potash. The experiment was laid out in Split Plot Design with four replications keeping *kharif* crops viz., clusterbean as green manure, greengram, fodder sorghum and fallow in main plots and combinations of spacings (90 cm x 60 cm and 90 cm x 60 cm) and nitrogen levels (40, 80 and 120 kg N/ha) in subplots. The *kharif* crops were grown at the onset of monsoon and castor was sown on 7th and 9th, October of 2003 and 2004, respectively. All the *kharif* crops were raised with recommended package of practices. All the plots were uniformly fertilized with 60 kg P₂O₅/ha as basal at the time of sowing of castor, while half dose of nitrogen was applied as basal and remaining half dose was applied in two equal splits at 30-33 and 80-85 days after sowing (DAS). The rainfall received during 2003-04 and 2004-05 was 691.8 and 250.6 mm with 31 and 20 rainy days, respectively.

Results and discussion

Kharif crops

Clusterbean grown as green manure added an average of 20.9 t/ha green biomass in the soil (Table 1). Greengram produced on an average of 1274 and 1715 kg/ha of seed and fodder while, fodder sorghum gave 24.95 and 7.85 tonnes/ha of green and dry fodder, respectively.

Rabi castor

Effect of *kharif* crops: The pooled results revealed that different *kharif* crops influenced the seed and stalk yields of castor significantly. Higher seed and stalk yields of castor were produced when it was grown after green manure than grown after greengram, fodder sorghum or on fallow land. The increase in seed yield of castor grown after green manure was to the tune of 14.1, 30.7 and 20.1% over greengram, fodder sorghum and fallow, respectively. This could be attributed to the favourable condition prevailing for growth and development of castor grown after green manuring, which improves the physical,

chemical and biological environments of soil. Fodder sorghum being an exhaustive crop, produces huge quantity of fodder and thereby removes large amount of nutrients from the soil, thus resulted in poor growth and development of succeeding crop grown after fodder sorghum as compared to grown after green manure, greengram and fallow. These results are in agreement with the finds of Kumpawat and Rathore (2003) and Mishra and Giri (2004).

Effect of spacing: The seed and stalk yields were remarkably increased with the decrease in spacing. The magnitude of increase in seed yield of castor under 90 cm x 45 cm spacing was to the tune of 4.1% over 90 cm x 60 cm spacing. The higher seed and stalk yields recorded with 90 cm x 45 cm spacing was mainly attributed to the fact that this spacing acquired 33.3% higher plant population than 90 cm x 60 cm spacing.

Effect of nitrogen: The different doses of nitrogen influenced the seed and stalk yields of castor significantly. Fertilizing castor with 120 kg N/ha recorded significantly higher seed and stalk yields over 40 and 80 kg N/ha (Table 2). The probable reason for such a positive response due to addition of higher rate of nitrogen might

be tended to put more vegetative growth, better root development and resulted in efficient photosynthesis and finally produced more seed yield. The maximum mean net monetary realization was registered with greengram cultivation over fodder sorghum, however monetary loss was recorded with green manure practice. In case of BCR, the highest value (3.16) was observed under greengram followed by fodder sorghum (1.06). Significantly the highest net realization with BCR (2.37) were registered when castor was grown after green manure. While, minimum net realization and BCR (1.58) was recorded after fodder sorghum. Net realization received from castor was not influenced significantly due to spacings, while the net realization secured from castor was significantly dose depended.

The castor equivalent yield, net realization, system productivity and system profitability obtained from different crop sequences indicated that the highest castor equivalent yield, total net realization, system productivity, system profitability and BCR (2.35) were received under green gram-castor sequence while, fallow-castor sequence recorded the minimum, net realization, system productivity and system profitability (Table 3).

Table 1 Grain/fodder/stover yield and economics of *kharif* crops (pooled)

Crops	Yield (kg/ha)		Gross realization (Rs/ha)	Cost of cultivation (Rs/ha)	Net realization (Rs/ha)	BC ratio
	Seed	Green biomass/stover/dry fodder				
Green manure	--	20900	--	5,898	(-)5,898	-
Greengram	1,274	1,795	28,173	6,768	21,405	3.16
Fodder sorghum	--	7,850	15,700	7,615	8,085	1.06
Fallow	--	--	--	--	--	--

Table 2 Seed and stalk yield and economics of castor (pooled)

Treatment	Yield (kg/ha)		Gross realization (Rs/ha)	Cost of cultivation (Rs/ha)	Net realization (Rs/ha)	BC ratio
	Seed	Stalk				
Main plot treatments - <i>Kharif</i> crops						
Green manure	3,185	4,340	45,677	13,573	32,104	2.37
Greengram	2,794	3,848	40,080	13,573	26,507	1.95
Fodder sorghum	2,436	3,307	35,017	13,573	21,444	1.58
Fallow	2,651	3,513	37,997	13,573	24,424	1.80
SEm ±	47	72	-	-	658	-
CD (P=0.05)	141	214	-	-	1,955	-
Sub-plot treatments: Spacing x Nitrogen - Spacings						
90 cm x 60 cm	2,711	3,654	38,871	12,986	25,885	1.99
90 cm x 45 cm	2,825	3,850	40,512	14,159	26,353	1.86
SEm ±	28	39	-	-	387	-
CD (P=0.05)	77	108	-	-	NS	-
Nitrogen levels : (kg/ha)						
40	2,434	3,354	34,914	13,060	21,854	1.67
80	2,841	3,824	40,928	13,581	27,347	2.01
120	3,015	4,078	43,232	14,077	29,155	2.07
SEm ±	34	48	-	-	475	-
CD (P=0.05)	95	132	-	-	1,329	-

Table 3 Economics, system productivity and system profitability of different crop sequences (pooled of two years)

Crop sequence	Castor EY (kg/ha)	Gross realization (Rs/ha)	Cost of cultivation (Rs/ha)	Net realization (Rs/ha)	System productivity (kg/ha/day)	System profitability (Rs/ha/day)	BC ratio
Green manure-castor	3262	45,673	19,471	26,202	8.94	71.79	1.34
Greengram -castor	4875	68,253	20,341	47,912	13.36	131.27	2.35
Fodder sorghum - castor	3622	50,712	21,187	29,525	9.92	80.89	1.40
Fallow- castor	2714	37,997	13,573	24,425	7.44	66.92	1.80

References

- Mishra, Bhartendu and Gajendra Giri. 2004. Influence of preceding season practices and direct application of fertilizers on growth, yield, oil content and oil production of Indian mustard (*Brasica juncea*). *Indian Journal of Agronomy*, 49 (4): 264-267.
- Kumpawat, B.S. and Rathore, S.S. 2003. Effect of preceding grain legumes on growth, yield, nutrient content and uptake by wheat (*T. aestivum*) under different nitrogen levels. *Crop Research*, 25 (2): 209-14.
- Rajaram, Sah, R.K., Rai and Mukherjee, P.K. 2003. Effect of green-manuring on available N and P status of the soil after wheat. *Annals of Agricultural Research New series*, 22 (3): 414-16.

Effect of potassium fertilization on cationic nutrient content and uptake in seeds of two castor, *Ricinus communis* L. cultivars

I.Y.L.N. Murthy and P. Padmavathi

Directorate of Oilseeds Research, Rajendranagar, Hyderabad-500 030, AP

(Received: December, 2008; Revised: August, 2009; Accepted: September, 2009)

Abstract

Field experiments were conducted with castor variety DCS-9 and hybrid DCH-32 during the *kharif* season of 2004, in red sandy loam (Alfisol) soil having initial available potassium 202 kg K₂O/ha, under rainfed conditions. Potash treatments comprised 7 levels viz., 0, 20, 40, 60, 80, 100 and 120 kg K₂O/ha. Results showed that castor seed yield responded positively to the potassium (K) doses and optimum doses of K derived with linear response plateau equation were 87 ($r^2 = 0.98$) for DCS-9 variety and 72 ($r^2 = 0.79$) kg K₂O/ha for DCH-32 hybrid. Cationic nutrient content and uptake in the seeds followed the order K > Mg > Ca > Fe > Zn > Cu > Mn irrespective of cultivar. Further, the K fertilization has influenced the Mg, Fe, Mn and Cu uptake in DCS-9 seed while Mg and Cu uptake in DCH-32 seed. The boundary line technique showed the internal nutrient requirement ratios, Ca: K, Mg: K, Fe: K, Zn: K, Mn: K and Cu: K associated with 94% of relative seed yield to be 0.049, 0.495, 0.050, 0.0057, 0.0024 and 0.0034 for DCS-9 and 0.084, 0.613, 0.072, 0.0094, 0.0030 and 0.0061 for DCH-32 respectively. The significance of these results is that the analysis of seed has a good potential as a diagnostic tool for evaluating mineral nutrition of castor.

Key words: Castor cultivars, cationic nutrient content and uptake, internal nutrient ratio, potassium fertilization, seed analysis

Introduction

Castor (*Ricinus communis* L) is an industrial oilseed crop. In Andhra Pradesh, it is cultivated under rainfed conditions in red sandy loam (Alfisol) soils. These soils are inherently low in essential nutrients (Katyal, et al., 1997; Dar, 2004). Under intensive cultivation, positive response to potassium (K) application has been reported in castor (Murthy, 2001), indicating the plausible inadequacy of available K in these soils. Plant analysis is one of the important diagnostic tools to identify nutrient deficiency, sufficiency, and toxicity. Timely corrective measures either to crop or soil are imperative to sustain higher crop yields. Seed analyses have been used to determine the nutrient requirement of crops and uptake from the soils

(Moussavic-Nik, 1997; Murthy and Muralidharudu, 2003). Advantages of seed as diagnostic tissue is that they can be easily secured, cleaned and processed. Low nutritional status of seeds has been reported to reduce plant growth under conditions of low nutrient availability (Mengel and Kirkby, 1987). Genotypic variation in nutrient content of seed reflects the capability of cultivar to extract and accumulate nutrients from the soil. Present study was conducted to know the cationic nutrient content and uptake in seed of castor cultivars as influenced by K fertilization and to establish internal nutrient ratio requirements.

Materials and methods

Field experiments were conducted separately at Directorate of Oilseeds Research, Rajendranagar, Hyderabad, research farm with castor variety DCS-9 and hybrid DCH-32 as test crops during *kharif*, 2004, in red sandy loam (Alfisol) soil under rainfed conditions. Physico-chemical properties of the soil were pH 7.9, E.C 0.11 d S/m, Org. C 3.5 g/kg, cation exchange capacity 9.3 c mol (p+)/kg, available N 180, P₂O₅ 12 and K₂O 202 kg/ha. DTPA extractable micronutrients viz., Fe, Zn, Mn and Cu in soil were 2.53, 0.57, 7.20 and 0.62 mg/kg respectively. The treatments comprised 7 levels viz., 0, 20, 40, 60, 80, 100 and 120 kg K₂O/ha supplied through muriate of potash and phosphorus @ 40 kg/ha as basal doses to all the plots. Recommended dose of N (60 kg/ha) in three splits was applied to each castor cultivar. Experiments were conducted in RBD and replicated thrice. The spacing was 90 x 60 cm. Recommended crop husbandry practices were followed. Soil samples were collected from the experimental plots on 45th day after sowing at primary spike initiation stage and analysed for available potassium content with 1N ammonium acetate extractant as per the standard procedure. Calcium and magnesium content in the soil were determined by Versenate method. The available Zn, Fe, Cu and Mn in the soil samples extracted with DTPA solution (Lindsay and Norvell, 1978) were analysed using atomic absorption spectrophotometer (AAS - GBC 332 model). Seed samples collected at maturity, from the three picks (primary, secondary and tertiary) were composited and digested in 9:4 HNO₃: HClO₄ diacid. Potassium concentration in the acid digest aliquot was analysed by flame photometer. Calcium, magnesium, iron, zinc,

manganese and copper concentrations were analysed with AAS.

Internal nutrient requirements of the nutrient ratio in seed, which were sufficient for 94% maximum yield, instead of 95% (Fox *et al.*, 1986) were determined from graphs of relative yield versus nutrient ratios using boundary line technique (Webb, 1972). For brevity, one graph each for DCS-9 and DCH-32 are presented in Fig.1 and Fig.2, respectively.

Results and discussion

Seed yield: Castor cultivars showed significant variation in seed yields due to K fertilization. Seed yield of castor variety DCS-9 and DCH-32 varied from 852.2 to 1304.4 kg/ha and 871.1 to 1302.2 kg/ha respectively among the K doses. In the present study, optimum doses of K derived with linear response plateau equation were 87.1 ($r^2 = 0.98$) for DCS-9 and 72.5 ($r^2 = 0.79$) kg K_2O /ha for DCH-32. Castor seed yield responses to K fertilization up to 20 kg K_2O /ha were reported earlier under K stress conditions (Murthy and Muraidharudu, 2003).

Cationic nutrient content and uptake: Influence of K levels on K, Ca, Mg, Fe, Zn, Mn and Cu contents and uptake by the castor seeds was shown in Table 1. These cationic nutrient contents in the seed did not show any significant variation due to K fertilization in both the castor cultivars, DCS-9 and DCH-32. However, the nutrient contents in seeds were relatively higher than in control and followed the order $K > Mg > Ca > Fe > Zn > Cu > Mn$ irrespective of cultivar. Uptake of K, Mg, Fe, Mn and Cu by DCS-9 seed varied significantly due to the K application (Table 1). Potassium uptake of seed was significant over control at 20 kg K_2O /ha and was at par from 20 to 120 kg K_2O /ha. Magnesium uptake by DCS-9 seed showed a statistically significant variation at 60 kg K_2O /ha. The antagonistic K x Mg effect was observed at higher K levels while low level of K application could not show much antagonistic influence. Further, the movement of K and Mg from source to sink i.e., seed also might have been influenced by K levels. Increasing K supply affects the Mg content of different plant organs to a varying extent (Mengel and Kirkby, 1987). Interestingly, above 60 kg K_2O /ha, the Mg uptake was at par, indicating that increasing levels of K did not influence the Mg uptake by seed further. Similar trends were observed with micronutrients, Fe, Mn and Cu uptake in DCS-9 seed. Zinc uptake in seed varied from 23.5 to 40.4 g/ha, but was statistically non-significant. Relatively higher nutrient content and uptake noticed in DCH-32 hybrid seed could be attributed to vigour, better ability to extract and mobilise nutrients to economic parts. Significant K uptake by the DCH-32 seeds was observed at 60 kg K_2O /ha, while Mg and Cu uptake was influenced significantly at 20 kg K_2O /ha over control.

Nutrient ratios being found as relatively better index to

interpret plant analysis data than single nutrient content, cationic nutrient content ratios of soil and seed were computed (Table 2). Available soil cationic nutrient ratios have decreased with increasing levels of K application with few exceptions. Owing to ionic competition and antagonistic effects, the higher levels of K application might have suppressed the other nutrients. Amongst K levels, the seed nutrient ratios have not shown any definite trend in both the castor cultivars although there was a general decrease in ratio at 20 kg K_2O /ha over control. Growth factors (nutrients) interact together within plants, in ways that influence their effectiveness.

Internal nutrient ratio requirement: The "internal nutrient requirement" is the concentration of a nutrient in particular plant tissue associated with 95% of the estimated highest yield attainable when that nutrient, the primary limiting nutrient, is just adequately supplied for nutritional purposes (Fox *et al.*, 1986). Rashid and Fox (1992) evaluated internal zinc requirements of grain crops by seed analysis using boundary line technique. In the present investigation, the nutrient ratios were considered at 94% of the estimated highest castor seed yields and reported (Table 2, Fig. 1 and 2). The nutrient ratios, Ca: K, Mg: K, Fe: K, Zn: K, Mn: K and Cu: K associated with 94% of relative yield of castor DCS-9 are 0.049, 0.495, 0.050, 0.0057, 0.0024 and 0.0034 respectively. These values are considered as "internal nutrient ratio requirement" (INR) expected in DCS-9 seed under K fertilization. Similarly, the INR for castor hybrid, DCH-32 can be perceived (Table 2). These INR values are useful guidelines for assessing plant nutrition. Values observed above and below these may be due to the unknown source-sink relationships luxury consumption of nutrients and Steenbjerg (1951) effects. Boundary line approach is suggested superior to traditional regression analysis when the datum is influenced by unknown and uncontrolled factors (Rashid and Fox, 1992).

Soil nutrient ratios: Soil nutrient ratios have correlated significantly and negatively with the seed yields for both the castor cultivars, which mean lower the soil nutrient ratio higher the seed yields (Table 3). An optimum soil nutrient cation ratio necessary to achieve higher castor seed yields of DCS-9 and DCH-32 at 45th day was determined with quadratic equation. The optimum soil Ca+Mg: K, Fe: K, Zn: K, Mn: K and Cu: K ratios for DCS-9 were 0.074, 0.033, 0.0016, 0.081 and 0.0087 and for DCH-32 0.057, 0.069, 0.020, 0.011, 0.0084, respectively.

Thus, the present study showed that K has a significant influence on seed yields of both the castor variety DCS-9 and hybrid DCH-32 at 87 and 72 kg K_2O /ha respectively in red sandy loam soil under rainfed conditions. Cationic nutrient content and uptake in seeds of both the castor cultivars have varied by K fertilization. The INR established for seeds provided an initial guideline of these limits.

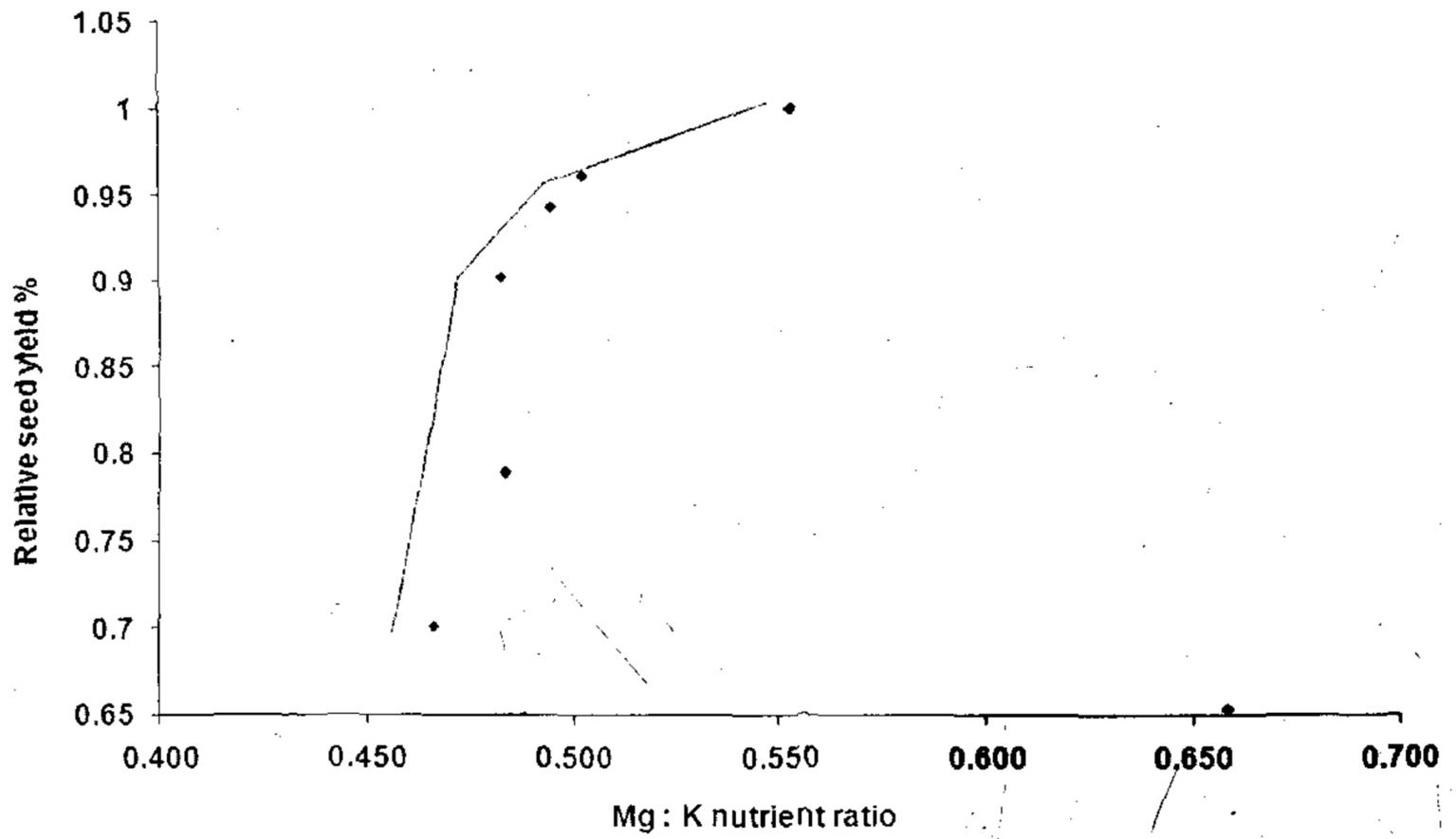


Fig. 1. Relationship between Mg:K nutrient ratio and seed yield of castor variety (DCS-9)

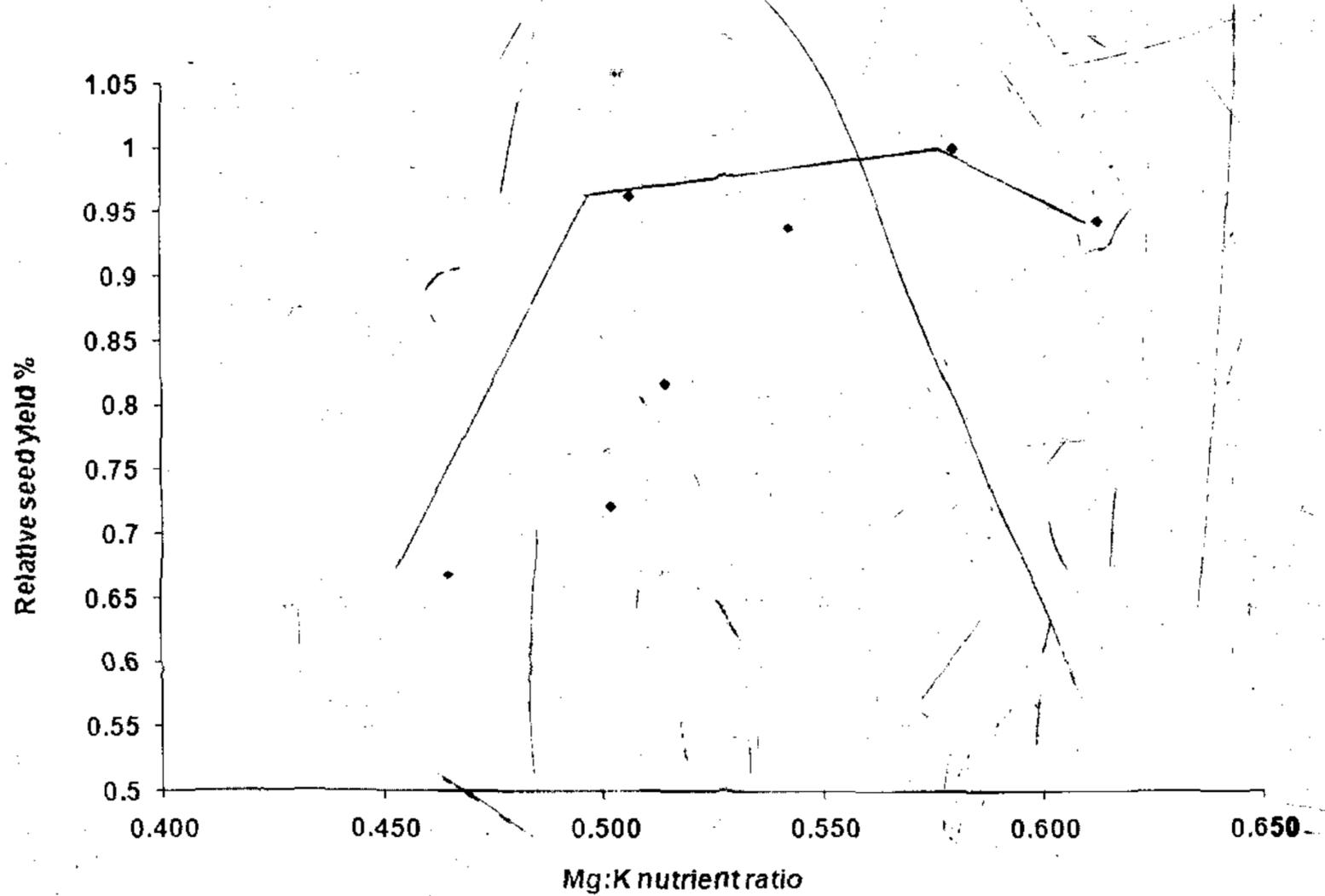


Fig. 2. Relationship between Mg:K nutrient ratio and seed yield of castor hybrid (DCH-32)

Table 1 Influence of potassium levels (kg/ha) on potassium, secondary and micronutrient cation i.e., Ca, Mg, Fe, Zn, Mn and Cu content (mg/kg) and uptake (g/ha) of castor seeds

K ₂ O levels (kg/ha)	K	Ca	Mg	Fe	Zn	Mn	Cu	K	Ca	Mg	Fe	Zn	Mn	Cu	Total seed yield (kg/ha)
	Content (mg/kg)							Uptake (g/ha)							
DCS-9															
0	3127.6	199.3	2058.1	182.4	27.5	9.7	15.3	2724.8	170.0	1753.6	155.4	23.5	8.3	13.0	852.2
20	4770.9	363.8	2222.2	242.9	30.9	11.9	15.2	4510.5	335.1	2035.5	222.6	28.4	10.9	13.9	914.4
40	4320.3	292.0	2086.6	207.9	29.4	11.3	14.2	5336.1	298.4	2152.5	213.9	30.3	11.6	14.7	1030.0
60	4724.5	352.8	2278.7	244.5	32.1	11.3	15.9	5020.4	414.5	2681.1	287.3	37.7	13.3	18.7	1176.7
80	4260.7	208.3	2107.4	214.4	24.5	10.1	14.7	5231.7	256.1	2590.1	263.5	30.1	12.5	18.0	1228.9
100	3982.4	286.5	2202.9	274.2	31.0	13.5	15.9	4993.3	373.9	2873.2	357.6	40.4	17.6	20.7	1304.4
120	4167.9	289.8	2094.3	189.0	31.1	9.3	16.4	5419.7	365.9	2628.7	236.9	38.9	11.7	20.5	1253.3
SEm±	342.0	72.0	126.2	28.9	3.0	1.3	0.6	542.5	78.9	166.1	32.6	3.6	1.6	0.8	10.6
CD (P=0.05)	NS	NS	NS	NS	NS	NS	NS	1672.1	NS	512.2	100.5	NS	4.9	2.5	33.0
DCH-32															
0	4035.4	277.4	1875.7	239.8	25.4	8.0	23.2	3438.5	241.3	1624.9	208.7	22.0	6.9	20.3	871.1
20	5029.3	564.9	2523.3	277.5	43.1	12.6	23.5	4622.8	534.1	2387.2	259.0	41.0	11.9	22.3	940.0
40	4313.7	408.7	2341.6	297.7	36.7	12.1	22.7	4455.2	513.5	2837.8	358.0	44.2	14.6	27.6	1222.2
60	4154.7	496.8	2136.8	257.7	32.9	10.8	22.5	4887.7	529.6	2272.5	274.0	34.9	11.4	23.8	1064.4
80	4035.4	338.3	2472.7	289.4	37.9	12.0	24.5	4959.1	416.4	3040.6	357.5	46.7	14.9	29.9	1228.9
100	4439.6	504.7	2248.1	288.6	37.7	13.3	23.7	5792.9	640.1	2814.1	365.4	47.7	16.8	29.8	1253.3
120	4028.8	435.6	2338.0	253.1	39.1	11.9	25.2	5050.8	582.9	3057.9	333.8	52.0	15.9	32.9	1302.2
SEm±	326.3	70.2	169.0	30.7	5.4	1.7	0.8	345.1	108.3	249.3	43.6	7.6	2.3	2.0	15.6
CD (P=0.05)	NS	NS	NS	NS	NS	NS	NS	1063.7	NS	768.5	NS	NS	NS	NS	48.0

Table 2 Influence of potassium levels on secondary and micro nutrient ratios in red soil and castor seeds of DCS-9 and DCH-32

K ₂ O levels	Ca + Mg: K	Fe: K	Zn: K	Mn: K	Cu: K	Relative yield %	Ca: K	Mg: K	Fe: K	Zn: K	Mn: K	Cu: K
	Soil						Seed					
DCS-9												
0	0.163	0.022	0.0094	0.051	0.0051	65.3	0.064	0.658	0.058	0.0088	0.0031	0.0049
20	0.143	0.040	0.0077	0.068	0.0074	70.1	0.076	0.466	0.051	0.0065	0.0025	0.0032
40	0.137	0.039	0.0056	0.102	0.0115	79.0	0.068	0.483	0.048	0.0068	0.0026	0.0033
60	0.121	0.019	0.0046	0.057	0.0061	90.2	0.075	0.482	0.052	0.0068	0.0024	0.0034
80	0.105	0.015	0.0037	0.053	0.0046	94.2	0.049	0.495	0.050	0.0057	0.0024	0.0034
100	0.100	0.014	0.0044	0.040	0.0038	100	0.072	0.553	0.069	0.0078	0.0034	0.0040
120	0.053	0.010	0.0018	0.035	0.0032	96.1	0.070	0.502	0.045	0.0075	0.0022	0.0039
DCH-32												
0	0.169	0.028	0.0060	0.092	0.0071	66.9	0.069	0.465	0.059	0.0063	0.0020	0.0057
20	0.166	0.019	0.0042	0.070	0.0079	72.2	0.112	0.502	0.055	0.0086	0.0025	0.0047
40	0.137	0.018	0.0049	0.075	0.0079	93.9	0.095	0.543	0.069	0.0085	0.0028	0.0053
60	0.121	0.018	0.0058	0.070	0.0063	81.7	0.120	0.514	0.062	0.0079	0.0026	0.0054
80	0.082	0.018	0.0035	0.068	0.0062	94.4	0.084	0.613	0.072	0.0094	0.0030	0.0061
100	0.075	0.014	0.0028	0.053	0.0040	96.2	0.114	0.506	0.065	0.0085	0.0030	0.0053
120	0.060	0.012	0.0025	0.044	0.0035	100	0.108	0.580	0.063	0.0097	0.0030	0.0063

Table 3 Correlation values @ between available soil nutrient ratios and castor seed yield

Available soil nutrient ratio	Seed yield	
	DCS-9	DCH-32
Ca+Mg: K	-0.841**	-0.889**
Fe: K	-0.713**	-0.852**
Zn: K	-0.912**	-0.741**
Mn: K	-0.472*	-0.778**
Cu: K	-0.494*	-0.643**

*,** = significant at 5% and 1% level, respectively.

References

Dar William, D. 2004. Macro benefits from micronutrients for grey to green revolution in agriculture. *IFA International symposium on micronutrients*. 23-25 February 2004. New Delhi, India.

Fox, R.L., Saunders, W.M.H. and Rajan, S.S.S. 1986. Phosphorus nutrition of pasture species: Phosphorus requirement and root saturation values. *Soil Science Society of America Journal*, 50: 144-148.

Effect of potassium fertilization on cationic nutrient content and uptake in seeds of two castor cultivars

- Katyal, J.C., Sharma, K.L., Srinivas, K. and Narayana Reddy, M. 1997.** Balanced fertilizer use in semi-arid soils. *Fertilizer News*, **42** (4): 59-69.
- Lindsay, W.L. and Norvell, W.A. 1978.** Development of a DTPA soil test for zinc, iron, manganese and copper. *Soil Science Society of America Journal*, **42**: 421-428.
- Mengel, K. and Kirkby, E.A. 1987.** *Principles of plant nutrition*. Bern, Switzerland, International Potash Institute.
- Moussavic-Nik, M. 1997.** Seed quality and crop establishment in wheat. Ph.D thesis, University of Adelaide, Adelaide, South Australia.
- Murthy, I.Y.L.N. 2001.** Micronutrient uptake in seed of sunflower (*Helianthus annuus L.*) hybrids. *Journal of Oilseeds Research*, **18** (1): 128-130.
- Murthy, I.Y.L.N. and Muralidharudu, Y. 2003.** Response of Castor (*Ricinus communis L.*) under K stress condition. In: *National seminar on Stress Management in Oilseeds for attaining self-reliance in Vegetable Oils*. Jan. 28-30, 2003. Indian Society of Oilseeds Research, DOR, Hyderabad. pp.304-305.
- Rashid, A. and Fox, R.L 1992.** Evaluating internal zinc requirements of grain crops by seed analysis. *Agronomy Journal*, **84**: 469-474.
- Steenbjerg, F. 1951.** Yield curves and chemical plant analysis. *Plant and Soil*, **3**: 97-109.
- Webb, R.A. 1972.** Use of the boundary line in the analysis of biological data. *Journal of Horticultural Science*, **47**: 309-319.

Nutrient requirements of *kharif* castor, *Ricinus communis* L. under irrigated conditions of Uttar Pradesh

S.K. Srivastava and D.R. Chandra

Oilseed Section, Chandra Shekhar Azad University of Agriculture & Technology, Kanpur-208 002, UP

(Received: January, 2009; Revised: August, 2009; Accepted: September, 2009)

Abstract

Field experiment conducted at Kanpur (UP) during *kharif* 2006-07 and 2007-08 to find out the optimum fertilizer needs of castor under irrigated conditions, revealed that with increase in nitrogen dose, there was significant increase in seed yield upto 80 kg N/ha. Application of 30 kg phosphorus/ha was found optimum. The crop responded to potassium application in the year 2007-08 upto 30 kg K₂O/ha. Application of 80 kg N/ha, 30 kg P₂O₅ and 30 kg K₂O/ha gave maximum net returns and B:C ratio during both the years.

Key words: Castor, irrigated conditions, fertilizer, economics

Introduction

Castor (*Ricinus communis* L.) is an important non-edible oilseed crop of India which fetches sizeable amount of foreign exchange to the country through export (Reddy *et al.*, 2006). India's share in world's area and production of castor is 68% and 76%, respectively (Reddy *et al.*, 2006). Oilseeds are energy rich crops and require high amount of nutrients for their production. Inadequate or imbalanced supply of nutrients has been identified as one of the critical constraint limiting oilseeds production. Castor is found to respond to major nutrients, more particularly to nitrogen and phosphorus together rather than singly (Patel *et al.*, 2007). Agro-climatic conditions of Uttar Pradesh are quite suitable for castor cultivation. The present study was made to find out the optimum requirement of fertilizers under irrigated conditions of Uttar Pradesh.

Materials and methods

The field experiment was carried out at Oilseeds Research Farm, Kalyanpur, C.S.A. University of Agriculture & Technology, Kanpur during *kharif* 2006-07 and 2007-08 to find out the optimum fertilizer doses (N, P and K) for castor to obtain maximum yield and returns under irrigated conditions of Uttar Pradesh. The soil of the experimental field was sandy loam in texture, with pH 7.8, low in organic carbon (0.43%), low in available phosphorus (11.7 kg/ha) and medium in available potash (150 kg/ha). Treatment combinations consisting of 4 nitrogen levels (0, 40, 80 and 120 kg N/ha), 3 phosphorus levels (0, 30 and 60 kg P₂O₅/ha) and 2 potassium levels (0

and 30 kg K₂O/ha) were laid out in Randomized Block Factorial Design with 3 replications. The fertilizers were applied as per treatment in the form of urea, diammonium phosphate, single super-phosphate and muriate of potash. Half of N with full dose of P and K was applied basal, and the remaining half nitrogen was top dressed in two equal splits at 30 and 45 days after sowing. Seeds were treated with carbendazim @ 1g/kg of seeds to protect from seed borne diseases. Castor crop was sown on 14 July, 2006 and Aug. 10, 2007 using variety Kalpi-6 in 2006 and hybrid GCH-4 in 2007. Seeds were dibbled @ 2 seeds/hill at a depth of 4-5 cm in the rows 90 cm apart keeping 60 cm plant spacing. All the recommended cultural practices and required suitable plant protection measures were adopted to raise a good crop. The crop was harvested in three pickings manually based on physiological maturity of the capsules. The crop received a rainfall of 499.5mm and 525.6mm during crop season in 2006 and 2007, respectively.

Results and discussion

Response to nitrogen: Significant linear enhancement in plant height, number of spikes/plant, spike length, number of capsules/spike and finally seed yield was recorded with increase in nitrogen dose from 0 to 80 kg N/ha. Venugopal *et al.* (2006) also found the highest number of spike/plant, spike length and number of capsules/spike with the application of 80 kg N/ha. Application of 80 kg N/ha remained comparable with 120 kg N/ha, gave significantly higher value of all these growth and yield attributes than 0 and 40 kg N/ha. Similar results were reported by Anonymous, (2008) from Bawal (Haryana). Better nutrition could have resulted in more number of spikes/plant and capsules/spike (Mathuria and Modhwadia, 1993).

Application of 80 kg N/ha produced significantly higher seed yield than no nitrogen and 40 kg N/ha. Venugopal *et al.* (2006), Anonymous (2007) under Jagdalpur (Chhattisgarh) conditions, Anonymous (2008) under Bawal (Haryana) and Hiriyur (Karnataka) agro-climatic conditions reported similar results. Patel *et al.* (1991) and Sardana *et al.* (2008) recorded the response upto 75 kg N/ha. Supply of 40 kg N/ha also achieved significantly higher seed yield than control while no application of nitrogen (control) resulted in significantly lowest seed yield. There was no remarkable variation in seed yield with 80 and 120 kg N/ha

during both the years. Nitrogen application in adequate quantity has promoted the growth and improved yield attributes of castor, resulting in highest seed yield. This confirms the findings of Ganga Saran and Gajendragiri (1987).

Response to phosphorus: Application of phosphorus 30 kg P_2O_5 /ha remaining comparable with 60 kg P_2O_5 /ha recorded significantly higher seed yield than no phosphorus application. Thus, 30 kg P_2O_5 /ha was found optimum. Similar findings were also reported by Anonymous (2007) from Jagdalpur (Chhattisgarh) and Anonymous (2008) from Hiriyur (Karnataka). Patel *et al.* (2007) observed that application of 25 kg P_2O_5 /ha gave significantly higher seed yield of castor than no fertilization (control) while application of 40 kg P_2O_5 /ha was found optimum for castor production by Sardana *et al.* (2008) under Punjab agro-climatic conditions.

Response to potassium: Castor responded to potassium application only in the year 2007-08, where application of 30 kg K_2O /ha produced significantly taller plants, more number of spikes/plant, longer spikes and more number of capsules/spike than control. Seed yield was also found significantly superior with 30 kg K_2O /ha to no potassium application during the year 2007-08, while no significant response was noticed to application of potash during the

year 2006-07. Response of castor upto 30 kg K_2O /ha was also reported by Anonymous (2007) from Jagdalpur (Chhattisgarh), Anonymous (2007) from Bawal (Haryana) and by Anonymous (2008) from Hiriyur (Karnataka) and Bawal (Haryana) did not observe significant response to potassium application in castor.

Thus, application of 80 kg N/ha and 30 kg P_2O_5 /ha was found optimum for castor variety, while castor hybrid required 80 kg N/ha, 30 kg P_2O_5 /ha and 30 kg K_2O /ha for its optimum yield. Application of 80 kg N/ha, 30 kg P_2O_5 /ha and 30 kg K_2O /ha was found optimum under Hiriyur (Karnataka) agro-climatic conditions.

Economics: Economic evaluation of N,P and K combinations showed that application of 80 kg N/ha, 30 kg P_2O_5 /ha and 30 kg K_2O /ha resulted in maximum gross returns, net returns and B:C ratio during the year 2007-08, while net returns and B:C ratio were maximum with this N,P and K combination during both the years. The highest net returns accrued with the application of 80 kg N/ha, 30 kg P_2O_5 /ha and 30 kg K_2O /ha could be attributed to higher seed yield. Venugopal *et al.* (2006) also reported the highest net returns of castor with 80 kg N/ha. Highest gross returns, net returns and benefit cost ratio were also obtained with the application of 100% N through fertigation in Gujarat in castor (Patel *et al.*, 2006).

Table 1 Effect of N, P, K fertilizers on growth, yield attributes and bean yield of castor

Fertilizers	Plant height (cm)		Spikes/plant		Spike length (cm)		Capsules/spike		Total seed yield (kg/ha)	
	2006-07	2007-08	2006-07	2007-08	2006-07	2007-08	2006-07	2007-08	2006-07	2007-08
Nitrogen levels (kg N/ha)										
0	73.3	80.5	3.4	4.3	18.7	57.4	25	67	511	1742
40	103.4	87.3	3.7	4.7	27.1	62.0	38	74	883	2120
80	128.1	92.3	5.0	5.3	35.9	66.6	55	83	1374	2701
120	135.0	92.3	4.9	5.2	35.9	65.4	55	81	1387	2627
SEm±	0.5	0.7	0.1	0.07	0.4	0.7	0.4	0.6	23	36
CD (P=0.05)	1.5	1.9	0.2	0.19	1.0	1.9	1.0	1.8	67	102
Phosphorus levels (kgP₂O₅/ha)										
0	102.8	85.0	3.8	4.3	26.6	57.4	39	67	856	1763
30	113.5	89.0	4.5	5.2	30.8	66.0	45	81	1117	2623
60	113.6	90.4	4.5	5.1	30.9	65.1	46	79	1142	2505
SEm±	0.4	0.6	0.1	0.06	0.3	0.6	0.3	0.6	20	31
CD (P=0.05)	1.3	1.6	0.2	0.17	0.9	1.6	0.9	1.6	58	89
Potassium levels (kgK₂O/ha)										
0	110.0	87.5	4.3	4.7	29.4	60.9	43	73	1032	2146
30	109.9	88.8	4.3	5.1	29.4	64.8	43	79	1045	2449
SEm±	0.4	0.5	0.1	0.05	0.3	0.5	0.3	0.5	17	25
CD (P=0.05)	N.S	1.3	N.S	0.14	N.S	1.3	N.S	1.3	N.S	72

Table 2 Effect of fertilizers on economics of castor production

Treatment	Seed yield (kg/ha)		Gross returns (Rs./ha)		Cost of cultivation (Rs./ha)		Net returns (Rs./ha)		B.C. ratio	
	2006-07	2007-08	2006-07	2007-08	2006-07	2007-08	2006-07	2007-08	2006-07	2007-08
Nitrogen levels (kg/ha)										
0	511	1742	8080	34843	11819	18347	-3739	15496	0.68	1.80
40	883	2120	13937	42400	12102	19698	1835	22702	1.15	2.15
80	1374	2701	21626	54000	12570	20147	9056	33853	1.72	2.68
120	1387	2627	21907	52533	13044	20624	8863	31909	1.68	2.55
Phosphorus levels (kgP2O5/ha)										
0	856	1763	13510	35262	11796	19352	1714	15910	1.15	1.82
30	1117	2623	17629	52462	12390	19952	5239	32510	1.42	2.63
60	1142	2505	18024	50107	12965	20558	5059	29549	1.39	2.44
Potassium levels (kgK2O/ha)										
0	1032	2146	16322	42913	12246	19816	4076	23097	1.33	2.17
30	1045	2449	16453	48975	12521	20092	3932	28883	1.31	2.44

References

Anonymous. 2007. Annual Progress Report of castor for year 2006-07, Directorate of Oilseeds Research, Hyderabad, pp. 143-145.

Anonymous. 2008. Annual Progress Report of castor for year 2007-08, Directorate of Oilseeds Research, Hyderabad, pp. 118-119.

Ganga Saran and Gajendragiri. 1987. Effect of seeding time and nitrogen on summer castor. *Indian Journal of Agronomy*, **32** (2) : 155-157.

Mathuria, R.K. and Modhwadia, M.M. 1993. Response of castor (*Ricinus communis* L.) to nitrogen and phosphorus. *Indian Journal of Agronomy*, **38**(1) : 152-153.

Patel, K.S., Patel, B.A., Patel, M.K., Patel, G.N. and Pathak, H.C. 2007. Integrated Nutrient Management for Castor - Sorghum (fodder) Cropping System. National Seminar of Indian Society of Oilseeds Research, 29-31 January 2007, pp. 345-346.

Patel, K.S., Patel, M.K., Patel, G.N. and Pathak, H.C. 2006. Fertigation study in castor, *Ricinus communis* L.. *Journal of Oilseeds Research*, **23** (1) : 122-123.

Patel, M.K., Fateh, U.G. and Patel, V.J. 1991. Effect of nitrogen and its time of application on yield of castor GAUCH-1 (*Ricinus communis* L.) under irrigated condition in North Gujarat. *Journal of Research, Gujarat Agricultural University*, **17** : 27-29.

Reddy, A. Pratap Kumar, Reddy, A. Sambasiva and Padmavathi, P. 2006. Effect of irrigation and integrated nutrient management on seed and oil yield of rabi castor, *Ricinus communis* L. *Journal of Oilseeds Research*, **23** (2) : 239-241.

Sardana, Virender, Singh, Jayesh and Bajaj, R.K., 2008. Investigation on sowing time, plant density and nutrient requirements of hybrid castor, *Ricinus communis* L. for the non-traditional area of Punjab. *Journal of Oilseeds Research*, **25** (1) : 41-43.

Venugopal, C., Reddy, G., Krishna and Reddy, D. and Srinivasulu. 2006. Seed yield and net returns of rainfed castor, *Ricinus communis* L. as influenced by plant geometry and nitrogen levels. *Journal of Oilseeds Research*, **23** (2) : 356-357.

Cultural and morphological variability in *Alternaria brassicae* isolates of Indian mustard, *Brassica juncea* L. Czern & Coss.

Dhiraj Singh, Rajender Singh, Harbinder Singh, Ram Chander Yadav, Neelam Yadav, Martin Barbetti¹ and Phil Salisbury²

Chaudhary Charan Singh Haryana Agricultural University, Hisar-125 004, Hisar

(Received: September, 2007; Revised: August, 2009; Accepted: December, 2009)

Abstract

Alternaria brassicae is the most virulent on all brassicaceous plants and cause adverse effect on both quality and quantity of the crop. The present investigation was carried out to know the cultural and morphological variability in *Alternaria brassicae* causing *alternaria* blight of oilseeds Brassica. One hundred and five disease samples were collected from 18 districts of Haryana (India) at 20 to 25 km intervals. The spot/lesion size on leaves of the collected samples ranged from 3.0 to 11.5 mm. These samples were isolated and purified by single spore technique to study the morphological, cultural and radial growth behavior at 20 and 25°C. The radial growth varied from 34.6-81.1 mm with creamish, light brown to dark brown in colour and compressed to fluffy mycelial growth. The average conidial length ranged from 117.0 to 192.0 µm and breadth from 14.0 to 24.0 µm. The conidial beak length varied from 42.0 to 116.0 µm, number of horizontal/longitudinal septa ranged from 6 to 9 and vertical/transverse septa ranged from 1 to 3 and average distance between two septa have also been determined.

Key words: *Alternaria brassicae*, morphological variability, cultural variability

Introduction

Alternaria blight caused by *Alternaria brassicae* (Berk) Sacc., is an economically important disease of oilseed brassica in many parts of the world which cause severe losses both in terms of quantity and quality. In India, the yield losses to the extent of 70% have been reported (Chahal, 1986; Saharan, 1991). Out of four species of *Alternaria* known to occur on this crop, *Alternaria brassicae* is more severe one (Verma and Saharan, 1994). Preliminary reports on variability in *Alternaria* species were made from Holland (Van Schreven, 1953) and UK (Mridha, 1983). There are number of reports on the existence of variability in *A. brassicae* in India on the basis of morphology, growth, cultural characteristics and

reaction to host differentials. However, complete information is not available on morphological variability of *A. brassicae* isolates collected from Haryana state. Therefore present investigation was undertaken to find out the cultural and morphological variability in *A. brassicae* isolated collected from all over Haryana.

Materials and methods

Alternaria blight infected 105 samples were collected from 18 districts of Haryana at a distance of 20-25 km. The pathogen *A. brassicae* (Berk) Sacc., was isolated from diseased leaves as per Dhingra and Sinclair (1985). The fungal colonies showing characteristics of *A. brassicae* was picked up and sub cultured in petri plates containing PDA supplemented with rose Bengal. These plates were incubated in BOD incubator at 25±1°C for 4 to 5 days. Isolates were then purified by single spore technique (Toussoun and Nelson, 1976). Isolates were maintained on PDA slants in a refrigerator at 5°C for further studies. The spot/lesion size on leaves of each collected sample was measured. Cultural characteristics of each isolates such as radial growth at 20 and 25°C, colony colour characteristics and growth behavior was observed. Morphological characteristics of each isolates such as conidia length, breadth, number of septations, beak length and average distance between septum were recorded.

Results and discussion

The collected *Alternaria* blight samples have variation in spot/lesion size and symptoms produced. The spot size varies from 3.0 to 11.5 mm in diameter having light brown to dark brown coloration with less to prominent concentric rings. The sample collected from Yamuna Nagar district have minimum spot size (3.0 mm) having light brown spot with less visible rings. The maximum spot size was observed from Sirsa district i.e., 11.5 mm with medium to large dark brown spots with well developed concentric rings (Table 1) The cultural characteristics such as growth behavior and colony character were studied at 20 and 25°C.

¹ School of Plant Biology, Faculty of Natural and Agr. Sciences, The University of Western Australia, 35 Stirling Highway, Crawlye, WA, 6009, Australia

² School of Agriculture and Food Systems, The University of Melbourne, Victoria 3010, Australia

Table 1 Symptoms and spot size of *Alternaria* samples collected from different districts of Haryana (India)

District	Avg. Spot size (mm)	Symptoms
Jind	7.8	Dark brown irregular shape spot with concentric ring
Panipat	5.4	Brown spots with yellowish margin
Sonepat	5.1	Brown spot with less visible concentric rings
Fatehabad	3.8	Dark brown spot with yellowish margin
Kurukshetra	4.8	Irregular, brown spots with less visible concentric rings
Karnal	8.1	Light brown spots with whitish margin, less visible concentric rings
Yamuna nagar	3.0	Light brown spots, less visible rings
Hisar	4.8	Dark brown spot with concentric rings
Bhiwani	6.2	Medium light to dark brown spots with concentric rings
Rohtak	5.3	Small greenish brown spots with irregular shape
Jhajjar	9.4	Light brown spots, concentric rings with papery growth
Rewari	5.5	Small dark brown spots with concentric rings
Kaithal	6.5	Light brown spots with yellowish margin
Mahender Garh	7.0	Dark brown spots with well developed concentric rings
Sirsa	11.5	Medium to large dark brown spots with concentric rings
Faridabad	5.5	Dark brown spots
Gurgaon	8.2	Brown spots with concentric rings
Ambala	5.2	Whitish to brown irregular spots

The pure culture of isolates collected from Yamuna Nagar have maximum growth i.e., 73.6 and 81.1 mm both at 20°C and 25°C followed by isolates collected from Hisar with creamy to dark brown colony character. Isolates collected from Bhiwani district had minimum growth i.e., 34.6 and 40.5 mm with dark brown colony characteristics both at 20 and 25°C (Table 3). Morphological observations recorded on each isolate revealed that isolates differed in their conidial size. The range of conidial length varies from 81.0 to 293.5 µm. Isolates collected from Jind have maximum conidial length. The conidial breadth also ranges from 8.1 to 31.5 µm. The thickest conidium was of isolate Kurukshetra and thinnest of isolate Sonepat. The horizontal septation varied from 4-12 and vertical from 0 to 5. The septum distance between two septa also showed some variation which ranges from 6.7 to 14.8 µm. Some variation also recorded in beak length and width. The average beak length varied from 26.2 to 225.2 µm. The longest beak was of isolate Rohtak and smallest of Ambala. The beak septation no. ranges from 0-9, (Table 2, Fig. 1).

Isolates collected from Rohtak have

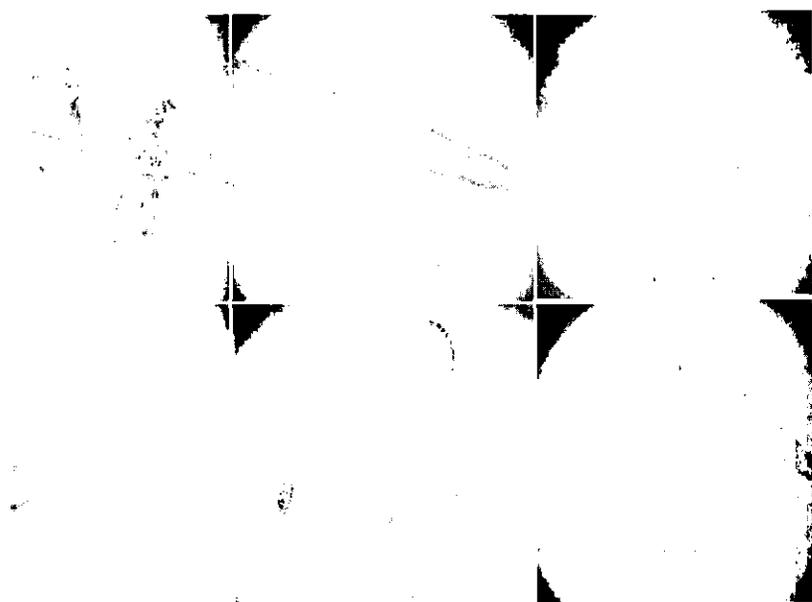
Fig. 1. Morphological variability in *Alternaria brassicae* isolates

Table 2 Morphological variation in conidial size of *Alternaria brassicae* collected from different districts of Haryana (India)

District	Conidial length (μm)	Conidial width (μm)	No. of Septation		Beak		Septum distance (μm)
			Horizontal	Vertical	Length (μm)	Septation No.	
Ambala	118.5(94.5-148.5)	13.8(11.5-19.2)	6.2(5-7)	0.7(0-3)	41.5(27.0-54.0)	1.8(0-5)	6.7(5.4-9.4)
Jind	117.1(81.0-163.4)	16.2(12.1-20.2)	5.6(4-8)	0.8(0-3)	41.8(30.4-51.5)	2.2(1-6)	13.5(12.8-16.2)
Fatehabad	151.2(108.0-192.5)	16.2(13.5-18.9)	5.7(5-8)	1.2(1-5)	54.0(27.0-94.5)	2.1(1-5)	8.1(5.4-10.8)
Gurgaon	191.7(135-233.5)	18.3(14.8-18.3)	7.8(5-12)	1.3(0-4)	54.0(40.5-82.3)	2.7(0-4)	9.4(8.1-13.5)
Y. Nagar	120.2(87.5-154.6)	16.2(14.8-21.6)	5.5(4-7)	1.0(0-4)	42.5(27.0-67.5)	2.2(1-4)	14.8(13.5-16.2)
Faridabad	191.6(121.0-293.5)	18.9(14.8-22.95)	8.4(7-12)	1.3(1-3)	60.7(27.0-82.7)	2.1(0-5)	10.8(9.4-13.5)
Panipat	141.7(94.5-192.5)	17.5(13.5-27.0)	6.8(5-9)	1.1(1-4)	52.5(40.5-67.5)	1.6(1-4)	10.8(8.1-12.7)
Karnal	121.5(91.5-148.5)	16.2(13.5-21.6)	5.5(5-8)	0.8(0-3)	67.5(40.5-87.7)	1.2(1-5)	11.7(8.1-13.5)
Kurukshetra	135.0(94.0-168.5)	21.5(16.5-31.5)	6.8(5-9)	1.0(0-4)	62.5(33.5-81.0)	1.4(1-3)	13.5(9.4-16.2)
Sonepat	148.5(108-186.5)	13.5(8.1-20.2)	5.6(5-8)	0.8(0-3)	54.0(40.5-94.5)	1.2(1-4)	8.1(5.4-13.5)
Hisar	117.3(81.8-161.6)	15.3(9.0-27.1)	6.5(5-9)	1.5(0-3)	47.1(26.2-81.8)	2.2(1-3)	9.4(7.5-12.4)
Bhiwani	137.7(89.9-180.8)	16.8(10.1-25.7)	7.2(6-10)	1.2(0-3)	67.1(45.2-116.1)	4.7(2-8)	10.7(9.2-15.5)
Rohtak	192.5(112.0-290.9)	20.5(9.3-27.2)	6.2(5-9)	2.1(0-4)	116.0(34.3-225.2)	5.6(3-9)	6.9(5.3-10.2)
Jhajjar	165.1(87.8-206.0)	18.2(12.5-25.2)	6.5(4-9)	2.1(0-4)	65.1(35.3-116.1)	1.7(0-3)	12.2(8.7-15.8)
Rewari	157.5(90.9-241.4)	14.4(12.8-18.1)	7.3(5-9)	2.3(0-4)	80.7(35.2-136.3)	3.2(2-5)	13.6(11.8-14.9)
Kaithal	132.9(86.8-207.8)	17.5(16.6-20.8)	7.7(6-11)	2.6(0-5)	42.9(27.2-86.9)	1.8(0-3)	9.8(7.3-12.3)
M. Garh	135.5(90.9-180.8)	17.2(9.9-24.7)	7.4(6-11)	1.2(0-3)	65.2(45.2-118.2)	4.7(2-9)	9.9(7.5-13.4)
Sirsa	146.2(89.9-189.7)	18.1(17.3-21.7)	8.5(7-11)	2.0(0-4)	43.7(31.2-90.9)	2.7(1-5)	11.1(9.8-14.5)

Table 3 Radial growth, colour and growth pattern of different isolates of *A. brassicae* collected from different districts of Haryana (India) on Indian mustard

District	Radial growth at		Colour	Growth pattern
	20°C	25°C		
Karnal	41.3	45.5	Dark brown	Compressed
Panipat	44.6	48.3	Brownish	Compressed
Fatehabad	38.3	44.6	Light Brown	Compressed
Bhiwani	34.6	40.5	Dark Brown	Compressed
Mahendergarh	42.7	50.0	Brown	Compressed
Rewari	51.3	60.0	Light Brown	Slightly Compressed
Gurgaon	45.5	56.6	Creamy whitish	Fluffy
Faridabad	55.5	66.7	Rough whitish	Fluffy
Hisar	57.6	68.6	Brown to dark brown	Compressed
Kaithal	51.3	56.6	Light brown to rough whitish	Fluffy
Sirsa	37.3	41.3	Light brown	Fluffy
Jind	40.0	46.6	Light brown	Compressed
Rohtak	46.6	55.5	Whitish	Compressed to Fluffy
Ambala	43.3	48.3	Creamy whitish	Fluffy
Jhajjar	46.6	50.0	Creamy white to light brown	Fluffy
Sonepat	53.3	60.0	Whitish	Fluffy
Kurukshetra	45.5	52.5	Light brown	Compressed
Yamuna Nagar	73.6	81.1	White to creamy chocolate	Compressed to Fluffy

Earlier workers also described morphological variation in *A. brassicae* isolates (Saharan and Kadian, 1983). The current results also describe that collected *A. brassicae* isolate have lot of morphological and cultural variation. The conidial length ranges from 81.0 to 293.5 μm and breadth from 8.1 to 31.5 μm . Kumar *et al.* (2003) observed average conidial length from 118.62 to 194.52 μm and breadth from 14 to 23 μm . They further observed some variation in beak length and number of septation. Mehta *et al.* (2003) categorized *A. Brassicae* isolates in four groups i.e., small (<100 μm), Medium (101-150 μm), long (151-200 μm) and very long (>200 μm). These variation in conodinal size may be due to variation in medium concentration and location/site of different field.

The cultural and morphological difference in *A. brassicae* isolates indicates significant variation in the pathogen, it may be attributed due to environmental variation and racial differences.

References

- Chahal, A.S. 1986. Losses and chemical control of *Alternaria* in rapeseed-mustard in Punjab. *Plant Disease Research*, 1: 46-50.
- Dhingra, O.D. and Sinclair, J.B. 1985. *Basic Plant Pathology Methods*. CRC Press, Inc. Boca Raton, Florida. pp.335.
- Kumar Satish, Sangwan, M.S., Mehta Naresh and Kumar Rakesh. 2003. Pathogenic diversity in Isolates of *Alternaria brassicae* infecting rapeseed and mustard. *Journal of Mycology and Plant Pathology*, 33(1) :59-64.
- Mehta Naresh, Sangwan, M.S. and Srivastava. M.P. 2003. Morphological and pathological variations in rapeseed and mustard isolates of *Alternaria brassicae*. *Indian Phytopathology*, 56(2) : 188-190.
- Mridha, M.A.U. 1983. Virulence of different isolates of *Alternaria brassicae* on winter oilseeds rape cultivars. 6th International Rapeseed Conference. Paris, France, pp 1025-1029.
- Saharan, G.S. 1991. Assessment of losses, epidemiology and management of black spot disease of rapeseed-mustard, GRIC 8th Proceedings of International Rapeseed Congress, July 9-11, Saskatoon, Canada, II: 465-470.
- Saharan, G.S. and Kadian, A.K. 1983. Physiological specialization in *Alternaria brassicae*. *Crucifereae Newsletter*, 8:32-33.
- Tousson, T.A. and Nelson, P.E. 1976. A Pictorial guide to the Identification of *Fusarium* spp. According to the Taxonomic system of Synder and Hanson. University Park, Pennsylvania, USA; Pennsylvania State University Press, p.43.
- Van Shreven, D.A. 1953. *Alternaria*, *stemphylium* and *Botrytis* infection of colza (*Brassica napus*). *Tizdschr. Planterzickten*, 59: 105-136.
- Verma, P.R. and Saharan, G.S. 1994. *Monograph on Alternaria diseases of crucifers*. Research Branch, Agriculture and Agriculture Food Canada, Saskatoon Res. Centre, Canada, pp.162.

Influence of castor, *Ricinus communis* L. as relay crop in controlling root knot disease of groundnut caused by *Meloidogyne arenaria* (Neal) chitwood

C. Lukose, A.M. Moradia, I.U. Dhruj, R.S. Savalia, L.D. Vavadia and B.A. Kunadia

Junagadh Agricultural University, Junagadh-362 001, Gujarat

(Received: January, 2008; Revised: July, 2009; Accepted: July, 2009)

Abstract

Root knot disease of groundnut caused by *Meloidogyne arenaria* is a serious problem in a few farmers' fields of Junagadh and Rajkot districts in Saurashtra region of Gujarat. An experiment was conducted during *kharif* season 2002-03-04 and 2005 in farmers' fields in Junagadh district to find out the effect of relay crops *viz.*, castor (GCH-4), pigeon pea (BDN-2), and sesame (G Til-1) in groundnut in combination with nematicide in managing *M. arenaria*. The data on nematode population, root-knot index and pod equivalent yield data revealed that relay cropping of groundnut with castor (row ratio of 2:1) along with soil application of carbofuran 3G @ 1 kg a.i./ha was highly effective in managing the root knot disease and also in increasing the yield with a CBR of 1:2.35.

Key words: Castor, relay crop, control, root knot nematode, groundnut

Introduction

Groundnut is the most important oilseed crop grown in India, particularly in Gujarat. Plant parasitic nematodes are widespread and important pests of groundnut. Root knot disease caused by *Meloidogyne arenaria* in groundnut has become widespread and causes serious problem at farmers' fields in Junagadh and Rajkot districts in Saurashtra region of Gujarat. The heavily infested plants remain stunted with chlorotic dwarf leaves and profusely galled roots with few lateral roots (Kalaiarasan *et al.*, 2006). Crop losses due to *M. arenaria* on groundnut were about 18% in Saurashtra. Patel *et al.* (1985) observed reduced penetration of *Meloidogyne arenaria* on groundnut with seed treatment of carbofuran and aldicarb as compared to control. Soil application of carbofuran has also been found to reduce the root knots and induce good plant growth. In the past, considerable attention has been paid towards the control of nematode diseases by intercropping/ relay cropping of non-host plants. Gowda and Reddy (1995) studied the management of root knot nematode in tobacco nursery with castor as trap crop along with phenamiphos. The use and practical implication of non-host plants as relay crop has also been reported (Alam *et al.*, 1977). Therefore, the present

investigation was carried out to find out the effect of growing relay crops in groundnut growing areas in combination with nematicide on managing *M. arenaria*.

Materials and methods

The experiment was carried out during *kharif* 2002-03-04 and 2005 at farmers' fields located in Junagadh district of Gujarat which were severely infested with *M. arenaria*. Groundnut variety GG-20 was sown with a spacing of 60 cm and three relay crops *viz.*, castor (GCH-4), pigeon pea (BDN-2), and sesame (G Til-1) were sown with a row ratio of 2:1 on different dates using recommended seed rate. Carbofuran 3G @ 1 kg a.i./ha was applied in furrows at the time of sowing. This experiment comprising eight treatments was conducted in a Randomized Block Design with three replications. Initial nematode population/200 cm³ soil was recorded at sowing and final nematode population/200 cm³ soil and root-knot index were recorded at 15 days before harvest.

Results and discussion

The initial nematode population (pooled) in the experimental field ranged from 272 to 399 (Table 1). The year-wise data and pooled data (2002-05) on the effect of intercrop on nematode population on groundnut crop indicated that the final nematode population was reduced by 52% when castor was sown as relay crop along with carbofuran followed by sesame and pigeon pea as relay crops (Table 1). The root-knot index was also significantly lowest (1.7) in the treatment where castor was sown as relay crop along with carbofuran followed by pigeon pea along with carbofuran as compared to other treatments (Table 1). Treatment of castor as relay crop along with carbofuran provided significantly higher pod yield followed by pigeon pea as relay crop with carbofuran which did not give any significant difference (Table 1). The highest net realization was obtained in groundnut + castor + carbofuran treatment followed by groundnut + pigeon pea + carbofuran with CBR of 1:2.35 and 1: 2.14, respectively.

Therefore, it is concluded that relay cropping of groundnut with castor (row ratio of 2:1) along with soil application of carbofuran 3G @ 1 kg a.i./ha was highly effective in managing nematode population and the root knot

nematode disease and also increasing the yield (CBR 1:2.35). The present investigations are in agreement with the earlier reports that better plant growth and reduction in root knot index and nematode population in tobacco intercropped with castor and phenamiphos (Dhanger *et al.*, 2002). Gowda *et al.*, (1995) also reported that castor caused maximum trapping of root knot nematodes. Control of *M. incognita* through cropping pattern was also

reported by Castillo *et al.*, (1975). Intercropping with marigold resulted in decrease in the number of plant parasitic nematodes on chilli, eggplant, cabbage, cauliflower and tomato (Alam *et al.*, 1977).

It is therefore apparent that growing castor as relay crop followed by application of carbofuran 3G @ 1 kg a.i./ha is more effective, economical and eco-friendly as compared to other treatments.

Table 1 Effect of different relay crops/nematicide on nematode population, root knot index and groundnut pod equivalent yield

Treatment	Initial nematode population/200cm ² soil (Pooled mean of 2002- 05)	Final nematode population/ 200cm ² soil (Pooled mean of 2002- 05)	Root knot index (1- 5) (Pooled mean of 2002- 05)	Pod equivalent yield (kg/ha) (Pooled mean of 2002- 05)	C B R
G'nut + Castor + Carbofuran @ 1 kg a.i./ha as furrow	272	131	1.72	2374	1:2.35
G'nut + Castor	308	315	2.81	1881	1:2.10
G'nut + Sesame+ Carbofuran @ 1 kg a.i./ha as furrow	288	159	2.84	1652	1:1.92
G'nut +Sesame	335	434	3.68	1279	1:1.76
G'nut + Pigeon pea+ Carbofuran @ 1 kg a.i./ha as furrow	292	179	2.54	2212	1:2.14
G'nut + Pigeon pea	346	432	3.53	1899	1:2.15
G'nut + Carbofuran @ 1 kg a.i./ha as furrow	310	213	2.88	1856	1:1.89
G'nut alone	399	571	4.52	1485	1:1.74
SEm ±	25.6	20.8	0.19	149.7	
CD (P=0.05)	75.5	61.2	0.57	440.4	
CV (%)	5.2	10.3	16.2	7.6	
Pooled:					
Year SEm ±	18.1	14.7	0.1	105.8	
CD (P=0.05)	53.4	43.3	NS	311.4	
Y x T SEm ±	9.5	18.1	0.3	80.2	
CD (P=0.05)	27.1	51.4	0.8	227.2	

G'nut = Groundnut

References

- Alam, M.M., Saxena, S.K. and Khan, A.M. 1977. Influence of interculture of marigold and margosa with some vegetable crops on plant growth and nematode population. *Acta Botanica Indica*, 5 : 33-39.
- Castillo, M.B., Alejar, M.S. and Harwood, R.R. 1975. Nematode in cropping patterns. II. Control of *Meloidogyne incognita* through cropping patterns and cultural practices. *Philippine Agriculturist*, 59: 295-312.
- Dhanger, D.S., Gupta, D.C. and Jain, R.K. 2002. Studies on intercropping of marigold (*Tagetes* sp.) with brinjal on plant growth and population of root-knot nematode, *M. javanica*. *Indian Journal of Nematology*, 32 : 220-221.
- Gowda, D. Nanje and Surayanarayana Reddy, V. 1995. Effect of castor and phenamiphos in management of root knot nematode in tobacco nursery. *Indian Journal of Nematology*, 25 : 221-222.
- Kalaiarasan, P., Rajendran, G. and Lakshmanan, P.L. 2006. Pathogenic potential of root-knot nematode, *M. arenaria* on groundnut (*Arachis hypogaea* L.), *Indian Journal of Nematology*, 36 : 278-279.
- Patel, H.R., Vaishnav, M.U. and Dhruj, I.U. 1985. Efficacy of aldicarb sulfone and carbofuran (flowable) seed treatments on plant growth against *Meloidogyne arenaria* on groundnut. *Pesticides*, 20: 29-31.

Productivity potentials and profitability of non-monetary, low-cost and cost-effective oilseeds production technologies

R. Venkattakumar, S.V. Ramana Rao, M. Padmaiah and D.M. Hegde

Directorate of Oilseeds Research, Rajendranagar, Hyderabad-500 030, AP

(Received: August, 2008; Revised: December, 2009; Accepted: December, 2009)

Abstract

Annual oilseeds support the livelihood earnings of 14 million farmers, majority of them are small and marginal in the arid and semi-arid eco-systems of the country, while one million are involved in processing of oilseeds and oils. The annual oilseed production of the country is faced with high degree of variation, as nearly 76% of the oilseeds area is under rainfed conditions and therefore subjected to uncertainties of moisture availability. Low relative comparative advantage, lower productivity and rainfed prone nature of oilseeds crops and the alarming increase in domestic oil demand, thrust upon the delineation of non-monetary, low-cost and cost-effective oilseeds production technologies for popularization among the oilseed growers. The delineation was done through ex-post-facto analysis on secondary data pertaining to frontline demonstrations (FLDs) in annual oilseed crops conducted under Technology Mission on Oilseeds (TMOP) and Integrated Scheme on Oilseeds, Pulses, Oilpalm and Maize (ISOPOM). The non-monetary technologies demonstrated through frontline demonstrations, resulted in seed yield increase, ranging from 19 to 50% whereas, the low-cost technologies from 22 to 24%. The demonstration of cost-effective technologies resulted in seed yield increase ranging from 9 to 63%. In the present scenario of oilseeds cultivation in the country, popularization of these proven non-monetary, low-cost and cost-effective oilseeds production technologies to the resource poor oilseed growers, will help them to get remunerative and sustainable yield as well as economic returns.

Key words: Non-monetary, low-cost and cost-effective oilseed technologies, productivity potentials and profitability

Introduction

Nine annual oilseeds crops are being cultivated in 26.51 million ha in India, with the production of 24.29 million tonnes and the productivity of 916 kg/ha (2006-07). India accounts for 15.6, 9.2, 6.8, 5.9, 6.1, 9.0 and 9.3% of

world's oilseed area, production, oil meal production, oil meal export, vegetable oil export, vegetable oil import and edible oil consumption. About 12-13% of dietary energy to human beings is supplied by oilseeds. Among the annual oilseeds grown in the country, groundnut, rapeseed-mustard and soybean account for nearly 78% of the oilseed area and 87% of the production. Madhya Pradesh, Gujarat, Rajasthan, Andhra Pradesh, Maharashtra, Karnataka, Tamil Nadu and Uttar Pradesh account for nearly 90 and 92% of oilseeds area and production of the country (Hegde, 2007).

Rationale behind delineation of non-monetary, low-cost and cost-effective oilseeds production technologies

- Annual oilseeds support the livelihood earnings of small and marginal farmers of arid and semi-arid eco-systems of the country. It is estimated that 14 million farmers are involved in oilseed cultivation, while one million persons are involved in processing of oilseeds and oils (Hegde and Venkattakumar, 2007).

The annual oilseed production of the country is faced with high degree of variation, as nearly 76% of the oilseeds area is under rainfed conditions and therefore subjected to uncertainties of moisture availability.

The country's minimum support price (MSP) programme that has often favoured production of crops that compete with oilseeds for area. Hence, only by popularizing non-monetary, low-cost and cost-effective oilseeds production technologies among the resource-poor farmers, oilseeds can sustain their competition with the competing crops.

The continuous cultivation of oilseed crops without proper crop rotation has led to depletion of soil nutrients as well as increase in pest and disease incidence causing upto 40% yield loss (Gowda *et al.*, 2007). This implies the need for use of cost-effective oilseeds-based cropping sequences and intercropping systems.

Oilseed crops are prone to damage by more than 64 major diseases. For as many as 26 pests in different

crops, no resistant source is available, vulnerability of majority of the cultivars of oilseed crops to insect pests and diseases continues to be one of the major factors responsible for the lower productivity and wider fluctuations in production. (Rabindra *et al.*, 2007). Thus, the need for use of low-cost and cost-effective integrated pest management practices in oilseeds cultivation is significantly important.

Specific attention needs to be given to harness the residual effects of fertilizers containing P, K and S. Sound fertilizer management for intercropping systems involving oilseeds which can meet the nutrient needs of both main and intercrop will go a long way in enhancing the productivity of the system.

Population growth, rising standard of living, aberrant weather for few years in succession and liberalization of export-import policy are the causes for rapid surge in imports (Rao, 2007). The relative advantage is less in case of oilseeds and oils than other food crops. Over the last one decade, the irrigation coverage merely increased by 4% from 23 to 27%.

To obtain the realistic assessment of potential of newly generated oilseed technologies through FLDs, the following thrust areas need to be emphasized while implementing (Rai, 2002): Cropping system demonstrations *viz.* relay, sequential and intercropping systems; maximizing returns per unit area, time and input; cluster area approach; narrowing down the various levels of gaps through efficient resource use management and effective integration of all the research and development agencies that are involved in the development of a particular oilseed crop in a given locality.

As per the data pertaining to the FLDs conducted during 1996-97 to 2006-07 and the national average yield of oilseeds for 2006-07, it was estimated that 227.76 lakh tonnes of additional production could be achieved, apart from 238.84 lakh tonnes of actual production (total could be 466.60 lakh tonnes), if the reservoir of oilseed production projected through FLDs, was utilized through complete adoption of improved technologies advocated for oilseed cultivation. The yield gap was 208% for groundnut and 44, 196, 110, 58, 89, 186, 100 and 148% for rapeseed-mustard, sunflower, safflower, soybean, sesame, niger, castor and linseed respectively. The above-mentioned data suggests the need for popularization of improved oilseed production technologies for use by the oilseed growers.

There is a potential to increase production of oilseeds by using best production practices and right combination of inputs at right time (Kumar and Chauhan, 2007).

The recent demand projections made by Rabo bank

for Department of Agriculture and Cooperation (DAC) indicated that the edible oil demand might rise up to 14.8, 18.3 and 21.8 million tones by 2010, 2015 and 2020 respectively. This means, the present level of oilseed production of the country needs to be increased by three times to meet out the projected demand for edible oil for which popularization and use of improved oilseed production technologies is a pre-requisite.

The above points reinforce the necessity for delineation of non-monetary, low-cost and cost-effective oilseeds production technologies and in turn effective transfer of these technologies to the oilseed growers for adoption under real farm situations.

Materials and methods

TMOP and ISOPOM: The Technology Mission on Oilseeds (TMO) launched by Government of India (GOI) in 1986, had a significant impact on overall production of oilseeds. To meet out the challenges that posed through huge demand for vegetable oils in the country, the DAC started implementing Integrated Scheme on Oilseeds, Pulses, Oilpalm and Maize (ISOPOM) to promote crop diversification and to provide focused approach to the oilseed development programmes from April 2004 onwards. In both the schemes, FLDs in oilseeds have been a major mode of transfer of technology effort to prove the improved oilseed production technology under real farm conditions. The delineation of non-monetary, low-cost and cost-effective oilseeds production technologies was done through ex-post-facto analysis on secondary data FLDs in oilseeds conducted. For this purpose the Annual reports on 'Frontline Demonstrations of Oilseeds' for the period of 1996-97 to 2006-07 were utilized (DOR, 2006a, DOR, 2006b, DOR, 2006c and DOR, 2007).

Operationalization of concepts used in the study

Non-monetary oilseed production technologies: The technologies classified under this category need right decision-making (psychomotor) behaviour of the oilseeds growers for adoption. The intelligence on oilseeds production accumulated based on the past experience and awareness, knowledge on improved technological recommendations, is needed to be utilized by the farmers in adopting such oilseeds technologies.

Low-cost oilseed production technologies: The oilseed production technologies with low and minimum cost to be spent apart from the usual cost of cultivation, but results in considerable additional yield and economic benefits are included under this category.

Cost-effective production technologies: The oilseeds production technologies to be adopted by spending considerable cost, but result in sustainable yield and economic returns of the oilseed crops, are operationalized as cost-effective technologies.

Results and discussion

Impact of non-monetary oilseed production technologies

Adoption of recommended spacing: Adoption of recommended spacing between plants and rows, resulted in 5 and 15% seed yield increase under irrigated and rainfed conditions of castor respectively, with corresponding additional net returns of Rs.268 and 1845/ha (Table 1). The same technology in linseed, gave 65% seed yield increase and additional net returns of Rs.2075/ha. In sunflower, the technology gave around 25% seed yield increase under both irrigated and rainfed conditions, whereas, the corresponding additional net returns was Rs.3733 and 4760/ha. Under rainfed conditions, adoption of recommended spacing resulted in 11 and 25% seed/ pod yield increase for sesame and groundnut resulting in additional net returns of Rs.1551 and 5923/ha respectively. As a whole for oilseeds, the technology gave 19% seed yield increase and Rs. 2853/ha additional net returns. The B:C ratio was 2.8 and 2.5 with IT and FP plots respectively (Table 4).

Adoption of right method of sowing: Adoption of the recommended method of sowing in sunflower gave 51% seed yield increase under irrigated conditions, whereas 25% under rainfed conditions. The additional net returns obtained was Rs.5254 and 3435/ha respectively for irrigated and rainfed conditions (Table 1). The seed/ pod yield increase as a result of adoption of this technology over the local practice was four times in irrigated than rainfed conditions in safflower and 2.5 times in groundnut. The seed yield increase was 90 and 76% for sesame and niger under rainfed conditions with corresponding additional net returns of Rs. 1408 and 2184/ha. For rapeseed-mustard, the seed yield increase was 36% with Rs.4072/ha additional net returns under irrigated conditions. As an overall impact, the seed yield increase was observed as 39% with Rs.3286/ha additional net returns. The B:C ratio was 2.4 and 2.1 with IT and FP plots respectively (Table 4).

Adoption of timely sowing: Adoption of timely of sowing resulted in 44 and 55% seed yield increase for sunflower and rapeseed-mustard respectively under irrigated conditions, with accrued additional net returns of Rs.3997 and 1061/ha respectively (Table 1). Under rainfed conditions, the same technology resulted in 182 and 13% seed/ pod yield increase for niger and groundnut with Rs.3150 and 2889/ha additional net returns. The overall impact was 36% for seed yield increase and Rs.4190/ha for additional net returns (Table 4). The B:C ratio was 2.5 and 2.3 with IT and FP plots respectively.

Adoption of right method of fertilizer application: Adoption of right method of fertilizer application, under rainfed conditions, gave 42% seed yield increase with Rs.2771/ ha additional net returns for rapeseed-mustard.

The B:C ratio was 3.6 and 2.5 with IT and FP plots respectively (Table 1 and 5).

Adoption of timely weeding: There was 39% seed yield increase in rapeseed-mustard under rainfed conditions (Table 1 and 5), when timely weeding was adopted under real farm conditions.

Selecting oilseed crops to competing crops: Selecting castor instead of maize, cotton and sorghum gave 104, 87 and 5% seed yield increase under rainfed conditions (Table 1). The corresponding additional net returns was 3121, 8237 and 5674/ha. Though tomato gave 21% seed yield equivalent increase than castor, there was Rs.675/ha additional net returns from castor, since the labour requirement of castor is lower than that of tomato. Under irrigated conditions, sunflower was proved to be superior to finger millet by 86% in seed yield equivalent and Rs.12340/ha in net returns. As a whole, the above-mentioned oilseed crops proved 50% seed yield equivalent increase and Rs.6009/ha additional net returns to competing crops. The B:C ratio was 3.0 and 2.1 with IT and FP plots respectively (Table 4).

Adoption of improved varieties and hybrids of oilseeds: Choice of improved varieties of groundnut to locals and existing ones, gave 25% pod yield increase both under rainfed and irrigated conditions. In rapeseed-mustard, the technology gave seed yield increase to the tune of 58% under irrigated conditions with Rs.4171/ha additional net returns. Improved dual-purpose linseed variety gave 68% seed yield enhancement and Rs.15453/ha additional net returns. Improved niger variety resulted in 120% seed yield increase and Rs.2383/ha additional net returns. Similarly, improved varieties gave seed yield increase to the tune of 27 and 45% under rainfed conditions in soybean and castor respectively, with corresponding additional net returns of Rs.2960 and 5340/ha (Table 1). Overall, improved varieties/ hybrids of oilseeds resulted in 36% seed yield increase, with Rs.3645/ha additional net returns. The B:C ratio was 2.4 and 2.1 with IT and FP plots respectively (Table 4).

Adoption of recommended crop sequence: Adoption of recommended crop sequence resulted in 83 and 62% seed yield increase in linseed under rainfed and irrigated conditions respectively (Table 1). The additional net returns obtained was Rs.4184 and Rs.3443/ha, under rainfed and irrigated conditions respectively. In castor, it resulted in 33% seed yield increase with Rs. 3900/ha additional net returns under rainfed conditions and 11% seed yield increase and Rs. 2004/ ha additional net returns, under irrigated conditions. In case of rapeseed-mustard, the seed yield increase as result of adoption of recommended crop sequence was 46 and 87% in irrigated and rainfed conditions respectively, with corresponding additional net returns of Rs.4955 and 6749/ha. In sunflower, under irrigated conditions, the technology resulted in 19% seed yield increase with

Rs.3491/ha additional net returns. The technology as a whole, gave 37% seed yield increase and Rs.5896/ha additional net returns. The B:C ratio was 2.0 and 1.8 with IT and FP plots respectively (Table 4).

Impact of low-cost oilseed production technologies

Adoption of seed treatment practice: Adoption of seed treatment with chemicals and/or biofertilizers for pest management and nutrient supplement, resulted in 10% seed yield increase both under rainfed and irrigated conditions in sunflower, with additional net returns of Rs.1525/ha. (Table 2). In safflower, the technology gave 12% seed yield increase and Rs.413/ha additional net returns under rainfed conditions. Under similar conditions in rapeseed-mustard, it gave 24% seed yield increase with Rs.2550/ha additional net returns. In groundnut, under irrigated conditions, this technology gave 44% pod yield increase and Rs.1957/ha additional net returns. This technology gave 24% seed yield increase and Rs.1083/ha additional net returns as an overall impact (Table 4). The B:C ratio was 2.3 and 2.6 with IT and FP plots respectively.

Adoption of thinning: Thinning excess plant population to maintain the recommended plant stand, resulted in seed yield increase to the tune of 26 and 32% in sunflower and safflower under irrigated conditions, with corresponding additional net returns of Rs.3576 and 4624/ha respectively (Table 2). In rapeseed-mustard under irrigated conditions, this technology gave 16% seed yield increase, with Rs.2258/ha additional net returns. Overall, this technology gave 24% seed yield increase and Rs.3238/ha additional net returns (Table 4). The B:C ratio was 2.6 and 2.3 with IT and FP plots respectively.

Use of biofertilizers: Application of biofertilizers, as a nutrient supplement, resulted in 9, 24 and 40% seed yield increase in sunflower, safflower and niger with Rs.1101, 1871 and 537/ha additional net returns respectively (Table 2). Under irrigated conditions, this technology gave seed/pod yield increase to the tune of 44% for groundnut and 6% for rapeseed-mustard with corresponding additional net returns of Rs.1957 and 1883/ha. Overall, this technology gave 22% seed yield increase with Rs.1925/ha additional net returns. The B:C ratio was 2.2 and 2.1 with IT and FP plots respectively (Table 4).

Application of gypsum: Application of gypsum in groundnut to increase pod yield and oil content of the crop and physio-chemical properties of the soil, resulted in 25% pod yield increase and Rs.342/ha additional net returns under rainfed conditions (Table 2 and 5). The B:C ratio was 1.5 and 1.6 with IT and FP plots respectively.

Application of sulphur: Application of sulphur to increase oil content in rapeseed-mustard resulted in 11% seed yield increase and Rs.3277/ha additional net returns under rainfed conditions (Table 2 and 5). The B:C ratio was 2.6 and 2.4 with IT and FP plots respectively.

Spraying of cycocel: Spraying cycocel as a growth regulator for safflower resulted in seed yield increase to the tune of 33% under irrigated conditions with Rs.5030/ha additional net returns (Table 2). Overall, the seed yield increase was 24% and the additional net returns was Rs.2778/ha (Table 4). The B:C ratio was 2.4 and 2.2 with IT and FP plots respectively.

Application of sulphur+boron: Sulphur+boron application to improve the oil content in sunflower resulted in 34% seed yield increase and Rs.8358/ha additional net returns under rainfed conditions (Table 2 and 5).

Impact of cost-effective oilseed production technologies

Integrated nutrient management (INM): Adoption of INM practices to improve the productivity of the crop as well as soil fertility in a sustained manner resulted in seed/pod yield increase to the tune of 22 and 19% in groundnut and rapeseed-mustard under irrigated conditions. The corresponding additional net returns was Rs.4806/ha for groundnut (Table 3). Overall, the technology gave 20% seed yield increase and Rs.1577/ha additional net returns (Table 4). The B:C ratio was 2.2 and 2.4 with IT and FP plots respectively.

Integrated weed management (IWM): IWM practices to manage the weeds in a sustained and cost-effective manner resulted in pod yield increase of 10% in groundnut under irrigated conditions. The corresponding additional net returns was Rs.3984/ha (Table 3 and 5). The B:C ratio was 2.6 and 2.4 with IT and FP plots respectively.

Integrated water management: Integrated water management practices to meet the minimal water requirement of the crop with higher use efficiency and ensuring the sustained availability of water to the crop eco-system gave 9% pod yield increase and Rs.2481/ha additional net returns in groundnut (Table 3 and 5). The B:C ratio was 2.3 and 2.2 with IT and FP plots respectively.

Integrated pest management (IPM): IPM practices to ensure the arrest of resilience of insect pests of crops through integrated management practices and ecological balance in pest management in groundnut gave pod yield increase to the tune of 18% with Rs. 3354/ha additional net returns under rainfed conditions, whereas, 19% seed yield increase for rapeseed-mustard under irrigated conditions (Table 3). Overall, the seed yield increase was 17% and additional net returns was Rs.35771/ha (Table 4). The B:C ratio was 2.1 and 2.0 with IT and FP plots respectively.

Polythene mulching: Polythene mulching for arresting the weed population and conserving the soil moisture by preventing evaporation losses in groundnut gave pod yield increase to the tune of 51% with Rs.15643/ha additional net returns (Table 3 and 5). The B:C ratio was 2.6 and 2.2 with IT and FP plots respectively.

Table 1 Impact of non-monetary oilseeds production technologies

Crop	Situation	No of demos.	Mean seed yield (kg/ha)		% increase in yield	Cost of cultivation (Rs./ha)		Gross returns (Rs./ha)		Addl.net returns (Rs./ha)	B.C ratio	
			IT	FP		IT	FP	IT	FP		IT	FP
Adoption of Recommended Spacing												
Castor	I	3	2332	2213	5	7056	7355	29375	29407	268	4.0	4.0
	R	6	1696	1474	15	6474	5999	22696	20377	1845	3.8	3.4
Linseed	I	2	576	350	65	3855	3218	6912	4200	2075	2.1	1.3
	I	22	1464	1173	25	10194	9556	22170	17799	3733	2.3	1.9
Sunflower	R	31	1345	1067	26	6710	6253	19661	14444	4760	3.1	2.3
Sesame	R	1	427	385	11	2972	2517	8855	6849	1551	3.5	2.7
Groundnut	R	9	1965	1575	25	12527	11250	31448	24428	5923	2.8	2.2
Adoption of Right Method of Sowing												
Sunflower	I	2	1378	913	51	5470	4344	17379	10999	5254	4.0	2.5
	R	5	1108	885	25	5604	5370	16053	12396	3435	3.0	2.3
Safflower	I	1	2500	2025	23	5505	6415	35000	28350	7560	5.5	4.4
	R	5	806	762	6	6204	4641	8618	7914	-859	1.9	1.7
Sesame	R	2	275	145	90	1960	1448	3880	1960	1408	2.7	1.4
	I	1	2625	2000	31	18062	16340	34725	26687	6316	2.1	1.6
Groundnut	R	5	550	492	12	7256	6479	9355	8361	217	1.4	1.3
Rapeseed-mustard	I	9	1389	1023	36	9896	8186	22430	16647	4072	2.7	2.0
Niger	R	42	402	228	76	3393	2156	6992	3571	2184	3.2	1.7
Following Timely Sowing												
Sunflower	I	2	1015	703	44	8100	8066	13219	9188	3997	1.6	1.1
Niger	R	104	316	112	182	3440	3017	5389	1817	3150	1.8	0.6
Rapeseed-mustard	I	33	1559	1009	55	6193	5723	20495	13302	1061	3.6	2.3
Groundnut	R	4	1938	1719	13	11436	10950	34137	30762	2889	3.1	2.8
Adoption of Right Method of Fertilizer Application												
Rapeseed-mustard	R	31	936	661	42	4723	3541	12727	8774	2771	3.6	2.5
Adoption of Timely Weeding												
Rapeseed-mustard	R	3	1635	1175	39	-	-	-	-	-	-	-
Selecting Oilseeds to Competing Crops**												
Castor/maize	R	4	2157	1060	104	6608	6013	18404	14688	3121	2.8	2.4
Sunflower/ finger millet	I	10	2260	1215	86	13873	10149	34754	18690	12340	2.5	1.8
Castor/ cotton	R	2	2338	1250	87	6739	8882	31782	25688	8237	4.7	2.9
Castor/ sorghum	R	19	1998	1900	5	5489	5036	14306	8179	5674	2.6	1.6
Castor/ tomato	R	2	688	-	-21	3827	8257	9369	13125	675	2.4	1.6
Adoption of Improved Varieties/ Hybrids												
Groundnut	I	213	2483	1976	26	16911	15255	40564	31973	6934	2.4	2.1
	R	1084	1839	1476	25	15310	13955	34596	27847	5393	2.3	2.0
Rapeseed-mustard	R	71	967	697	39	6187	5704	13120	9009	3628	2.1	1.6
	I	27	555	351	58	8859	6528	16070	9567	4171	1.8	1.5
Sesame	R	136	515	367	40	6440	5344	16431	11414	3920	2.6	2.1
	DP	3	1567	933	68	10470	8456	3350	15583	15453	3.2	1.8
Linseed	I	41	1160	848	37	10852	9269	21400	15794	4022	2.0	1.7
	R	25	634	420	51	4802	4196	12212	8456	3150	2.5	2.0
Niger	R	81	352	160	120	3527	1778	7188	3057	2383	2.0	1.7
	I	9	460	408	13	2477	2440	19929	17158	2734	8.0	7.0
Soybean	R	36	756	594	27	6943	6422	13556	10076	2960	2.0	1.6
	I	105	2618	1970	33	11262	11838	35384	21425	14535	3.1	1.8
Castor	R	208	1393	961	45	8186	6794	27637	20906	5340	3.4	3.1
	I	12	1386	783	77	6969	5032	20132	11006	7189	2.9	2.2
Safflower	R	209	922	719	29	6205	5427	13423	10301	2344	2.2	1.9
	I	69	1609	1331	21	8771	8373	25426	21023	4004	2.9	2.5
Sunflower	R	93	1193	985	21	7469	7389	20294	16708	3506	2.7	2.3
Adoption of Recommended Crop Sequence												
Linseed	R	1	1434	785	83	13400	5870	21527	9813	4184	3.7	1.7
	I	5	1014	625	62	7542	5879	13298	8193	3443	2.3	1.4
Castor	R	1	1600	1200	33	7910	7410	17600	13200	3900	2.4	1.8
	I	6	2677	2419	11	11586	10307	35178	31896	2004	3.4	3.1
Rapeseed-mustard	I	93	1959	1339	46	8996	5713	24725	16488	4955	4.3	2.9
	R	8	1323	707	87	7558	5879	17844	9416	6749	3.0	1.6
Safflower	R	6	1462	1161	26	13160	10451	5938	3434	-205	0.6	0.3
Sunflower	I	11	1446	1213	19	7052	7048	18895	15399	3491	2.7	2.2

IT= Improved technology; FP=Farmers' practice; I=Irrigated; R=Rainfed; DP=Dual purpose;

*-The data pertaining to timely weeding, selecting oilseeds crops to competing crops and selection of improved varieties/ hybrids were of the period from 2002-03 to 2006-07, whereas the remaining data were of the period from 1996-07 to 2006-07; **-expressed in seed yield equivalent

Table 2 Impact of low-cost oilseeds production technologies*

Crop	Situation	No. of demos.	Mean seed yield (kg/ha)		% increase in yield	Cost of cultivation (Rs./ha)		Gross returns (Rs./ha)		Addl net returns (Rs./ha)	B:C ratio	
			IT	FP		IT	FP	IT	FP		IT	FP
Seed Treatment												
Sunflower	I	4	1187	1079	10	4313	4088	19000	17250	1525	4.6	4.2
Safflower	R	2	811	725	12	6665	4800	14603	12325	413	3.0	2.6
Groundnut	I	3	2333	1625	44	20558	14799	33533	25817	1957	2.3	1.7
Rapeseed-mustard	R	6	968	778	24	3400	3198	14031	11279	2550	4.4	3.5
Thinning												
Sunflower	I	25	1303	1034	26	8282	7692	19938	15766	3576	2.6	2.0
	R	52	1131	945	20	7357	6919	20277	17209	2629	2.9	2.5
Safflower	I	32	1445	1093	32	7807	7046	21118	15733	4624	3.0	2.2
Rapeseed-mustard	R	108	1170	908	29	6402	5855	16219	12573	3099	2.8	2.1
	I	25	1555	1336	16	7865	7130	21770	18778	2258	3.1	2.6
Biofertilizer												
Sunflower	R	8	856	783	9	4425	4257	14974	13705	1101	3.5	3.2
Safflower	R	7	867	697	24	4295	3750	11568	9152	1871	3.1	2.4
Niger	R	6	208	149	40	1758	1399	2877	1982	537	2.1	1.4
Groundnut	I	3	2333	1625	44	20558	14799	33533	25817	1957	2.3	1.7
Rapeseed-mustard	R	110	1790	1500	19	14926	13108	34591	28570	4203	2.6	2.2
	I	8	1787	1681	6	15026	15016	35655	33762	1883	2.4	2.2
Gypsum Application												
Groundnut	R	3	1077	865	25	7691	5788	11405	9160	342	1.5	1.6
Sulphur Application												
Rapeseed-mustard	R	37	1710	1535	11	11496	10962	29767	25956	3277	2.6	2.4
Cycocel Spray												
Safflower	I	2	1380	1038	33	6396	5433	24150	18157	5030	3.8	3.3
	R	11	1063	884	20	7933	7138	16387	13565	2027	2.1	1.9
Sulphur+Borax												
Sunflower	R	10	1441	1073	34	5724	5304	25938	17160	8358	4.5	3.2

IT= Improved technology; FP=Farmers' practice; I=Irrigated; R=Rained; *.The data were of the period from 1996-07 to 2006-07

Table 3 Impact of cost-effective oilseeds production technologies*

Crop	Situation	No. of demos.	Mean seed yield (kg/ha)		% increase in yield	Cost of cultivation (Rs./ha)		Gross returns (Rs./ha)		Addl net returns (Rs./ha)	B:C ratio	
			IT	FP		IT	FP	IT	FP		IT	FP
Integrated Nutrient Management												
Groundnut	I	116	2424	1995	22	17770	15933	40387	33744	4806	2.3	2.1
	R	224	1579	1334	18	13281	11691	30113	24973	3550	2.3	2.1
Rapeseed-mustard	I	5	1737	1470	19	15500	7930	30870	26926	-3626	2.0	3.4
Integrated Weed Management												
Groundnut	R	14	2216	2023	10	16531	16684	43734	39903	3984	2.6	2.4
Integrated Water Management												
Groundnut	I	23	1588	1461	9	15105	14913	35024	32350	2481	2.3	2.2
Integrated Pest Management												
Groundnut	I	42	2439	2138	14	18340	16774	42732	37367	3838	2.3	2.2
	R	106	1632	1380	18	16576	14681	32073	26824	3354	1.9	1.8
Rapeseed-mustard	I	2	1768	1490	19	-	-	-	-	-	-	-
Integrated Disease Management												
Groundnut	I	30	1528	1135	35	13690	11206	32047	23765	5798	2.3	2.1
	R	119	1677	1354	24	12257	9848	27805	22161	3236	2.3	2.3
Rapeseed-mustard	I	8	1714	1575	9	12688	12375	29137	26912	1913	2.3	2.2
Oilseed-based Intercropping Systems												
Linseed	I	90	1342	638	110	8945	6277	24399	13131	8600	2.7	2.1
	R	53	1048	566	85	7513	5303	19926	9622	8694	2.7	1.8
Sesame	I	7	549	382	44	3624	2861	17651	11485	5402	4.9	4.0
	R	73	559	362	54	7292	5468	16344	10746	3774	2.2	2.0
Niger	R	22	554	234	137	4898	2694	9765	4050	3511	2.0	1.5
Soybean	R	27	1691	1127	50	7497	5358	21905	14801	4966	2.9	2.8
Castor	R	40	1789	900	99	8429	6186	29480	16791	10446	3.5	2.7
	I	38	2600	1725	51	16101	11318	53440	37162	11495	3.3	3.3
Safflower	R	119	1094	792	38	6208	5593	15230	10805	3810	2.5	1.9
Sunflower	R	61	975	780	25	7992	7382	17698	14122	2966	2.2	1.9
Polythene Mulching												
Groundnut	I	11	2660	1785	49	18326	14881	47878	32518	11915	2.6	2.2
	R	13	3032	2005	51	20184	16870	56684	37728	15643	2.8	2.2
Thio urea Application												
Rapeseed-mustard	I	18	1808	1644	10	10677	9944	27551	25146	1673	2.6	2.5

IT=Improved technology; FP=Farmers' practices; I=Irrigated; R=Rained; *.The were of the period from 2002-03 to 2006-07

Table 4 Impact of non-monetary, low-cost and cost-effective oilseeds production technologies

Technology	No. of demos.	Mean seed yield (kg/ha)		% increase in yield	Cost of cultivation (Rs./ha)		Gross returns (Rs./ha)		Addl net returns (Rs./ha)	B:C ratio	
		IT	FP		IT	FP	IT	FP		IT	FP
Non-monetary oilseeds production technologies											
Selecting oilseeds to competing crops*	37	1888	1260	50	7307	7667	21723	16074	6009	3.0	2.1
Choice of improved varieties*	2422	1201	881	36	8332	7306	20042	15371	3645	2.4	2.1
Recommended Spacing	74	1401	1177	19	7113	6593	20160	16786	2853	2.8	2.5
Correct method of sowing	72	1226	941	39	7039	6153	17159	12987	3286	2.4	2.1
Correct time of sowing	143	1207	886	36	7292	6939	18310	13767	4190	2.5	2.3
Correct method of fertilizer application	31	936	661	42	4723	3541	12727	8774	2771	3.6	2.5
Timely weeding*	3	1635	1175	39	-	-	-	-	-	-	-
Adoption of Recommended crop sequence	131	1614	1181	37	9651	7320	19376	13480	5896	2.0	1.8
Low-cost production technologies											
Seed treatment	17	1228	994	24	8168	6095	19049	15893	1083	2.3	2.6
Thinning	242	1321	1063.2	24	7543	6928	19864	16012	3238	2.6	2.3
Biofertilizer application	142	1307	1073	22	10165	8722	22200	18831	1925	2.2	2.1
Application of gypsum in groundnut	3	1077	865	25	7691	5788	11405	9160	342	1.5	1.6
Application of sulphur in rapeseed-mustard*	37	1710	1535	11	11496	10962	29767	25956	3277	2.6	2.4
Application of cycocel in safflower*	13	1142	923	24	7549	6712	18328	14713	2778	2.4	2.2
Application of Sulphur+Borax in sunflower*	10	1441	1073	34	5724	5304	25938	17160	8358	4.5	3.2
Cost-effective production technologies											
Integrated nutrient management*	345	1913	1600	20	15517	11851	33790	28548	1577	2.2	2.4
Integrated weed management*	14	2216	2023	10	16531	16684	43734	39903	3984	2.6	2.4
Integrated water management*	23	1588	1461	9	15105	14913	35024	32350	2481	2.3	2.2
Integrated pest management*	150	1946	1669	17	17458	15728	37403	32096	3577	2.1	2.0
Integrated disease management*	157	1640	1355	21	12878	11143	29663	24279	3648	2.3	2.2
Oilseed-based intercropping systems*	530	1220	751	63**	7850	5844	22584	14272	6306	2.9	2.4
Polythene mulching in groundnut*	11	2660	1785	49	18326	14881	47878	32518	11915	2.6	2.2
Application of thio-urea in rapeseed-mustard*	18	1808	1644	10	10677	9944	27551	25146	1673	2.6	2.5

*-The data pertaining to these technologies were of the period from 2002-03 to 2006-07, whereas the remaining data were of the period from 1996-07 to 2006-07; **-Seed equivalent yield

Application of thio-urea: Thio-urea application to improve the oil content of rapeseed-mustard resulted in 10% seed yield increase and Rs. 1675/ha additional net returns under irrigated conditions (Table 3 and 5). The B:C ratio was 2.6 and 2.5 with IT and FP plots respectively.

Oilseeds-based intercropping systems: Intercropping systems are recommended with a view to improve the yield and returns per unit area, while helping in efficient and effective use of available space as well as critical inputs of the crops. These practices also help as an insurance against crop failure. Use of linseed-based intercropping systems gave seed equivalent yield increase to the tune of 110%, with Rs. 8600/ha additional net returns (Table 3). Sesame-based intercropping systems gave seed equivalent yield increase to the tune of 54%, with Rs. 3774/ha additional net returns. Similarly, niger, soybean, safflower, sunflower and castor-based intercropping systems resulted in seed equivalent yield increase of 137, 50, 38, 25 and 99%, under rainfed conditions with corresponding additional net returns of Rs. 3511, 4966, 3810, 2966 and 10446/ha. Overall, the oilseeds-based intercropping systems gave 63% seed equivalent yield advantage, with Rs. 6306/ha additional net returns (Table 4). The B:C ratio was 2.9 and 2.4 with IT and FP plots respectively.

Conclusion

The non-monetary technologies that just need right decision making (psychomotor) behaviour of the oilseeds growers for adoption, demonstrated through frontline demonstrations, resulted in seed yield increase, ranging from 19 to 50% for adoption of recommended spacing and selecting oilseeds to competing crops respectively. The average cost of these non-monetary technologies was Rs. 7351/ha, with a range of Rs. 4723 to 9651/ha. The low-cost technologies to be adopted with minimal cost to be spent, which results in considerable additional yield and economic benefits, resulting in seed yield increase ranging from 22 to 24% for biofertilizer application to seed treatment respectively. The average cost of these low-cost technologies was Rs. 8334/ha, ranging from Rs. 5724 to 11496/ha. The demonstration of cost-effective technologies to be adopted by spending considerable cost, but result in sustainable yield and economic returns of the oilseed crops, resulted in seed yield increase ranging from 9 to 63% for integrated water management to oilseeds-based intercropping systems respectively. The cost of these cost-effective technologies was ranging from Rs. 7850 to 17458/ha, with an average of Rs. 14293/ha. In the present scenario of oilseeds cultivation in the country, popularization of these proven non-monetary, low-cost and cost-effective oilseeds production technologies among

resource poor oilseed growers, will help them to get remunerative and sustainable yield as well as economic returns, in the backdrop of import of huge quantity of edible oils in the globalized market economy and remote chances for acquiring additional area for cultivation of oilseeds from that of food grains.

References

- DOR. 2006a. *Frontline demonstrations in oilseeds-An overview, 1997-98 to 2001-02*. Directorate of Oilseeds Research, Rajendranagar, Hyderabad-500 030. P 184.
- DOR. 2006b. *Frontline demonstrations in oilseeds-Progress Report, 2002-03 to 2004-05*. Directorate of Oilseeds Research, Rajendranagar, Hyderabad-500 030. P 164.
- DOR. 2006c. *Frontline demonstrations in oilseeds-Annual Report, 2005-06*. Directorate of Oilseeds Research, Rajendranagar, Hyderabad-500 030. P 115.
- DOR. 2007. *Frontline demonstrations in oilseeds-Annual Report, 2006-07*. Directorate of Oilseeds Research, Rajendranagar, Hyderabad-500 030. P 137.
- Gowda, M.V.C., Lingaraju, S., Basappa, H. and Kusuma, V.P. 2007. Breeding for biotic stresses in oilseed crops: Current status and future threats. In: Hegde, D.M. 2007. *Changing global vegetable oil scenario: Issues and challenges before India*. Indian Society of Oilseeds Research, Hyderabad, pp.47-64.
- Hegde, D.M. 2007. An overview of oilseed scenario in India and scope and potential of bio-fuel crops. Paper presented in the *Expert Group Discussion on Role of Bio-fuels for Decentralized Application in Agriculture and Allied Activities*, May 14-15, 2007 at Administrative Staff College of India, Hyderabad.
- Hegde, D.M. and Venkattakumar, R. 2007. Development of oilseeds- a step towards self-sufficiency in India. *Souvenir*. All India Convention on Oilseeds, Oil trade and Industry. Nagpur. 27-28 October 2007, pp.41-48.
- Kumar Sant and Chauhan Sonia. 2007. Production performance and total factor productivity growth of oilseeds in India. In: Hegde, D.M. 2007. *Changing global vegetable oil scenario: Issues and challenges before India*. Indian Society of Oilseeds Research. Hyderabad, pp.239-246.
- Rabindra, R.J., Sunil Joshi and Veenakumari, K. 2007. Biological control of insect pests of oilseeds in India. In: Hegde, D.M. 2007. *Changing global vegetable oil scenario: Issues and challenges before India*. Indian Society of Oilseeds Research. Hyderabad, pp.101-142.
- Rai, S.D. 2002. Technology transfer by public sector. In: Rai, Mangala, Singh, Harvir and Hegde, D.M. (Eds). *Oilseeds and oils: Research and development needs*. Indian Society of Oilseeds Research. Hyderabad, pp.438-445.
- Rao, K.P.C. 2007. Can India achieve self-sufficiency in edible oils once again? In: *Souvenir*. National Seminar on changing global vegetable oils scenario: Issues and challenges before India. January 29-31, 2007. Indian Society of Oilseeds Research. Hyderabad, pp.9-15.

Short communication

Genetic divergence in groundnut, *Arachis hypogaea* L.

V.P. Korat, R.H. Kavani, M.S. Pithia, J.J. Savaliya and A.G. Pansuriya

Department of Agricultural Botany, Junagadh Agricultural University, Junagadh-362 001, Gujarat

(Received: November, 2008; Revised: October, 2009; Accepted: October, 2009)

Abstract

D²-statistics, grouped 80 genotypes of groundnut into 9 clusters. The clustering pattern of the genotypes did not confirm to the geographical distribution. The maximum intercluster distance was observed between clusters I and VIII followed by clusters IV and VIII, clusters III and VIII and clusters II and VIII. The cluster VII showed high mean in respect to pod yield/plant, number of secondary branches/plant, number of aerial pegs/plant, number of kernels/pod, 100-kernel weight and harvest index. The cluster I had desirable value for days to 50% flowering and days to maturity. While higher number of primary branches was found in cluster V. The cluster VII was the best for plant height and biological yield/plant. The cluster IV and III had desirable values for shelling percentage and oil content, respectively. The cluster IX was the best for number of underground pegs/plant. It will be advisable to intercross among the genotypes from clusters I, II, III, IV and VIII for generation of transgressive segregants and wide spectrum genetic variability for improvement of pod yield in groundnut.

Keywords: D² statistics, groundnut, diversity

In India groundnut is principal oilseed crop and contributes about 35% of area and 40% of oilseed crops grown in the country. Among the major groundnut growing states Gujarat rank first in production (1.8 million tonnes) and second in area (1.85 million hectares) in the country. Plant breeders are always interested in assessing genetic divergence among the varieties or advance breeding line available with them so as to utilize in directed breeding programme because genetically diverse parents are likely to produce high heterotic effects and the distinctly related parents within species, when utilized in cross breeding programme, are likely to produce a wide spectrum of variability (Arunachalam, 1981). D²-statistic developed by Mahalanobis (1936) was used to measure genetic divergence and to classify the genetic stock into distinct groups.

The experimental material comprised of 80 bunch groundnut genotypes. The experimental was laid out in Randomized Block Design with three replications at

instructional Farm of College of Agriculture Junagadh Agricultural University, Junagadh under irrigated condition during summer 2006. Each entry was accommodated in a single row of 3.0 m length with a spacing of 30 x 10 cm. The observations were recorded on five randomly selected plants in each entry and replication and their mean values were used for the analysis of genetic divergence using Mahalanobis's D²-statistics.

The analysis of variance revealed significant differences among the genotypes for all the characters under study thereby indicating the presence of ample variability among the genotypes. On the basis of magnitude of D² values all the 80 genotypes of groundnut for 14 characters showed that the generalized distance (D²) between two populations varied from 8.43 to 26.91, which was an indicator of considerable diversity available in the material evaluated. On the basis of D²-values, 9 clusters were formed from 80 genotypes (Table 1). The cluster I was the largest, accommodating as many as 19 genotypes followed by cluster II with 17 genotypes from different origins. Cluster III possessed 15 genotypes, while clusters IV and V contained 6 and 8 genotypes, respectively and clusters VI, VII and VIII possessed 4 genotypes each. Cluster IX having 3 genotypes. The clustering pattern of genotypes showed that the genotypes of different origins were clubbed into one cluster whereas, the genotypes belonging to same state or origin were grouped into different clusters indicating that the geographic distribution was not the sole criterion of genetic diversity. Murthy and Arunachalam (1966) also stated that genetic drift and selection in different environments could cause greater diversity than geographic distance. Further, the free exchange of seed materials among the different regions consequently causes characters constellations because of the human interference and material may lose its individuality. The results obtained in the present study are in accordance with the findings of Yadava *et al.* (1991), Golakia and Makne (1992), Katule *et al.* (1992), Reddy and Reddy (1993) and Venkataravana *et al.* (2000) who also reported no parallelism between geographic distribution and genetic diversity. Intra-cluster distance values were lower than the inter-cluster distances. Thus, the genotypes included within a cluster tended to diverse less from each other. Intra-cluster distance (D) ranged from 8.43 to 12.97 for

cluster VI and VIII, respectively (Table 3). The maximum inter-cluster distance (D) was observed between cluster I and VIII (D=26.91) followed by that between cluster IV and VIII (D=25.90), cluster III and VIII (D=25.62) and cluster II and VIII (D=25.25), while the closest proximity was noticed between the cluster II and III (D=11.22). Inter-cluster distance is the main criterion for selection of genotype. In this context, the genotypes from cluster I, II, III and IV could be selected as parents for hybridization with those of cluster VIII. A wide to moderate range of variation for several characters among clusters was noticed (Table 3). However, the differences were more pronounced for number of secondary branches/plant and number of aerial pegs/plant. The present findings are in conformity with those reported earlier in groundnut by Golakia and Makne (1992), Katule *et al.* (1992) and Reddy and Reddy (1993). The clustering pattern could be utilized in selecting the parents and deciding the cross combinations which may generate the highest possible variability for various traits. The genotypes with high values of divergent clusters can be used in hybridization programme for further selection and improvement. In the present study, the

cluster VIII differed from other clusters in respect of pod yield/plant, number of secondary branches/plant, number of aerial pegs/plant, number of kernels/pods, 100-kernel weight and harvest index. The cluster I had desirable value for days to 50% flowering and days to maturity. The cluster IV and III had desirable values for shelling percentage and oil content, respectively. The analysis for percent contribution of various characters toward expression of total genetic divergence indicated that number of secondary branches/plant (37.09%) followed by shelling per cent (22.53%), number of aerial pegs (14.34%), oil content (11.71%) and number of under ground pegs (4.97%) contributed maximum towards divergence. These characters accounted for more than 90% of the total divergence in the material investigated. Therefore, in the present investigation based upon high yielding genotypes and large inter-cluster distances, it is advisable to attempt crossing of the genotypes from cluster I cluster IV cluster III and cluster II with genotypes of cluster VIII. This may lead to broad spectrum of favourable genetic variability for pod yield improvement in groundnut.

Table 1 Grouping of 80 genotypes of bunch groundnut in various clusters on the basis of D² statistics

Cluster	Number of Genotypes	Name of the genotypes	Origin
I	19	URR 304 ICAS 109 JB FDR 29, JB(E) 336, JE 6, JB 459, JB HOC 6, JB 904, GG 2, GG 4, GG 5, GG 6, GG 7, JB 823, JB 605, J 28, Jawan BDH 510 NCAC 995	Maharashtra Andhra Pradesh Gujarat Orissa Karnataka U.S.A.
II	17	JB 436, JB 60, JB FDR 59, Ginar 1, JBD 36, FeEsg 10, FeEsg 8 AK 7171, TG 22, TG 3, AK 408 DH 20 NIAC 1727 DSA 201 EC 21139, EC 21051 Kisan	Gujarat Maharashtra Karnataka U.S.A. Argentina Cuba Orissa
III	15	ICGV 87935, ICGV 92127, ICGV 89359, ICGV 86590, ICGS 10, ICG FDRS 10 EC 100827 J 54, JB HOC 4, JB 452 TG 40, TG 50, TAG 24 ALR 1 R 9251	Andhra Pradesh Paraguay Gujarat Maharashtra Tamil Nadu Karnataka
IV	6	GC 400, JB 779, JB 959, JB FDR 55 VRI 4 BH 532	Gujarat Tamil Nadu Bihar
V	8	JB 579, JB 696 AH 7228, AK 159 SG 84 ICGS 76, ICGV 86934 EC 24405	Gujarat Maharashtra Punjab Andhra Pradesh
VI	4	AH 8253, AH 6902 Gemuma CSMa 841	South Africa Maharashtra Sudan
VII	4	IJS 71 NCAC 519 HNG 37 VG 8934	Uttar Pradesh Andhra Pradesh U.S.A. Rajasthan
VIII	4	ICHMG 88409, ICHMG 88398 JVB 148, JVB 185	Tamil Nadu Andhra Pradesh Gujarat
IX	3	M 522, M 335 JVB 151	Punjab Gujarat

Table 2 Average inter- and intra-cluster distance ($D=D^2$) values in bunch groundnut

	I	II	III	IV	V	VI	VII	VIII	IX
I	8.84	11.23	11.81	14.26	14.09	13.60	19.04	26.91	21.70
II		8.82	11.22	15.95	12.54	12.38	18.60	25.25	18.21
III			9.83	17.26	12.80	14.17	18.56	25.62	18.70
IV				10.91	14.81	17.36	21.12	25.90	22.94
V					10.12	11.56	14.54	19.23	13.86
VI						8.43	11.72	18.82	14.25
VII							9.55	15.29	13.48
VIII								12.97	15.18
IX									9.93

Table 3 Cluster means for 14 different characters in 80 genotypes of bunch groundnut

Clusters	Pod yield/plant (g)	Days to 50% flowering	Days to maturity	No. of primary bran./plant	No. of secondary bran./plant	Plant height (cm)	No. of aerial pegs/plant	No. of underground pegs/plant	Biological yield/plant (g)	No. of kernels/pod	100 kernel weight (g)	Harvest index (%)	Shelling per cent	Oil content (%)
I	13.1	47.8	115.3	4.7	1.2	22.0	4.4	28.2	33.8	2.0	37.3	38.6	71.6	48.4
II	11.6	48.4	116.2	4.7	1.3	17.8	5.5	27.7	28.3	1.9	34.8	40.6	66.0	48.5
III	12.8	48.9	116.5	5.0	1.6	20.7	4.4	30.7	31.9	1.9	40.3	40.3	67.3	51.2
IV	11.1	48.9	116.6	5.0	1.4	21.1	13.6	31.3	32.0	2.0	35.2	35.5	73.6	48.5
V	12.2	49.2	116.5	5.5	3.5	19.5	8.7	33.8	31.9	2.0	37.8	38.4	67.4	49.7
VI	11.6	50.3	117.7	4.6	4.3	22.1	4.3	28.5	31.2	1.8	38.9	38.0	68.2	47.5
VII	11.0	52.5	120.0	5.4	6.7	24.4	4.0	25.3	34.2	1.9	35.3	32.9	70.0	49.4
VIII	14.1	49.4	118.1	5.2	9.2	21.8	10.5	30.1	33.4	2.1	43.1	41.8	66.9	49.8
IX	10.3	50.1	117.6	5.3	6.2	15.1	6.9	34.3	32.4	2.0	35.5	33.6	62.6	50.4
Mean	12.2	48.9	116.5	4.9	2.6	20.4	6.1	29.6	31.8	2.0	37.4	38.7	68.5	49.2
SEm ±	1.1	1.1	1.2	0.4	0.5	2.4	1.1	3.3	2.2	0.1	3.1	2.4	1.5	0.6
C.V.%	15.8	4.0	1.7	13.8	31.3	20.4	31.1	19.3	11.8	8.8	14.2	10.8	3.7	2.1
Percentage contribution of characters towards total divergence														
No. of times appearing first	22	0	14	23	1172	109	453	157	22	74	32	0	712	370
Per cent contribution	0.7	0.0	0.4	0.7	37.1	3.5	14.3	5.0	0.7	2.3	1.0	0.0	22.5	11.7

References

- Arunachalam, V. 1981. Genetic distance in plant breeding. *Indian Journal of Genetics*, **41**: 226-236.
- Golakia, P.R. and Makne, V.G. 1992. D^2 -analysis in Virginia runner groundnut genotypes. *Indian Journal of Genetics*, **52**: 252-256.
- Katule, B.K., Thombre, M.V., Dumbre, A.D. and Pawar, B.B. 1992. Genetic diversity in bunch groundnut. *Journal of Maharashtra Agricultural Universities*, **17**: 302-303.
- Mahalanobis, P.C. 1936. On the generalized distance in statistics. *Proceeding of Indian National Institute of Science, (India)*, **2**: 49-55.
- Murthy, B.R. and Arunachalam, V. 1966. The nature of divergence in relation to breeding systems in crop plants. *Indian Journal of Genetics*, **25**: 188-198.
- Reddy, K.H.P. and Reddy, K.R. 1993. Genetic divergence in groundnut (*Arachis hypogaea* L.). *Annals of Agriculture Research*, **14**: 9-14.
- Venkataramana, P., Sheriff, R.A., Kulkarni, R.S., Shankaranarayana, V. and Fathima, P.S. 2000. Genetic divergence in groundnut (*Arachis hypogaea* L.). *Mysore Journal of Agricultural Sciences*, **34**: 325-329.
- Yadava, J.S., Yadav, I.S. and Chaudhry, M.S. 1991. Genetic diversity in bunch groundnut (*Arachis hypogaea* L.). *Haryana Agricultural University Journal of Research*, **21**: 133-136.

Short communication

Phenotypic evaluation of RILs for rust and late leaf spot and their association on productivity traits in groundnut, *Arachis hypogaea* L.

Sarvamangala Cholin and M.V.C. Gowda

Department of Genetics and Plant Breeding, University of Agricultural Sciences, Dharwad, Karnataka

(Received: September, 2009; Revised: December, 2009; Accepted: January, 2010)

Abstract

The mapping population used in the present study was showing better genetic variability and consistent positive association between the stages and seasons for both the diseases i.e., rust and leaf spot indicating the usefulness of population for mapping and tagging for these two diseases using appropriate molecular markers. The association pattern for productivity with rust indicates the importance of resistance for improved productivity in groundnut. However, the association was not clearly observed with late leaf spot revealing the need to screen in multiple seasons.

Keywords: Groundnut, rust, late leaf spot

Groundnut (*Arachis hypogaea* L.) is damaged by late leaf spot and rust diseases (Kokalis-Burette *et al.*, 1997). Moderate level of resistance is available in the cultivated groundnut gene pool for late leaf spot and several wild *Arachis* species also possess very high levels of rust resistance.

There has been limited success in transferring resistance from wild *Arachis* to cultivated groundnut, mainly because of inter-specific compatibility barriers, resistance being linked with many undesirable pod/seed characteristics and longer periods required for developing stable tetraploid interspecific derivatives. However, an inter-specific derivative GPBD 4 released at U.A.S. Dharwad combined early maturity, high yield potential and high shelling outturn with minimum yield reduction due to high level of resistance to rust and late leaf spot with higher pod growth rate, partitioning coefficient and harvest index (Gowda *et al.*, 2001). The present study has aimed at evaluating the mapping population for the genetic variability components including heritability and correlation among these two diseases and the correlation of them with important productivity traits.

A mapping population consisting of 146 recombinant inbred lines obtained ($F_9:F_{10}$) by crossing TG 26 x GPBD 4 was used for the study. TG 26 is an early maturing, semi dwarf, erect variety with high pod growth rate, high harvest index, greater partitioning efficiency, tolerance to bud

necrosis and rust with high linoleic acid content but susceptible to LLS (Kale *et al.*, 1997; Badigannavar *et al.*, 2002). GPBD 4 developed at University of Agricultural Sciences, Dharwad is a second cycle product of interspecific hybridization with desirable combination of early maturity, high yield, high pod growth rate, desirable pod and kernel features, high oil and protein content, better oleic/linoleic (O/L) ratio, resistant to late leaf spot and rust (Gowda *et al.*, 2002). Phenotyping for both the diseases was carried out according to modified 9 point scale Disease scoring on rust and LLS was recorded in Rainy 2005 and Rainy 2007 at 70 days (stage I) and 90 days (stage II). The productivity traits were evaluated during rainy 2007 for each RIL in two replications.

Phenotypic data obtained for both the diseases were subjected for pooled analysis of variance (ANOVA), mean and range and genetic variability components were estimated for individual season as well as pooled across the seasons. Correlation between the seasons, within the season between the stages and between both the diseases was done and the mean replicated data obtained for six productivity traits during rainy 2007 was used for correlation analysis with both rust and LLS during the same seasons. All the statistical parameters were estimated using SPAR software.

Mapping population exhibited significant variation for rust and LLS. Significant seasonal and genotype x season interaction indicated the need for screening in multiple environments (Table 1). Khedikar (2008) also found significant G x E interaction for these two diseases in a mapping population of 268 RILs obtained from the cross TAG 24 x GPBD 4.

The components of variation viz., PCV and GCV revealed substantial variation for both the diseases (Table 2). Further, moderate to high heritability and GAM indicated highly heritable nature of the variation. The estimates of components of variation were very low in one of the seasons (Rainy, 2007) for LLS which could be due to predominance of rust in that season. Usually both LLS and rust occur together but the incidence and severity vary between localities and seasons (Subramanyam *et al.*, 1984) and relative occurrence of these two diseases can

Phenotypic evaluation of RILs for rust and late leaf spot and their association on productivity traits in groundnut

influence precision and assessment of diseases in the genotypes. When compared across stages, the components were low at later stage especially for LLS indicating suitability of first stage for better discrimination. The reduction in different components of variability under pooled analysis was more for LLS as compared to rust indicating preponderance of G x E interaction.

The disease scores between the stages in a season and between seasons at a particular stage were highly correlated revealing the consistency of disease reaction in the individual genotypes for both the diseases in spite of significant G x E interaction. In contrast, correlation between rust and LLS was negative indicating differential

prevalence of resistance in the RILs for the two diseases (Table 3).

Most of the productivity traits exhibited negative correlations with rust disease indicating favorable influence of rust resistance on productivity (Table 4). In contrast, there was a lack of association for many of the productivity traits with late leaf spot. LLS at stage II had a negative correlation with 100-seed weight indicating its favorable association with resistance. At Stage I, positive correlation was evident with number of pods/plant and pod yield/plant revealing the association of higher productivity with susceptibility.

Table 1 Pooled ANOVA for rust and late leaf spot traits in TG 26 x GPBD 4 mapping population

Traits/Source of variation	Mean sum of squares							
	Season	Replication	S x R	Genotypes	S X G	Error	CV	Sed
Diseases								
Df	1	1	1	145	145	290		
Rust								
Stage I	51.48**	0.43	1.157	1.42**	0.58**	0.32	17.24	0.39
Stage II	33.08**	2.09	0.29	3.43**	1.60**	0.47	15.98	0.48
Late leaf spot								
Stage I	263.12**	30.75	75.51	4.44**	2.42**	1.11	21.33	0.74
Stage II	234.40**	15.78	0.36	2.11**	1.27**	0.73	12.58	0.60

Table 2 Mean, range and genetic variability components for rust and late leaf spot in TG 26 x GPBD 4 mapping population

Traits	Parents		RILs					
	TG 26	GPBD 4	Mean	Range	PCV	GCV	h ²	GAM
Rainy 2005								
Rust (Stage I)	3.0	2.0	2.9	2.0-5.0	31.4	23.6	56.6	36.6
Rust (Stage II)	5.0	3.0	4.0	2.0-8.0	37.5	31.7	71.7	55.2
LLS (Stage I)	7.0	2.0	5.6	2.5-7.0	26.4	23.2	77.2	42.2
LLS (Stage II)	8.0	3.0	6.1	3.0-8.0	19.4	14.2	52.9	21.2
Rainy 2007								
Rust (Stage I)	5.0	2.0	4.8	2.0-8.0	41.8	38.2	83.7	7.1
Rust (Stage II)	7.5	3.0	6.4	3.0-9.0	33.2	28.9	75.7	51.9
LLS (Stage I)	4.0	2.5	4.2	2.0-9.0	35.7	18.3	26.2	19.1
LLS (Stage II)	7.5	4.0	7.4	5.0-9.0	13.2	5.8	19.5	5.3
Plant height (cm)	15.9	29.4	23.0	8.0-33.3	21.9	16.0	53.3	1.0
Number of branches	4.7	4.5	6.0	3.0-13.5	22.5	0.5	0.1	0.0
Number of pods/plant	13.0	16.6	16.7	5.5-45.5	34.8	12.8	13.6	0.6
Pod yield/plant (g)	7.0	10.2	12.9	2.7-37.2	37.5	17.8	22.6	1.3
Shelling %	68.7	75.9	76.6	61.7-87.5	5.2	1.9	13.9	0.0
100-seed weight (g)	32.6	50.7	34.2	17.3-58.7	21.6	14.7	46.4	20.6
Across seasons								
Rust (Stage I)	4.0	2.1	3.7	2.1-5.1	28.7	22.7	62.5	37.0
Rust (Stage II)	5.0	2.7	4.9	2.7-6.6	27.9	22.1	62.7	36.1
Rust (Stage III)	6.4	3.0	5.6	3.0-7.5	26.4	18.7	50.1	27.2
LLS (Stage I)	5.5	2.2	4.9	2.7-7.0	25.7	14.3	31.2	16.4
LLS (Stage II)	7.7	3.5	6.7	5.5-8.0	14.2	6.6	22.0	6.4

Table 3 Correlation between rust and late leaf spot in various seasons

Traits	Rust Stage I	Rust Stage II	Late Leaf spot Stage I	Late Leaf Spot Stage II
Rust Stage I				
Rainy 2005	1.000	0.905**	-0.505**	-0.430**
Rainy 2007	1.000	0.943**	-0.273**	0.105*
Rust Stage II				
Rainy 2005		1.000	-0.591**	-0.526**
Rainy 2007		1.000	-0.301**	0.034
Late Leaf spot Stage I				
Rainy 2005			1.000	0.858**
Rainy 2007			1.000	0.441**
Late Leaf spot Stage II				
Rainy 2005				1.000
Rainy 2007				1.000

Table 4 Correlation for diseases (rust and LLS) with productivity traits in TG 26 x GPBD 4 mapping population

Traits	LLS Stage I	LLS Stage II	Rust Stage I	Rust Stage II	Rust Stage III	Rust (I) Stage I	Rust (I) Stage II
Plant height	0.208*	-0.161	0.124	0.042	-0.034	0.004	-0.028
No. of Branches	-0.006	-0.007	-0.001	0.019	0.006	0.021	-0.002
Number of pods/plant	0.211*	0.011	-0.238**	-0.264**	-0.298**	-0.297**	-0.322**
Pod yield/plant	0.242**	0.012	-0.314**	-0.318**	-0.344**	-0.386**	-0.402**
shelling %	-0.020	-0.064	-0.314**	-0.236*	-0.265**	-0.250**	-0.179*
100-seed weight	0.145*	-0.152*	-0.288**	-0.301**	-0.296**	-0.310**	-0.303**

References

- Badigannavar A.M., Kale, D.M. and Murty, G.S.S. 2002. Assessment of yielding ability of Trombay groundnut varieties through growth analysis. *Journal of oilseeds Research*, 19: 38-43.
- Gowda, M.V.C., Motagi, B.N., Chandran, K.P. and Ashalatha, K.V. 2001. Modelling yield loss due to rust and late leaf spot in Groundnut-Implications to crop improvement. *Proceedings of National Seminar on "Role of Plant Physiology for Sustaining Quality and Quantity of Food Production in Relation to Environment"* 5-7 December, pp 8-11.
- Gowda, M.V.C., Motagi, B.N., Naidu, G.K., Diddimani, S.B. and Sheshagiri, R. 2002. GPBD 4 : A Spanish bunch groundnut genotype resistant to rust and late leaf spot. *International Arachis Newsletter*, 22 : 29-32.
- Kale, D.M., Mouli C., Murthy, G.S.S. and Rao M.V.P. 1997. Development of a new groundnut variety, TG 26 by using induced mutations in cross breeding. *Mutation Breeding Newsletter*, 43: 25-27.
- Khedikar Y.P. 2008. Molecular tagging and mapping of resistance to late leaf spot and rust in Groundnut (*Arachis hypogaea* L.). Ph.D. Thesis, University of Agricultural Sciences, Dharwad (India).
- Kokalis-Burette, N., Porter, D.M., Rodriguez-Kábana, R., Smith, D.H. and Subrahmanyam, P. 1997. *Compendium of Peanut Diseases*. American Phytopathology Society, St. Paul, MN, USA, p. 94.
- Subrahmanyam, P., Williams, J.H., McDonald, D. and Gibbons, R.W. 1984. The influence of foliar diseases and their control by selective fungicides on a range of groundnut (*Arachis hypogaea* L.) genotypes. *Annals of Applied Biology*, 104: 467-476.

Short communication

Heterosis and genetic architecture for quality traits in sesame, *Sesamum indicum* L.

N.N. Prajapati, C.G. Patel, R.N. Patel, A.M. Patel and K.M. Patel

Department of Seed Technology, Sardarkrushinagar Dantiwada Agril. University, Sardarkrushinagar-385 506, Gujarat

(Received: July, 2009; Revised: September, 2009; Accepted: October, 2009)

Abstract

The studies on heterosis and genetic architecture for quality traits in sesame, *Sesamum indicum* L. indicated that non-additive gene action was predominantly responsible for inheritance of quality traits, heterosis breeding could be the best breeding method for improvement.

Keywords: *Sesamum indicum*, genetic architecture

Sesame (*Sesamum indicum* L.), is a very ancient oilseed crop, generally known as til, tal and gingelly. India is the largest producer and exporter of sesame in the world. The genetic make up of genotypes for inherited traits can be well understood by the study of genetic parameters. The combining ability analysis has been utilized to know the nature and extent of gene action controlling the inheritance of yield and quality traits for getting better recombinants. The present investigation was carried out to evaluate genotypes of sesame for their heterosis and combining ability.

A half diallel set was made using 10 sesame genotypes during *kharif* 2004. The 10 parents and their 45 F₁s were evaluated in a Randomized Block Design with three replications under four environments created by four different dates of sowing during *kharif* 2005. The field experiment was conducted at the main castor and mustard research station, S.D. Agricultural University, Sardarkrushinagar with 45 x 15 cm spacing. The observations were recorded on five randomly selected plants of each genotype in each replication and each environment. The data were analyzed for heterosis according to Fonseca and Petterson (1968) and combining ability for single environment using Griffing (1956), Model 1 and Method II.

The analysis of variance for combining ability over environments was carried out and the mean squares are presented in Table 1. The results revealed that mean squares due to GCA and SCA were highly significant for all the traits. The ratio of $\delta^2_{gca}/\delta^2_{sca}$ was less than unity suggesting the major role of non-additive gene action for the inheritance of all the characters except for protein content. For protein content variance ratio of $\delta^2_{gca}/\delta^2_{sca}$

was greater than unity indicating the importance of additive type of gene action.

The parents, C 1013, AT 123 and GT 2 were found to be good general combiners for seed yield (Table 2). Among these, C 1013 was also found to be best combiner for oil content and protein content. It was further noted that involvement of these parents in hybridization has resulted into hybrids expressing useful heterosis for various traits. The parent C 1013, Mrug 1 and Tapi were good combiners for oil content, whereas, parent Tapi, C 1013 and PT 64 were good combiners for protein content. Parent C 1013 should be utilized in breeding programme for improvement of seed yield and quality traits. Similar finding was also noted by Karuppaiyan and Sundaresan (2002), Krishnaiah *et al.* (2003) and Prajapati *et al.* (2006).

The estimates of *sca* effects pooled over environments revealed that GT 2 x GT 10, Mrug 1 x PT 64 and C 1013 x ABT 23 were found to be best specific combiners for seed yield, which also showed significant desirable *sca* effects for quality characters. The crosses, Mrug 1 x PT 64, Mrug 1 x ABT 26 and GT 2 x ABT 23 for oil content and C 1013 x GT 10, GT 10 x Tapi and Tapi x AT 123 for protein content were found to be best specific combiners.

A comparative study of hybrids on the basis of per se performance and *sca* effects revealed that the most of the crosses, which produced maximum yield, had at least one parent as good combiner for seed yield. This suggested that the parent with higher *gca* effects are desirable for a hybrid having *sca* effects for realization of the maximum heterosis. A perusal of the data given in Table 2 also revealed that, the cross having higher desirable estimates of *sca* which resulted from good x good, good x average and also good x poor general combiners. This supports the importance of both additive and non-additive genetic variance in quality traits.

The hybrid, ABT 23 x ABT 26 expressing the highest heterobeltiosis for seed yield/plant followed by Mrug 1 x PT 64 also manifested high heterobeltiosis for oil content on pooled basis (Table 2).

An over view of the results suggested that since non-additive gene action was predominantly responsible

for the inheritance of all the traits, heterosis breeding could be the best breeding method for improvement. Since both additive and non-additive genetic variances are important, to take advantage of both types of gene action, reciprocal recurrent selection (Dixit, 1976) or biparental mating

(Krishnadoss *et al.* 1987) followed by modified recurrent selection may be resorted for the improvement of yield and yield related traits for the development of high yielding stable genotypes of sesame.

Table 1 Pooled analysis of variance of combining ability for different quality characters in sesame

Source of variation	d. f.	Seed yield/plant	1000–seed weight	Oil content	Protein content
<i>gca</i>	9	10.31**	0.74**	59.09**	72.31**
<i>sca</i>	45	3.39**	0.12**	33.06**	0.86**
Error	432	0.49	0.01	0.37	0.06
$\delta^2 gca$	-	0.21	0.02	1.22	1.51
$\delta^2 sca$	-	0.73	0.03	8.17	0.20
$\delta^2 gca/\delta^2 sca$	-	0.28	0.58	0.15	7.49

*,** indicate significance at P=0.05 and P=0.01 levels, respectively.

Table 2 Three best performing parents, best general combiners and best performing hybrids along with their *sca* effects and best specific cross combinations and best heterotic combinations in desired direction for various characters in sesame

Character	Best performing parents	Best general combiners	Best performing hybrids	Status of parents	<i>sca</i> effects	Best cross combination	Best heterotic combination over	
							MP	BP
Seed yield/plant	GT 2	C 1013	GT 2 x GT 10	G x A	2.71**	GT 2 x GT 10	Mrug 1 x PT 64	ABT 23 x ABT 26
	AT 123	AT 123	C 1013 x ABT 23	G x P	1.49**	Mrug 1 x PT 64	C 1013 x ABT 23	Mrug 1 x PT 64
	C 1013	GT 2	ABT 26 x AT 123	A x G	1.40**	C 1013 x ABT-23	GT 2 x GT 10	C 1013 x ABT 23
1000–seed weight	Mrug 1	Mrug 1	GT 1 x Tapi	G x G	0.30**	Mrug 1 x ABT 23	GT 1 x ABT 23	GT 1 x Tapi
	PT 64	Tapi	Mrug 1 x ABT 23	G x A	0.32**	GT 1 x Tapi	Mrug 1 x ABT 23	GT 10 x ABT 23
	Tapi	GT 1	Mrug 1 x Tapi	G x G	0.04	GT 1 x ABT 123	GT 1 x Tapi	GT 1 x ABT 23
Oil content	GT 10	C 1013	Mrug 1 x PT 64	G x P	5.98**	Mrug 1 x PT 64	Mrug 1 x ABT 26	Mrug 1 x PT 64
	C 1013	Mrug 1	Mrug 1 x AB 26	G x P	5.37**	Mrug 1 x AB 26	Mrug 1 x PT 64	Mrug 1 x ABT 26
	GT 1	Tapi	C 1013 x Mrug 1	G x G	1.82**	GT 2 x ABT 23	C 1013 x ABT 26	GT 2 x AT 123
Protein content	Tapi	Tapi	C 1013 x Tapi	G x G	0.03	C 1013 x GT 10	Mrug 1 x PT 64	Mrug 1 x GT 10
	C 1013	C 1013	PT 64 x Tapi	G x G	0.20	GT 10 x Tapi	Mrug 1 x Tapi	C 1013 x PT 64
	PT 64	PT 64	C 1013 x PT 64	G x G	0.29**	Tapi x AT 123	C 1013 x GT 10	C 1013 x Tapi

*,** indicate significance at P=0.05 and P=0.01 levels, respectively.

References

Dixit, R.K. 1976. Inheritance of yield and its components in sesame. *Indian Journal of Agricultural Sciences*, **46**: 187-191.

Fonseca, S. and Patterson, F. 1968. Hybrid vigour in a seven parent diallel cross in common winter wheat (*Triticum aestivum* L.). *Crop Science*, **8**: 85-86.

Griffing, B. 1956. Concept of general and specific combining ability in relation to diallel crossing system. *Australian Journal of Biological Sciences*, **9**: 463-493.

Karupaiyan, R. and Sundaresan, N. 2002. Combining ability and heterosis for seed yield in sesame (*Sesamum indicum* L.). *Madras Agricultural Journal*, **89** (4-6): 359-361.

Krishnadoss, D., Kadabavana Sundaram, M., Ramalingam, R.S. and Rajasekaran, S. 1987. Combining ability in sesame. *Indian Journal of Agricultural Sciences*, **5**: 85-88.

Krishnaiah, G., Reddy, K.R. and Sekhar, M.R. 2003. Heterosis and combining ability in sesame (*Sesamum indicum* L.). *Journal of Oilseeds Research*, **20** (2): 229-233.

Prajapati K.P., Patel, K.M., Prajapati, B.H. and Patel, C.J. 2006. Genetic analysis of quantitative traits in sesame (*Sesamum indicum* L.). *Journal of Oilseeds Research*, **23** (2): 171-173.

Short communication

Evaluation of progenies selected from random mated population of safflower, *Carthamus tinctorius* L. using GMS lines

P.V. Mahajan, S.N. Deshmukh and R.D. Ratnaparkhi

Oilseeds Research Unit, Dr. Panjabrao Deshmukh Krishi Vidyapeeth, Akola-444 104, MS

(Received: August, 2007; Revised: November, 2008; Accepted: July, 2009)

Abstract

Safflower (*Carthamus tinctorius* L.) is one of the most important *rabi* oilseed crop belonging to family Asteraceae. Low seed yield and low market prices, due to low oil content coupled with long duration and susceptibility to biotic and abiotic stresses, restricted the spread of safflower crop beyond traditional safflower growing area of the region. So, this crop needs further improvement genetically with agronomic intervention. Recurrent selection is efficient method of population improvement which increases the frequency of desirable genes, maintain high genetic variability and breaks undesirable linkages. Genetic male sterility system available in safflower may help in this direction.

Keywords: Safflower, genetic male sterile

The material of the present study derived from the GMS based hybrids made earlier. During *rabi* 2002-03, 72 hybrids were made using nine GMS lines and eight pollen parents. These hybrids were evaluated during 2003-04 and superior 54 hybrids were selected to constitute base population-I. Equal amount of seeds of 54 hybrids (F_1 's seeds) were composited and sown during *rabi* 2004-05 for recombination in open pollinated condition in isolation and in this F_2 bulk 256 male sterile plants were tagged at the time of flowering and harvested individually. Out of these 196 plants having sufficient seeds were selected for evaluation and sown during *rabi* 2005-06 along with two checks *viz.*, AKS 207 and Bhima keeping remnant seeds to constitute next recombination cycle. Material was sown in two replications in modified Randomized Block Design. Each replication consisted of ten blocks and each block consisted of 20 progenies along with two checks. The observations were recorded on ten characters (Table 2). The data were statistically analysed to estimate mean, genotypic and phenotypic variances, genotypic and phenotypic coefficient of variation (Burton, 1951), heritability (broad sense) as per Johnson *et al.* (1955) and genetic advance (Lush, 1949). As well as the simple genotypic, phenotypic correlation coefficient were worked out from the respective variances and covariances as per

Hayes *et al.* (1955). Mean performance (Table 1) indicated that maximum range was existing in plant height followed by number seeds per capitulum, seed yield/plant and number of capitula/plant, primary branches/plant, oil content and 100 seed weight. Similar trend was reported by Eckebil *et al.* (1977) and Flores *et al.* (1986).

PCV was greater than GCV for all the characters indicating apparent variation was due to both environment and genotype (Table 2). Estimates of GCV and PCV were low for days to maturity and 30.33 and seed yield/plant. Number of branches, number of seeds/capitulum and number capitula/plant exhibited high GCV and PCV indicate large amount of variation and less amenability to environmental fluctuations. Similar results were recorded by Senapati *et al.* (1999) and Sarang *et al.* (2004). Low GCV and PCV for days for maturity and 50% flowering are in conformity with the finding of Nair *et al.* (2006). Heritability (broad sense) was low 64.31% for oil content and it was high 84.97% for seed yield/plant. Characters exhibited high heritability indicated greater scope for selection for improvement of yield and field component. High heritability are in conformity with the findings of Senapati *et al.* (1999) and Goyal (2006).

Expected genetic advance as mean of population at 5, 10 and 20% selection intensity was high for seed yield/plant. EGA over check varieties, i.e., AKS 207 and Bhima at 5, 10 and 20% selection intensity was high for seed yield/plant over AKS 207 and Bhima, respectively. High heritability along with high expected genetic advance indicated the presence of additive gene action.

Higher genotypic correlations than the phenotypic one indicated inherent relationship between the characters. Table 4 revealed that seed yield/plant had positive and significant association with number of primary branches/plant and number of capitula/plant at both genotypic and phenotypic levels. Similar findings was also recorded by Jwanjal *et al.* (2006). Seed yield/plant had significant positive correlation with number of seeds/capitulum at genotypic level. Diwakar *et al.* (2006) observed same results. Seed yield/plant had positive significant association at genotypic and phenotypic level with oil content which is in conformity with the findings of

Esendal and Tosun (1972) and Tunçtürk and Ciftci (2004). In general seed yield in negatively correlated with oil content as per Ghongade *et al.* (1993). He also suggested that it is possible to break such undesirable relations by random mating among segregants followed by selection. Seed yield/plant had significant and negative correlation with days to 50% flowering at genotypic and phenotypic

levels. These findings are also consonance with the findings of Diwakar *et al.* (2006). The present investigation clearly showed that the traits, viz., number of primary branches/plant, number of capitula/plant and number of seed/capitulum had strong association with seed yield/plant. This indicated that direct selection for these traits will definitely enhance the seed yield in safflower.

Table 1 Mean performance progenies

Statistic	Days to first flowering	Days to 50% flowering	Days to maturity	Plant height (cm)	No. of primary branches/plant	No. of capitula/plant	No. of seeds/capitulum	100 seed wt. (g)	Oil content (%)	Seed yield/plant (g)
Minimum (Progeny No.)	74 (PI-140)	82 (PI-82)	139 (PI-73)	75.2 (PI-18)	3.8 (PI-188)	6.4 (PI-188)	10.4 (PI-83)	4.66 (PI-95)	27.02 (PI-63)	8.73 (PI-83)
Maximum (Progeny No.)	103 (PI-194)	110 (PI-196)	160 (PI-196)	125.2 (PI-195)	17.4 (PI-193)	42.4 (PI-46)	58.0 (PI-178)	7.27 (PI-187)	33.48 (PI-152)	55.53 (PI-167)
Range	29	28	21	50.0	13.6	36.0	47.6	2.61	6.46	46.80
Mean (progenies)*	85.82	91.10	145.80	95.87	9.89	25.08	27.50	5.91	29.37	29.15
Checks										
AKS-2072	79.5	85.75	142.15	91.09	10.12	26.76	27.3	5.68	30.39	36.76
Bhima**	86.35	92.05	145.65	93.37	10.13	26.34	27.4	5.62	31.26	37.48

* Mean of 196 progenies; ** Mean of check varieties

Table 2 Parameters of genetic variability for different characters

Character	Range	Mean	Genotypic variance	Phenotypic variance	GCV (%)	PCV (%)	Heritability % in (Broad sense)	GA (%)	EGA as (%) over mean of population (at 5%)
Days to first flowering	74-103	85.56	13.53	18.12	4.29	4.97	74.68	6.54	7.65
Days to 50% flowering	82-110	90.90	10.24	12.32	3.52	3.86	83.16	6.01	6.81
Days to maturity	139-160	145.62	8.09	10.10	1.95	2.18	80.07	5.24	3.61
Plant height (cm)	75.2-125.2	95.53	33.59	41.43	6.06	6.73	81.05	10.74	11.23
No. of branches/plant	3.8 - 17.4	9.91	5.50	7.24	23.66	27.15	75.89	4.20	42.46
No. of capitula/plant	6.4 - 42.4	25.21	29.81	40.38	21.65	25.19	73.82	9.66	38.32
No. of seeds/capitulum	10.4 - 58	27.51	39.92	52.80	22.96	26.40	75.60	11.31	41.10
100 seed weight (g)	4.66 - 7.27	5.89	0.223	0.285	8.01	9.06	78.23	0.86	14.61
Oil content (%)	27.02-33.48	29.51	1.26	1.96	3.81	4.75	64.31	1.85	6.30
Seed yield/plant (g)	8.73 - 55.53	29.89	82.27	96.82	30.33	32.91	84.97	17.22	57.61

GCV = Genotypic coefficient of variation; PCV = Phenotypic coefficient of variation

Table 3 Estimates of expected genetic advance over mean of population and over checks AKS-207 and Bhima

Unit of evaluation and selection	Cycle	Selection intensity#	Days to first flowering	Days to 50% flowering	Days to maturity	Plant height (cm)	No. of primary branches/plant	No. of capitula/plant	No. of seeds/capitulum	100 seed wt. (g)	Oil content	Seed yield/plant (g)
Expected genetic advance as per cent over mean of populationⁿ												
Progeny	1	5	7.65	6.61	3.61	11.23	42.46	38.32	41.1	14.61	6.30	57.61
		10	6.53	5.65	3.08	9.59	36.27	32.74	35.1	12.48	5.38	49.22
		20	5.20	4.49	2.45	7.63	28.85	26.04	28	9.93	4.28	39.15
Expected genetic advance as per cent over AKS-207												
Progeny	1	5	8.23	7.01	3.69	11.78	41.59	36.11	41.48	15.13	6.12	46.85
		10	7.03	5.99	3.16	10.06	35.53	30.85	35.44	12.93	5.23	40.02
		20	5.58	4.76	2.51	8.00	28.26	24.54	28.19	10.28	4.16	31.84
Expected genetic advance as per cent over Bhima												
Progeny	1	5	7.58	6.53	3.61	11.49	41.55	36.69	41.35	15.31	5.95	45.95
		10	6.47	5.58	3.08	9.82	35.50	31.35	35.32	13.08	5.08	39.26
		20	5.15	4.44	2.45	7.81	28.24	24.93	28.10	10.40	4.04	31.22

#=Response to selection of top 5% (K=2.06), 10% (K=1.76) and 20% (K=1.40) of large number of progenies, where 'K' is the standardized selection differential

Table 4 Genotypic and phenotypic correlation coefficient among different characters in safflower

Character		Days to first flowering	Days to 50% flowering	Days to maturity	Plant height (cm)	No. of primary branches /plant	No. of capitula /plant	No. of seeds/ capitulum	100 seed wt (g)	Oil content	Seed yield/ plant (g)
Days to first flowering	G	1.00	0.918**	0.655**	0.636**	-0.342**	-0.292**	0.190**	0.004	-0.044	-0.247**
	P	1.00	0.797**	0.514**	0.480**	-0.230**	-0.175*	0.136	-0.013	-0.030	-0.190**
Days to 50% flowering	G		1.00	0.762**	0.631**	-0.279**	-0.198**	0.136	-0.019	0.013	-0.198**
	P		1.00	0.650**	0.511**	-0.222**	-0.143*	0.107	-0.032	0.032	-0.166*
Days to maturity	G			1.00	0.451**	-0.159*	-0.107	0.150*	-0.074	0.092	-0.158*
	P			1.00	0.344**	-0.104	-0.091	0.109	-0.049	0.060	-0.132
Plant height (cm)	G				1.00	-0.244**	-0.140*	0.060	0.075	-0.131	-0.099
	P				1.00	-0.193**	-0.100	0.078	0.076	-0.066	-0.105
No. of primary branches/ plant	G					1.00	0.708**	-0.425**	-0.036	0.110	0.245**
	P					1.00	0.612**	-0.309**	-0.045	0.057	0.205**
No. of capitula/ plant	G						1.00	-0.431**	-0.043	0.105	0.503**
	P						1.00	-0.324**	-0.042	0.091	0.418**
No. of seeds/ capitulum	G							1.00	-0.102	0.143*	0.150*
	P							1.00	-0.063	0.134	0.126
100 seed wt. (g)	G								1.00	-0.325**	-0.186**
	P								1.00	-0.224**	-0.149*
Oil content (%)	G									1.00	0.516**
	P									1.00	0.373**

G = Genotypic correlation coefficient; P = Phenotypic correlation coefficient; * = Significant at 5%, ** = Significant at 1%

References

- Burton, G.W. 1951. Quantitative inheritance in pearl millet (*Pennisetum glaucum* S. and H.). *Agronomy Journal*, **43** : 404-417.
- Diwakar, R., Sreedhar, N. and Mukta, N. 2006. Studies on character association in safflower (*Carthamus tinctorius* L.). *International Journal of Agricultural Sciences*, **2** : 154-156.
- Eckebil, T.F., Ross, W.H., Gardner, C.O. and Marranville, J.W. 1977. Heritability estimates, genetic correlations and predicted gains from S1 progeny tests in three grain sorghum random mating populations. *Crop Science*, **17**: 374-377.
- Esendal, E. and Tosun, F. 1972. Research on the yield, phenological, morphological and seed characteristics of some native and foreign varieties of safflower (*Carthamus tinctorius* L.) under Erzurum ecological condition. *Zirat Dergis*, **3** : 93-115.
- Flores, C.I., Ross, W.M. and Maranville, J.M. 1986. Quantitative genetics of agronomic and nutritional traits in related grain sorghum random mating population as affected by selection. *Crop Science*, **26** : 9-12.
- Ghongade, R.A., Joshi, B.P. and Navale, P.A. 1993. Correlations and path analysis of yield components in safflower. *Journal of Maharashtra Agricultural Universities*, **18**(2) : 240-243.
- Goyal, V.S. 2006. Evaluation of third cycle of recurrent selection for seed yield and its components in safflower (*Carthamus tinctorius* L.). M.Sc. (Agril.) Thesis, Dr. PDKV, Akola.
- Hayes, F.E., Immer and Dc Smith. 1955. *Methods of plant breeding*. McGraw Hill Book Co. New York. pp 439-442. 535-536.
- Jawanjal, S.S., Choulwar, S.B. and Patil, S.R. 2006. Characters association and path analysis for yield in safflower. *Journal of Maharashtra Agricultural Universities*, **31** (1): 030-032.
- Johnson, H.W., Robinson H.F. and Comstock R.E. 1955. Estimates of genetic and environmental variability in soybean. *Agronomy Journal*, **47** : 314-318.
- Lush, J.L. 1949. Heritability of quantitative characters in farm animals. *Hereditas*, **35** : 356-375.
- Nair, B., Kalamkar, V., Barsod, S. and Lakshmi, M.K. 2006. Genetic association, path analysis and heritability studies in safflower. *Journal of Soils and Crops*, **16**(1) : 194-198.
- Sarang, D.H., Chavan, A.A., Chinchane, V.N. and Gore, B.M. 2004. Correlation and path analysis in safflower. *Journal of Maharashtra Agricultural Universities*, **29**(1) : 36-39.
- Senapati, N., Samal, K.M. and Dhal, A. 1999. Performance variability and character association in safflower (*Carthamus tinctorius*). *Indian Journal of Agricultural Research*, **33**(4) : 254-258
- Tunctoruk, M. and Ciftci, V. 2004. Relationship among traits using correlation and path coefficient analysis in safflower (*Carthamus tinctorius* L.) sown different fertilization level and row spacing. *Asian Journal of Plant Sciences*, **3**(6) : 683-686.

Short communication

Genetic divergence in linseed, *Linum usitatissimum* L. under salt stress condition

R.L. Srivastava, H.C. Singh, Karam Husain, Y.P. Malik and Om Prakash

Project Coordinating Unit (Linseed), C. S. Azad University of Agriculture & Technology, Kanpur-208 002, UP

(Received: February, 2009; Revised: June, 2009; Accepted: July, 2009)

Abstract

Genetic divergence under salt stress conditions for nine yield contributing characters were studied in linseed using 105 diverse genotypes including five checks. The analysis of variance revealed highly significant differences of means for all the characters. The 105 genotypes were grouped into twelve clusters where, cluster X was the largest containing 24 genotypes followed by cluster IV with 12 genotypes whereas, cluster I contained only 3 genotypes. The inter-cluster distance was the maximum between cluster I and VI based on inter-cluster distance and per se performance of genotypes, the entries, viz., A-18-1-2, A-6-1-5, 5/37-2/1-61/1, 5/47, 9x12, 477-3/2, A-62 and A-44 can be intercrossed to obtain high heterotic response and also to recover desirable transgressive segregants. Seed yield/plant contributed maximum divergence (80.7%) followed by number of capsules/plant (8.2%) and days to flowering (6.6%).

Keywords: Linseed, D² statistic, genetic divergence

Linseed (*Linum usitatissimum* L.) is one of the important oilseed crops of India. The crop is grown under diverse agro-production situations, across the climatic and geographic boundaries, which necessitated the development of more productive varieties of diverse origin. Crosses between divergent parents usually produce greater heterosis than those between closely related ones (Moll and Stuber, 1971). Genetic divergence studies are the vital tools for the evaluation of germplasm lines and selection of parents for the breeding programme. So, the present study was undertaken to measure genetic diversity among the germplasm lines.

The material for the present study consisted of 100 linseed germplasm accessions from GMU-01 to GMU-100 along with five checks i.e., Gaurav, Neelum, Kiran, T-397 and Padmini. The experiment was conducted at Crop Research Farm, Nawabganj of the university during *rabi* 2002 under salt sick soil (pH 8.9). Each accession was sown in single row of 3m length with a spacing of 40cm. between row and 4-5 cm between plants. The experiment was laid out in Randomized Complete Block Design in

three replications. In each accession, five plants were randomly selected for collection of data on yield and its component traits. The data was analysed as per Mahalanobis D² statistics (Mahalanobis, 1936) and Toucher's method as described by Rao (1952) for determining group. Average intra and inter cluster distances were estimated as per the procedure outlined by Singh and Choudhary (1977).

The analysis of variance revealed significant differences among the genotypes for all the traits under study except number of seeds per capsule. Based on D² statistics and Toucher's method 105 accessions (100 germplasm + 5 checks) were grouped into 12 clusters (Table 1). Cluster X had the maximum number with 24 genotypes followed by cluster IV with 12 genotypes, cluster III with 10 genotypes, cluster VII, VIII and XI with 9 genotypes, cluster VI and XII with 8 genotypes, clusters IX, II and I with 5, 4 and 3 genotypes, respectively. The genotypes from one source of origin clustered with the genotypes of other source of origin. This indicated that there was no parallelism between geographical distribution and genetic diversity. Three checks (Kiran, T-397 and Padmini) were included in cluster X and one each check i.e., Neelum and Gaurav were grouped into clusters XI and XII, respectively indicating their distinctness or similarity from the germplasm accessions with respect to traits considered. Similar findings were also reported by Chawla and Singh (1984) and Mahto *et al.* (1995) in linseed.

The clusters VI (A-18-1-2, A-6-1-5, 5/37-2/1-61/1, 5/47, 9x12, 477-3/2, A-62 and A-44) recorded the highest test weight (8.48 g) and technical plant height (35.9 cm). The cluster XII recorded the highest number of capsules/plant (150.4), plant height (75.1 cm) and days to maturity (144.4 days). Seed yield/plant contributes more than 80% followed by number of capsules/plant (8.24%) and days to flowering (6.6%) contributes maximum role for the divergence.

Average intra and inter cluster D² values among 105 genotypes (Table 2) revealed that cluster I, X, IV, XI and XII have the minimum intra cluster values viz., 1.32, 2.28, 2.31, 2.92 and 3.00, respectively indicating that genotypes within this cluster were similar. While, cluster V showed

the maximum intra cluster D^2 value (5.24) followed by cluster VI (4.93) and cluster III (4.41) revealing, thereby, the existence of diverse genotypes that fell in these clusters. The inter cluster D^2 values ranged from 3.17 to 43.56. Minimum inter cluster D^2 values were observed between cluster IV and X (3.17) indicating the close relationship among the genotypes included in these clusters. Maximum inter cluster value was observed between II and XII (43.56) followed by cluster II and XI

(35.40) and cluster I and XII (33.76) which indicates that genotypes included in these clusters are genetically diverse and may give rise to high heterotic response in early generation and transgressive segregantes in advance generations, if utilized repeatedly in the crossing programme, which in turn may release hidden variability by breaking tight linkages, if any, as reported by Thoday (1960).

Table 1 Grouping of 100 germplasm + checks of linseed in 12 clusters

Cluster	Genotypes	No. of Genotypes
I	1795, 421 × RR-9, 491	03
II	1636, 1541, 1206, 1045	04
III	191×RR-9A, 191×RR-9/2, 302-63-32, 164/1, 121×RR-9-1, 169R-53-2, 149T-126-1, 1C-58-2676, 63/4-34, 39-1	10
IV	A-19-6, A-15-1-2, A-12-1-2, 59/25, 56120, 5629, 5620A, 19×11, A-63, A-51, A-39, A-47	12
V	A-3-1, A-2-1, 1/76, 3/1	04
VI	A-18-1-2, A-6-1-5, 5/37-2/1-61/1, 5/47, 9×12, 477-3/2, A-62, A-44	08
VII	A-4-2-1, 191×RR-9/1, 333-6, A/17/1, 104NP(RR)-573, A-61, A-59, 73×RR-9, 19-1-62	09
VIII	5610, 40617/1, 4602, 1937, 448-5, 476 ENG, 473, 474-3/2, 1491	09
IX	A-10-2-2, 11414, 191-RR-9, 18-RR-9, 18-B-1	05
X	A-6, A-4-3-2, A-3, A-1-3, 5630, 5613, 162-OR-3-1, 107-DOOMA, LC-2045, 1-60, A-58-A-56, A-52, A-24-1-2, A-4-2-1, A-40, A-42, A-43, 11×14, EC 97256, LCK-887, Kiran, T-397, Padmini	24
XI	A-4-3-1-13, 5/47-2/1-10/10, A-23-1-1, 9×14, 9×17, 11×12, 11×17, 12×15, Neelum	09
XII	388, A-49, 9×JBP-1986, Jabalpur12 -1986, 11×13, 23/3, 16×RR-9, Gaurav	08
	Total	105

Table 2 Intra and inter cluster (D and D^2) values among 12 clusters in linseed under salt affected soil

Cluster	I	II	III	IV	V	VI	VII	VIII	IX	X	XI	XII
I	1.15 (1.322)											
II	3.32 (11.022)	1.83 (3.348)										
III	4.03 (16.240)	5.21 (27.147)	2.10 (4.410)									
IV	4.07 (16.564)	4.83 (23.328)	2.36 (5.569)	1.52 (2.310)								
V	3.60 (12.960)	4.25 (18.062)	3.57 (12.744)	3.53 (12.460)	2.29 (5.244)							
VI	4.93 (24.304)	5.55 (30.802)	3.41 (11.628)	2.96 (8.761)	3.00 (9.00)	2.22 (4.928)						
VII	3.90 (15.210)	4.95 (24.502)	2.61 (6.812)	2.56 (6.553)	3.15 (9.922)	9.92 (8.526)	1.83 (3.348)					
VIII	3.09 (9.548)	4.13 (17.056)	4.23 (17.892)	3.23 (10.432)	4.26 (18.147)	3.71 (13.764)	3.47 (12.040)	1.89 (3.572)				
IX	3.57 (12.744)	5.13 (26.316)	4.08 (16.646)	4.06 (16.483)	3.66 (13.395)	4.49 (20.160)	3.08 (9.486)	4.32 (18.662)	1.58 (3.422)			
X	4.31 (18.576)	5.50 (3.250)	3.31 (10.956)	1.78 (3.168)	3.91 (15.288)	3.11 (9.672)	2.33 (5.428)	2.83 (8.179)	3.70 (13.690)	1.51 (2.280)		
XI	5.23 (27.352)	5.95 (35.402)	3.47 (12.040)	3.35 (11.222)	4.21 (17.724)	3.31 (10.956)	2.16 (4.665)	4.13 (17.056)	3.96 (15.681)	2.80 (7.840)	1.71 (2.924)	
XII	5.81 (33.756)	6.60 (43.560)	5.06 (25.603)	4.15 (17.222)	5.11 (26.112)	4.65 (21.622)	3.65 (12.673)	4.30 (18.490)	4.92 (24.206)	2.79 (7.784)	2.75 (7.562)	1.72 (2.958)

Figures in parentheses denote D values

References

- Chawla, B.K. and Singh, P. 1984. Genetic divergence in linseed. *Indian Journal of Agricultural Sciences*, **54**(4): 266-268.
- Mahto, J.L., Chaudhary, U. and Singh, S.N. 1995. Stability and genetic divergence in linseed (*Linum usitatissimum* L.) under rainfed situation. *Indian Journal of Agricultural Sciences*, **65**(8): 602-604.
- Mahalanobis, P.C. 1936. A statistical study at Chinese head measurement. *Journal of Asiatic Society Bengal*, **25**: 301-377.
- Moll, R.H. and Stuber, C.W. 1971. Quantitative genetics empirical results relevant to plant breeding. *Advances in Agronomy*, **26**: 277-313.
- Rao, C.R. 1952. *Advanced statistical methods in biometric research*. John Wiley and Sons, Inc., New York.
- Singh, R.K. and Choudhary, B.D. 1977. *Biometrical methods in quantitative genetic analysis*. Kalyani Publishers, New Delhi.
- Thoday, J.M. 1960. Effect of disruptive selection III coupling and repulsion. *Heredity*, **14**: 35-39.

Short communication

Genetic diversity and variability in sesame, *Sesamum indicum* L.

S.R. Kumhar and Z.S. Solanki

Agricultural Research Station, Rajasthan Agricultural University, Mandor-342 304, Jodhpur, Rajasthan

(Received: November, 2008; Revised: August, 2009; Accepted: September, 2009)

Abstract

Genetic divergence was studied by growing 82 genotypes of sesame, *Sesamum indicum* L. at Agricultural Research Station, Mandor, Jodhpur. The mean sum of squares were significant for all the characters studied except oil content, indicating the presence of variability. Characters like seed yield, primary branches/ plant, capsules/ plant and plant height to first capsule exhibited heritability coupled with high genetic advance revealing that these characters were controlled by additive gene action. The hierarchical cluster analysis indicated the presence of considerable genetic divergence among the genotypes. The genotypes were grouped into eight clusters using Ward's minimum variance method. The inter-cluster Euclidean² distance was maximum between cluster V and VIII followed by cluster V and VII and cluster II and V which indicated that the genotypes included in these clusters will give high heterotic response and thus better segregants. The maximum cluster means were revealed by cluster V for seed yield, plant height and number of capsules/ plant, cluster II for number of primary branches/plant and cluster III for test weight, while cluster VIII showed minimum cluster means for plant height to first capsule, days to 50% flowering and maturity. Among the eight characters studied seed yield contributed the most (38.9%) towards the divergence of genotypes.

Keywords: Sesame, Euclidean² distance analysis, genetic divergence, genetic variability

Bold and shining white seeded sesame also referred as confectionary sesame gaining much more importance in recent years in view of its export potential. Genetic variability and divergence are of great interest to the plant breeders as these play a vital role in framing a successful breeding programme. The objective of the research was to assess quantum of variability, heritability, genetic advance and genetic diversity in available breeding lines so that breeding efforts can be initiated to evolve high yielding sesame varieties.

The materials for present investigation comprised of 78 newly developed white seeded genotypes selected from F₆

generation of different crosses and four standard check varieties of sesame, *Sesamum indicum* L. A set of 82 entries/varieties of sesame was grown in Randomized Block Design with three replications during *kharif* 2006 at research farm of Agricultural Research Station, Mandor, Jodhpur. Each genotype was sown in five rows of 4 m length following crop geometry of 30 × 15 cm. The data were recorded on five competitive plants taken from each replication for plant height (cm), plant height to first capsule (cm), number of primary branches/plant and number of capsules/plant. The characters like seed yield (kg/ha), test weight (g), oil content (%), days to 50% flowering and maturity were recorded considering whole plot in each replication. The data were subjected to analysis of variance as suggested by Panse and Sukhatme (1985). Genotypic and phenotypic coefficients of variance were estimated as per Burton (1952). Broad sense heritability and genetic advance (GA) as per cent of mean at 5% selection intensity were estimated as per Johnson *et al.* (1955). Clustering was performed by procedure of Ward's minimum variance method (Ward, 1963).

The analysis of variance revealed significant differences among the genotypes for all the characters except oil content, hence not included for further analysis. Wide variability was observed for seed yield i.e., from 250 (RMT 207) to 891 kg/ha (RMT 240) (Table 1). The genotype RT 54 was found earliest in days to 50% flowering (38) and maturity (71 days). The range of plant height was from 95 (RT 54) to 145 cm (RMT 201), plant height to first capsule from 47 (RMT 243) to 93 cm (RMT 194), number of primary branches/plant from 1.0 (RMT 207) to 4.4 (RMT 162), number of capsules/plant from 28 (RMT 207) to 62 (RMT 194) and test weight from 2.44 (RMT 192) to 2.95 g (RMT 236). The high magnitudes of phenotypic as well as genotypic coefficient of variation for seed yield, primary branches/plant and capsules/plant indicated the presence of ample amount of variation for these characters. The high heritability (75-95%) combined with high genetic advance as per cent of mean for seed yield, primary branches/plant, capsules/plant and plant height to first capsule revealed that these characters were controlled by additive gene action, suggesting that selection for these traits would be effective for crop improvement. These

results are in agreement with those reported by earlier workers (Solanki and Gupta, 2004; Babu et al., 2004; Anuradha and Reddy, 2005).

A hierarchical cluster analysis of Ward's minimum variance method produced a dendrogram showing successive fusion of individuals which clearly partitioned the genotypes into eight clusters. The genotypes within each cluster were closer to each other than the genotypes grouped in to different clusters. The maximum intra-cluster distance was observed in cluster VII (13.9) followed by cluster VIII (12.0) and cluster VI (8.9) indicating wide genetic variability within the genotypes of these three clusters (Table 2). The highest inter-cluster distance was observed between cluster V and VIII (94.4) followed by cluster V and VI (69.7) and cluster II and cluster V (58.6), suggesting wide diversity between genotypes of these clusters. Therefore, genotype belonging to these clusters may be used in hybridization programme for improvement of sesame and may give better segregants. The least inter-cluster distance was observed between clusters I and II (8.6) indicating close relationship between the genotypes of these two clusters.

The diversity was also supported by the appreciable amount of variation among the cluster means for different characters (Table 3). The highest cluster mean revealed by cluster V for seed yield, plant height and number of capsules/plant, cluster II for primary branches/plant and cluster III for test weight, while cluster VIII showed minimum cluster means for plant height to first capsule,

days to 50% flowering and maturity. These results showed that different clusters genotypes were superior for different characters and genotypes much in use of these characters would offer a good scope of improvement of sesame through rational selection.

Amongst the characters, seed yield contributed maximum towards genetic divergence (38.9%) followed by days to maturity (11.7%), primary branches/plant (11.5%), test weight (10.6%) and number of capsules/plant (10.2%) while characters plant height to first capsule (5.1%) and days to 50% flowering (7.8%) contributed least to genetic divergence (Table 3). These results are in conformity with those reported by Rao (2004 and 2006), and Anuradha and Reddy (2005).

Since varieties with narrow genetic base are more vulnerable to diseases and adverse climatic conditions, therefore, the availability of the genetically diverse genotypes for hybridization programme becomes more important. In the present study the maximum inter-cluster distance observed between cluster V and VIII, cluster V and VII, and cluster II and V, and crosses among genotypes viz. RMT 194, RMT 241, RMT 242, RMT 256, RMT 184, RMT 201, TKG 22 and RT 54 would be resulted into transgressive segregation. Since, seed yield, days to maturity, primary branches/plant, test weight, and capsules/plant contributed maximum towards the divergence, direct selection of these traits help in crop improvement.

Table 1 Estimate of genetic parameters of seed yield and its component traits in sesame

Characters	Mean	Range		Coefficient of variation (%)		Heritability (%)	Genetic advance as % of mean
		Min.	Max.	GCV	PCV		
Seed yield (kg/ha)	571	250	891	25.4	26.0	95	50.9
Days to 50% flowering	42	38	45	3.2	3.6	82	6.0
Days to maturity	80	71	87	2.5	2.7	88	4.9
Plant height (cm)	127	95	145	5.4	6.2	75	9.7
Plant height to first capsule (cm)	59	47	93	8.9	10.3	76	16.0
Primary branches/plant	2.9	1.0	4.4	18.7	20.4	84	35.4
No. of capsules/plant	45	28	62	13.9	15.2	84	26.2
Test weight (g)	2.66	2.44	2.95	3.7	4.1	83	7.0

Table 2 Intra (diagonal) and inter cluster distance among eight clusters in sesame

Cluster	Cluster I	Cluster II	Cluster III	Cluster IV	Cluster V	Cluster VI	Cluster VII	Cluster VIII
Cluster I	6.6	8.6	10.4	15.1	41.5	13.4	27.3	34.8
Cluster II		5.7	14.9	19.9	58.9	11.4	22.5	28.8
Cluster III			7.2	12.9	38.9	14.4	30.4	38.7
Cluster IV				6.7	36.3	17.4	34.2	23.5
Cluster V					0.0	57.8	69.7	94.4
Cluster VI						8.9	16.4	30.2
Cluster VII							13.9	46.8
Cluster VIII								12.0

Table 3 Cluster means in different clusters and per cent contribution for different characters in sesame

Characters	Cluster I	Cluster II	Cluster III	Cluster IV	Cluster V	Cluster VI	Cluster VII	Cluster VIII	Contribution to diversity (%)
Seed yield (kg/ha)	611	535	768	681	846	493	328	547	38.9
Days to 50 % flowering	43	43	42	40	40	42	42	39	7.7
Days to maturity	79	80	79	77	78	81	83	75	11.8
Plant height (cm)	128.0	127.8	132.4	118.5	132.3	126.2	129.8	105.8	4.2
Plant height to first capsule (cm)	61.5	56.7	59.9	63.0	92.7	57.8	61.2	48.8	5.2
Primary branches/plant	3.2	3.3	3.0	2.8	2.8	2.7	1.7	2.8	11.5
Capsules/plant	51	45	48	48	62	39	36	38	9.8
Test weight (g)	2.65	2.55	2.79	2.75	2.78	2.72	2.64	2.52	10.5

References

- Anuradha, T. and Reddy, G.L.K. 2005. D² analysis in sesame (*Sesamum indicum* L.). *Journal of Oilseeds Research*, **22**: 174-175.
- Babu, J.S., Reddy, C.D.R. and Reddi, N.S. 2004. Studies on genetic variability in sesame (*Sesamum indicum* L.). *Annals of Agri Bio Research*, **9**:7-11.
- Burton, G.W. 1952. Quantitative inheritance in grasses. In: *Proceedings 6th International Grassland Congress*, **1**:227-233.
- Johnson, H.W., Robinson, H.F. and Comstock, R.E. 1955. Estimates of genetic and environmental variability in soybean. *Agronomy Journal*, **47**: 314-318.
- Panase V.G. and Sukhatme, P.V. 1985. *Statistical methods for agricultural workers*. ICAR, New Delhi, pp. 381.
- Rao, S.V.S.G. 2004. Genetic divergence in sesame, *Sesamum indicum* L. *Journal of Oilseeds Research*, **21**: 336-337.
- Rao, S.V.S.G. 2006. D² statistics in sesame (*Sesamum indicum* L.). *Crop Research*, **31**: 261-263.
- Solanki, Z.S. and Gupta, D. 2004. Genetic divergence, heritability and genetic advance in sesame, *Sesamum indicum* L. *Journal of Oilseeds Research*, **21**: 333-335.
- Ward, J.H. 1963. Hierarchical grouping to optimize an objective function. *Journal of the American Statistical Association*, **58**: 236-244.

Rajasthan Til 346 : A high yielding white and bold seeded sesame, *Sesamum indicum* L. variety for National Zone-I of India

S.R. Kumhar, Z.S. Solanki, T.S. Rajpurohit, M.M. Sundria and M.S. Chandawat

Agricultural Research Station, Rajasthan Agricultural University, Mandor-342 304, Jodhpur, Rajasthan

(Received: June, 2009; Revised: August, 2009; Accepted: September, 2009)

Abstract

A white and bold seeded variety of sesame 'RT 346' was found suitable for cultivation in National Zone-I of India i.e., Rajasthan, Haryana, Punjab, Himachal Pradesh, Gujarat, Western U.P., adjoining parts of Maharashtra and Karnataka.

Keywords: Sesame, RT 346, white and bold seeded variety

Sesame is a rich source of protein (24%) and carbohydrates (15%) in addition to good source of quality oil (50%). India is the world leader with the area of 17.5 lakh ha and production of 6.7 lakh tonnes (Anonymous, 2008a). In the country, Rajasthan among the sesame growing states contributes more than 18% in area and production, respectively (Anonymous, 2008a and b).

With a view to develop shining white and bold seeded sesame varieties, breeding efforts were initiated at Agricultural Research Station (Rajasthan Agricultural University), Mandor, Jodhpur. A cross was attempted between RT 127 and HT 24 in the year 1996 and segregating material was handled through pedigree method. As a result of these efforts, the culture, RT 346, was isolated and evaluated for its production potential in *kharif* seasons during 2003 to 2004 along with existing checks of sesame varieties in Randomized Block Design

with three replications at different locations of the region. The material was tested in PET (Preliminary Evaluation Trial) and SVT (State Varietal Trial) consisting of 5 m length plot with variable row numbers, i.e., 4 and 6, respectively. The spacing of 30 cm between the rows and 15 cm distance plant to plant was maintained. The agronomic package of practices recommended for the region were followed to reap the good yield.

The RT 346 recorded the mean seed yield (average of 26 trials) of 733 kg/ha as against 611 kg/ha of Pragati and TKG 22 with a yield improvement of 20% over both the check varieties (Table 1).

The morphological features of RT 346 and two check varieties are presented in Table 2. Data indicated that RT 346 is earlier in maturity (83 days) with average plant height of 101.3 cm., high oil content (50%) and more number of capsules/plant (51) with higher oil yield (363 kg/ha) over the checks. The genotype has shining white seed colour suitable for export purpose. Newly selected variety also showed resistance to phyllody and leaf curl viruses and moderately resistance to *Macrophomina* stem and root rot and *Alternaria* and *Cercospora* leaf spot, and capsule borer. In Adaptive trials, RT 346 recorded 19.4% higher seed yield over best adapted variety RT 127 (360 kg/ha) at farmers' fields (data not presented).

Table 1 Summary seed yield (kg/ha) data of coordinated varietal trials

Year of testing	No. of locations	RT 346	Pragati (ZC)	TKG 22 (NC)	Per cent increase over checks	
					Pragati	TKG 22
2005	7	668	568	640	17.6	4.4
2006	8	702	585	543	20.0	29.3
2007	11	797	657	641	21.3	24.3
Mean	-	733	611	611	20	20
Frequency in the top significant group (Pooled for 3 years)			14/26	10/26	5/26	

Table 2 Important characteristics of sesame variety RT 346 compared to checks (2005 to 2007)

Character	RT 346	Check varieties	
		Pragati	TKG 22
Seed colour	White	White	White
Days to 50% flowering	41	42	42
Days to maturity	83	85	83
Plant height (cm)	101.3	98.9	100.2
No. of branches/plant	3.4	3.2	3.5
No. of capsules/plant	51	45	47
1000 seed weight (g)	2.9	2.8	2.9
Oil content (%)	50.0	48.6	49.2
Oil yield (kg/ha)	363	298	309
Macrophomina stem and root rot (%)	9.0	17.1	7.6
Antigastra (% damage)	8.1	14.4	8.1

Looking to its consistence superior performance under rainfed conditions, variety RT 346 was identified in the meeting held during IX Annual Workshop of Sesame and Niger at Nagpur and subsequently released by Central Sub-Committee on Crop Standards, Notification and Release of Variety Vide notification S.O. 449 (E) dated 11.2.2009 for the commercial cultivation in hot and arid ecosystem areas of the National Zone-I, comprising of Rajasthan, Haryana, Punjab, Himachal Pradesh, Gujarat,

Western U.P. and adjoining areas of Maharashtra and Karnataka.

References

- Anonymous, 2008a.** <http://faostat.fao.org/site/612/default.aspx#ancor>.
- Anonymous, 2008b.** <http://www.rajasthankrishi.gov.in/Departments/Agriculture/RBDistCroppt.asp?var=1>.

Short communication

Effect of biofertilizer on sunflower, *Helianthus annuus* L.

Madhurendra, N. Prasad and R.K. Akhuri

Department of Biochemistry, Rajendra Agricultural University, Pusa, Samastipur, Bihar

(Received: May, 2009; Revised: September, 2009; Accepted: October, 2009)

Abstract

The pooled analysed data of year revealed that treatment T₉ (50 % Nitrogen + *Azospirillum* treatment) recorded the high seed yield (1685 kg/ha) followed by T₁₀ (100 % nitrogen + *Azospirillum* + *Azotobacter* seed treatment) seed yield (1656 kg/ha) but difference was at par (Table 1). Same trend of yield was found in treatment T₈ (50 % N + *Azotobacter* seed treatment) and T₃ (100 % N + *Azospirillum* seed treatment) seed yield 1646 kg/ha and 1627 kg/ha, respectively. Lowest seed yield (785 kg/ha) was obtained in T₁ (control).

Keywords: Sunflower, biofertilizer

Sunflower is an exhaustive oilseeds crop, giving good response to application of fertilizers. Continuous use of chemical fertilizers may, in some situation have detrimental effect on soil properties. Therefore to maintain soil fertility and productivity for long, use of organic manures and biofertilizers is very essential. Thus any attempt made to increase productivity of sunflower can satisfy the need of high oil production. Seed inoculation with *Azospirillum* and use of farm yard manure to sunflower increased its yield as well as oil content (Ram et al., 1992). Keeping these facts in view, an experiment was

conducted to study the effect of application of biofertilizers on yield and quality of sunflower. A field experiment was conducted during *kharif* season of 2008 with 11 treatments, 3 replications in a Randomized Block Design at T.C.A., Dholi centre farm which consisted of control (no nitrogen), 50% and 100% recommended doses of nitrogen (80 kg/ha) and seed inoculation with biofertilizer, *Azospirillum* and *Azotobacter* alone and in combination and also integration of these fertilizer with former two recommended doses of nitrogen (Table 1). Seed inoculation was done with *Azospirillum* or *Azotobacter* @ 50 g/kg of seed before sowing. *Azospirillum* or *Azotobacter* culture was added to 150 ml starch solution. This mixture was added to 1 kg of seed and mixed thoroughly. The crop cultivar, KBSH-1 was fertilized with a uniform dose of 50kg P₂O₅ and 30kg K₂O/ha in the form of single superphosphate and muriate of potash, respectively. The crop was sown with a spacing of 2.2 m × 2.2 m (7 row). All the cultural practices besides the treatments adopted as per the recommended package of practices. So application of *Azospirillum* and *Azotobacter* separately or jointly with 50% N (of recommended dose of N) can save the 50% nitrogen for higher seed yield of sunflower. These findings are in line with those of Kumari et al. (2004).

Table 1 Seed yield, oil content and oil yield of sunflower as influenced by different biofertilizer treatments

Treatment	Seed yield (kg/ha)	Oil content (%)	Oil yield (kg/ha)
T ₁ No nitrogen (control)	785	40.2	315
T ₂ 50 % recommended dose of nitrogen	1075	42.0	451
T ₃ 100 % recommended dose of nitrogen	1627	39.3	639
T ₄ <i>Azospirillum</i> seed treatment only	815	40.5	330
T ₅ <i>Azotobacter</i> seed treatment only	885	40.2	355
T ₆ <i>Azospirillum</i> <i>Azotobacter</i> + <i>Azotobacter</i>	890	39.6	352
T ₇ 50% Nitrogen + <i>Azospirillum</i> treatment	1600	40.4	646
T ₈ 50% Nitrogen + seed treatment	1646	39.8	655
T ₉ 50% nitrogen + <i>Azospirillum</i> + <i>Azotobacter</i> seed treatment	1685	40.0	674
T ₁₀ 100% Nitrogen + <i>Azospirillum</i> + <i>Azotobacter</i> seed treatment	1656	39.2	649
T ₁₁ 75% Nitrogen + <i>Azospirillum</i> + <i>Azotobacter</i> seed treatment	1608	39.0	627
SEm ±	23	0.5	5
CD (P=0.05)	81	1.7	15

References

Kumari, C.P., La Channa, A. and Satyanarayana, V. 2004. Effect of biofertilizers on seed yield and quality of sunflower, *Helianthus annuus* L. *Journal Oilseeds Research*, 21 (1) : 183-184.

Ram, G., Patel, J. K., Chaure, N.K. and Choudhary, K.K. 1992. Single and combined effect of bio, organic and Inorganic fertilizers on yield of sunflower and soil proportion under rainfed conditions. *Advances in Plant Sciences*, 5(1): 161-167.

Inter-relay cropping of castor in greengram under irrigated condition

R.M. Patel, G.N. Patel and S.S. Solanki

Main Castor-Mustard Research Station, S.D. Agricultural University, Sardarkrushinagar-385 506, Gujarat

(Received: July, 2009; Revised: September, 2009; Accepted: October, 2009)

Abstract

A field experiment was conducted on loamy sand soils during *kharif* seasons of 2006-07 to 2008-09 to study the effect of inter-relay cropping of castor in greengram under irrigated condition. The results showed that sole castor (4030 kg/ha) remaining comparable with castor intercropped with greengram (1:1) with simultaneous sowing (3924 kg/ha) gave significantly higher yield than castor relay intercropped with greengram staggered sowing at 15, 30, 45 DAS of greengram and harvest of greengram. Simultaneous sowing of castor in greengram gave higher castor equivalent yield, net realization and BCR as compared to delayed sowing. Seed yield of castor was decreased with extent in sowing period. Economics of different treatments indicated that the net realization and B : C ratio from both the crops were higher when the castor + greengram were seeded simultaneously. Sowing of castor after harvest of greengram is found not beneficial.

Keywords: Intercropping, castor

Castor is an important non-edible industrial oilseed crop of North Gujarat. During 2007-08 the state occupied the top

position in area (3.58 lakh ha), production (7.08 lakh tonnes) and productivity (1978 kg/ha). However, it is ideally suited for intercropping system by virtue of its drought tolerance, perenniating nature, branching habit and indeterminate penology (Hegde and Sudhakara Babu, 2002). To study the influence of staggered sowing of castor in greengram stand as inter cropping the present study was undertaken.

Field experiment was conducted at the Main Castor-Mustard Research Station, S.D. Agricultural University, Sardarkrushinagar, Gujarat during *kharif* seasons of 2006-07 to 2008-09. The soil of experimental plot was loamy sand in texture with pH 7.8. It was low in organic carbon (0.12%) and available nitrogen (157 kg/ha), medium in available phosphorus (50) and available potash (243). The experiment comprised of six treatments conducted in Randomized Block Design with four replications (Table 1). Castor hybrid GCH-7 and greengram variety GM-4 were selected as test varieties. A recommended dose of fertilizers was applied to both the crops. Greengram was sown at 45 cm row spacing in sole treatment while in intercropping, it was sown at 120 cm row spacing. Castor was grown at 120 cm x 60 cm both as sole and in intercropping.

Table 1 Seed yield of castor and castor equivalent yield (kg/ha)

Tr. No	Seed yield of castor (kg/ha)				Castor equivalent yield (kg/ha)				BCR
	2006-07	2007-08	2008-09	Pooled	2006-07	2007-08	2008-09	Pooled	
1	4039	4068	3521	3876	4039	4068	3521	3876	2.63
2	3963	3962	3589	3838	4707	4838	4378	4641	3.01
3	3499	3405	2872	3259	4339	4451	3722	4170	2.85
4	3362	3171	2766	3100	4262	4332	3730	4108	2.79
5	3195	2485	1732	2471	4099	3578	2726	3468	2.32
6	2729	2154	1382	2088	4344	4303	3421	4023	2.35
SEm±	178	128	181	135	208	140	157	152	-
CD (P=0.05)	535	385	546	425	NS	421	475	431	-
CV (%)	10.3	8.0	13.71	10.1	9.7	6.6	8.8	13.0	-

In this study, the influence of staggered sowing of castor in greengram stand as intercropping has been evaluated. The results showed that sole castor remaining comparable with castor intercropped with greengram (1:1) with simultaneous sowing gave significantly higher seed yield than castor relay intercropped with greengram staggered sowing at 15, 30, 45 DAS of greengram and harvest of greengram. With delay in castor sowing at fortnightly intervals, there was a drastic reduction in seed yield. The data further indicated that maximum castor equivalent yield from both the seasons was recorded when castor was intercropped in greengram (1:1) with simultaneous sowing followed by castor intercropped in greengram (1:1) and sown after 15 DAS of greengram. Economics of different treatments indicated that the net realization and B : C (3.01) ratio from both the crops were higher when the castor + greengram were seeded simultaneously. Similar results were reported by Patel *et al.* (2007), Reddy *et al.* (2008) and Lila Rani (2008). Sowing of castor after harvest of greengram is found not beneficial.

References

- Hegde, D.M. and Sudhakara Babu, S.N. 2002. Castor. In: *A Text Book of Field Crops Production*. (ed. Rajendra Prasad), ICAR, New Delhi, pp. 586-610.
- Lila Rani, P. 2008. Study on castor, *Ricinus communis* L. based intercropping system under rainfed condition. *Journal of Oilseeds Research*, **25** (1): 92-93.
- Patel, K.S., Patel, M.K., Patel, G.N. and Pathak, H.C. 2007. Intercropping in castor, *Ricinus communis* L. under irrigated condition. *Journal of Oilseeds Research*, **24** (1): 121-123.
- Ready, B.N., Raghavaiah, C.V. and Padmavathi, P. 2008. Fertilizer management in intercropping systems involving castor or sunflower under rainfed conditions. *Journal of Oilseeds Research*, **25** (1): 89-91.

Short communication

Study on intercropping of castor, *Ricinus communis* L. under irrigated condition

I. Singh

AICRP on Castor, Agricultural Research Station, Rajasthan Agricultural University, Mandor, Jodhpur-342 304, Rajasthan

(Received: December, 2008; Revised: September, 2009; Accepted: October, 2009)

Abstract

Castor and companion crops were sown at 120 cm x 60 cm row x plant spacing respectively at Mandor, Rajasthan. The castor yield was not reduced significantly by raising intercrops. Sole castor produced maximum seed yield of 3694 kg/ha closely followed by castor + greengram 1:1, castor + moth bean 1:1. The castor seed equivalent yield recorded with castor + greengram 1:1 (4531 kg/ha) was 837 kg/ha higher over sole castor. Corresponding increase due to castor + moth bean 1:2 system was 639 kg/ha. Castor + sesame intercropping system in 1:1 and 1:2 ratios was not found effective as seed equivalent yield, net return and B:C ratio were not influenced remarkably due to adoption of this system.

Keywords: Castor, intercropping, seed equivalent yield

Rajasthan ranks second in terms of castor acreage and production (104.7 thousand ha and 159.4 thousand tonnes, respectively) after Gujarat where intercropping of pulses and short stature crops with castor is very common (Anonymous, 2009). Castor growing areas in Rajasthan are characterized with frequent drought in light sandy soils. In western part of state, continuous availability of electricity using wind mills and good quality water through bore well technology has opened new era for castor cultivation. Therefore, studies on intercropping of castor under irrigated crop culture were planned.

The experiment was conducted at Agricultural Research Station, Mandor (Jodhpur) in three consecutive seasons 2005-06, 2006-07 and 2007-08. The soil of the site was loamy sand having pH 8.2 with low available nitrogen (149 kg/ha), medium available phosphorus (38 kg/ha) and potassium (380 kg/ha). The experiment was laid out in Randomized Block Design with seven treatments and replicated three times (Table 1). Castor crop was sown at 120 and 60 cm spacing (row x plant) in field. In between two rows of castor one or two rows of intercrop (as per treatment) were sown at 30 x 10 cm row x plant spacing. The net plot size was 4.8 m x 4.8 m. Castor as well as all the intercrops were sown in the first fortnight of July every season with the onset of rains. Crop was initially kept

rainfed but after withdrawal of rains, it was irrigated 8 times at 18-20 days interval. The total rainfall received during the seasons 2005-06, 2006-07 and 2007-08 was 227, 254 and 163 mm, respectively. Castor was fertilized with 80 kg N, 50 kg P₂O₅/ha whereas greengram and mothbean according to row ratio were fertilized with 20 kg N and 20 kg P₂O₅/ha. Sesame was fertilized with 25 kg N and 20 kg P₂O₅/ha. In castor, half nitrogen and full phosphorus was applied as basal dose and remaining nitrogen was applied in two equal splits at 35 and 90 days after sowing with irrigation water. Greengram, sesame and moth bean took 80, 85 and 70 days, respectively to mature whereas castor was harvested through 6 pickings at one month interval starting from 90 DAS. Seed yield of component crops were converted into castor equivalent seed yields, considering the prevailing prices of the produce during the year.

The results showed that growth attributes, seed yield and oil content of castor was not influenced significantly by intercropping system providing compatibility of the companion crops in the system. However, the highest seed yield was recorded in sole castor (3694 kg/ha) closely followed by castor + greengram 1:1 (3593 kg/ha) and castor + mothbean 1:1 (3525 kg/ha) system. Giridhari and Giri (1991) obtained the highest seed yield of intercropped castor (1018 kg/ha) at 90 cm with one row of greengram with seed yield of 200 kg/ha. The castor seed equivalent yield recorded with castor + greengram 1:1 and 1:2 systems was significantly more by 22.7 and 21.1%, respectively when compared with sole castor. Similarly, castor + mothbean 1:1 and 1:2 systems also increased castor seed equivalent yield by 16.1 and 17.3%, respectively when over sole castor. The castor seed equivalent yield was not influenced significantly when sesame was intercropped with castor. Sesame as an intercrop did not perform well because during early phase castor and sesame both grow vertically and competed for space and light. Whereas, greengram and mothbean adjusted horizontally in the available space between two rows of castor and produced bumper seed yield. Being legume crop, moth bean and greengram also benefitted castor which remained in the field for 150-160 days after harvest of both these crops. Prasad *et al.* (1993) reported that intercropping system decreased leaching of N, P and

K and increased soil fertility compared with pure stands of castor. Singh and Singh (1988) also reported similar results.

The spikes/plant and 100-seed weight of castor was reduced significantly by 2.8/plant and 2.7 g, respectively when sesame was intercropped with castor in 1:1 ratio. Castor seed yield was found to be reduced when intercropped with sesame, sorghum and maize (Gupta and Rathore, 1993). Economic evaluation reveal that castor + greengram in 1:1 ratio is the best option as it provided maximum net return and B:C ratio. Castor + greengram

1:1 ratio increased net compared to sole castor. Gupta and Rathore (1993) recorded maximum monetary advantage of Rs. 3907/ha from castor + greengram intercropping system followed by castor + blackgram. Castor + mothbean 1:1 and 1:2 are other options which increased the net return over sole castor. Increases in net return due to castor + sesame 1:1 and 1:2 ratios were not remarkable. The B:C ratios recorded with castor + sesame 1:1 and 1:2 systems were reduced to 3.08 and 3.01, respectively compared with maximum ratio of 3.58 observed with castor + greengram in 1:1 ratio.

Table 1 Seed yield, growth and yield attributes of castor as influenced by intercropping under irrigated conditions (Pooled mean of 3 seasons)

Treatment	Plant stand ('000 ha)	Plant height (cm)	Nodes upto primary raceme	100-seed weight (g)	Raceme length (cm)	Spikes/plant	Intercrop mean seed yield (kg/ha)	Castor seed equivalent yield (kg/ha)	Castor seed yield (kg/ha)	Oil content (%)	B:C ratio
Sole castor	13.1	61	15	27	41	18	-	3694	3694	50.3	3.20
Castor + Mungbean (1:1)	13.0	57	14	26	40	18	546	4531	3593	49.5	3.58
Castor + Mungbean (1:2)	13.1	57	13	26	40	16	606	4472	3429	49.7	3.44
Castor + Sesame (1:1)	13.5	66	15	24	38	15	280	3829	3356	48.0	3.08
Castor + Sesame (1:2)	13.1	64	15	25	37	15	321	3870	3332	48.9	3.01
Castor + Mothbean (1:1)	13.5	58	13	26	39	19	573	4287	3525	49.1	3.40
Castor + Mothbean (1:2)	13.4	62	14	26	42	18	629	4333	3491	49.7	3.37
SEm±	0.3	3.8	0.8	0.8	2.1	1.3	-	254	128	0.9	-
CD (P=0.05)	NS	NS	NS	2.3	NS	3.8	-	NS	NS	NS	-
CV (%)	3.2	14.2	10.3	1.2	9.6	3.6	-	13.5	13.8	1.2	-

References

Anonymous. 2009 Project Director's Report, All India Coordinated Research Project on Castor, Directorate of Oilseeds Research, Rajendranagar, Hyderabad-500030. p.32.

Giridhari, K. and Giri, G. 1991. Growth and yield of summer castor (*Ricinus communis*) and greengram (*Phaseolus radiatus*) in intercropping system. *Indian Journal of Agricultural Sciences*, 61(9) : 669-671.

Gupta, J.N. and Radhore, D.N. 1993. Intercropping in castor (*Ricinus communis*) under dryland condition in Rajasthan. *Indian Journal of Agronomy*. 38(2) : 182-186.

Singh, J.P. and Singh, S.P. 1988. Intercropping of mung bean and guar in castor under rainfed conditions. *Indian Journal of Agronomy*, 33(2) : 177-180.

Prasad, S.N., Ratan Singh, Prakash, C., Verma, B. and Singh, R. 1993. Effect of conversion measures on erosion, soil fertility and yield of castor (*Ricinus communis*). *Indian Journal of Agricultural Sciences*, 63(1) : 47-49.

Short communication

Efficacy of new insecticides against mustard aphid, *Lipaphis erysimi* (Kalt.)

S.S. Dhaka, Gaje Singh, Y.P.S. Malik and A. Kumar

Department of Entomology, Sardar Vallabhbhai Patel University of Agriculture and Technology, Meerut-250 110, UP

(Received: January, 2009; Revised: September, 2009; Accepted: December, 2009)

Abstract

Efficacy of newer insecticides was studied against mustard aphid, *Lipaphis erysimi* (Kalt.) and their effect on *Coccinella septempunctata* was also recorded at Pilibhit (UP) during 2007-08 on Indian mustard. The insecticides used are acetamiprid 20 SP (125 g/ha) and imidacloprid 17.8 SL (150 ml/ha). Acetamiprid 20 SP (125 g/ha) proved as the best insecticide followed by acephate, thiamethoxam, imidacloprid, profenofos, dimethoate and oxydemeton methyl for the management of aphids. Thiamethoxam was found as the safer insecticide to the coccinellids.

Keywords: *Brassica juncea*, *Lipaphis*, *Coccinellid*, efficacy

About three dozen insect pests have been found infesting mustard crop in India of which the mustard aphid, *Lipaphis erysimi* (Kalt.) is the key pest (Upadhyay *et al.*, 1999). In spite of having certain limitations, the use of insecticides still remains the practically and economically viable solution to combat the aphids. The use of older insecticides causes sudden decrease in the number of natural enemies also. Therefore, the present study was undertaken to test some newer insecticides against

mustard aphid and their safety to its predators on mustard crop.

An experiment was conducted at farmers' field in the Terai region of Uttar Pradesh in Pilibhit district during *rabi*, 2007-08 in Randomized Block Design with three replications in a net plot size of 15 m². Seven insecticides and a control (untreated) were tested on the crop, Indian mustard. Insecticides were sprayed once when the aphids reached economic threshold level (ETL 28/10 cm shoot). Mortality of aphids and coccinellids was recorded after 1, 2, 3, 7 and 10 days of spray.

All treated plots had drastic reduction in aphid population as compared to untreated control (Table 1). Acetamiprid was the most effective insecticide. Ali and Khan (2008) also found imidacloprid more effective than dimethoate and oxydemeton methyl against aphids. Thiamethoxam was the safest insecticide against coccinellids followed by acetamiprid and imidacloprid. Oxydemeton methyl and dimethoate caused 100% mortality of predators. Present results are in agreement with Kannan *et al.* (2004). Highest C:B ratio of 2.15 was found with acetamiprid followed by acephate and thiamethoxam.

Table 1 Effect of various insecticides against mustard aphid, *Lipaphis erysimi* (Kalt.)

Treatment	dose/ha	Before spray	Mean number of aphids/shoot					No. of lady bird beetles/shoot (10 days after spray)	Yield (kg/ha)	C:B ratio (Rs.)
			Days after spray							
			1	2	3	7	10			
Thiamethoxam 25 WDG	100 g	173	112.3 (34.9)	47.7 (72.4)	14.6 (91.8)	5.3 (96.9)	0.0 (100.0)	5.7 (-21.4)	1691	2.08
Profenofos 50 EC	1000 ml	168	123.6 (26.0)	50.7 (69.7)	21.3 (87.2)	9.67 (94.2)	4.3 (97.4)	0.0 (100.0)	1449	1.77
Imidacloprid 17.8 SL	150 ml	179	119.3 (33.2)	48.3 (72.9)	13.3 (92.5)	3.7 (97.9)	0.0 (100.0)	5.3 (0.0)	1657	2.06
Dimethoate 30 EC	1000 ml	160	114.6 (28.18)	53.7 (66.4)	21.6 (86.4)	8.3 (94.7)	4.5 (97.1)	0.0 (100.0)	1319	1.61
Acetamiprid 20 SP	125 g	159	108.6 (31.5)	46.3 (70.1)	12.3 (92.2)	4.3 (97.2)	0.0 (100.0)	6.3 (0.0)	1730	2.15
Oxydemeton methyl 25 EC	1000 ml	165	128.6 (21.8)	64.7 (60.7)	23.6 (85.6)	11.6 (92.9)	4.7 (97.2)	0.0 (100.0)	1246	1.52
Acephate 75 SP	500 g	167	107.6 (35.4)	44.7 (73.2)	8.3 (95.0)	2.3 (98.6)	0.0 (100.0)	6.3 (-11.6)	1723	2.13
Control	-	174	195.3 (11.8)	219.3 (-25.6)	279.3 (-59.9)	311.3 (-78.2)	389.7 (-123.1)	10.3 (-121.2)	873	1.10
CD (P=0.05)	-	NS	15.5	16.1	16.1	20.7	19.4	5.1	291	

Figures in parentheses are per cent mortality.

References

- Ali, Haider and Safiq Ansari. 2008. Efficacy of insecticides against *Lipaphis erysimi* on mustard crop. *Journal of Entomology Research*, 32(1) : 45-47.
- Kannan, M., Uthamasamy, S. and Mohan, S. 2004. Impact of

insecticides on sucking pests and natural enemy complex of transgenic cotton. *Current Science*, 86(5) : 726-729.

- Upadhyay, R., Rajeev, Mukherji, K.G. and Rajak, R.L. 1999. In *IPM System in Agriculture : Oilseeds*, Kailash Balani for Aditya Books Pvt. Ltd., 5:37-39.

Short communication

Efficacy of bioagents and organic amendments against *Macrophomina phaseolina* (Tassi) causing root rot of sesame

S. Usha Rani, R. Udhayakumar and D. John Christopher

Department of Plant Pathology, Faculty of Agriculture, Annamalai University, Annamalai Nagar-608 002, Chidambaram, TN

(Received: December, 2008; Revised: June, 2009; Accepted: July, 2009)

Abstract

Studies indicated that use of *Trichoderma viride* (seed treatment) 4 g/kg, soil application 5 kg/ha with FYM managed *Macrophomina phaseolina* causing root rot in sesame.

Keywords: *Macrophomina phaseolina*, root rot, sesame

Sesame, *Sesamum indicum* L., is the third major oilseed crop of India. Root rot caused by *Macrophomina phaseolina* (Tassi.) Goid, is one of the most serious diseases of sesame causing 42-45% yield losses. Biological control methods have shown promise in managing this pathogen, hence the present study was undertaken.

M. phaseolina was isolated from root rot infected plants of sesame from farmers' field of Vallampadugai Village, Cuddalore District of Tamil Nadu and pure culture was maintained on PDA slants at 4°C. Pure culture of *M. phaseolina* was inoculated on PDA medium in petriplates and incubated at 25±1°C for 5 to 7 days. *M. phaseolina* was multiplied on autoclaved sorghum seeds at 25±1°C for 12 days and the final population was adjusted to maintain 10⁶ cfu/g preparation.

Powdered, sieved and sterilized (1.4 kg/cm pressure, 2 hr for 3 consecutive days) silky clay soil incorporated with FYM @ 12.5 t/ha was filled in earthen pots (30 cm diameter) @ 6 kg/pot and inoculum of *M. phaseolina* was incorporated earlier (50 g/kg of soil and it was irrigated for 7 days and allowed the pathogen to multiply). Seeds were treated with talc based formulation of antagonist @ 4 g/kg of seed and its combination with FYM as soil application was done. The amendment was tested in combination with seed 4 g talk based formulation of *T. viride*/kg of seed and soil application of *T. viride* 5 kg/ha on 45 DAS. For comparison carbendazim @ 2 g/kg was used. Soil without treatment served as control. Three replications were maintained for each treatment.

All the treatments significantly increased both root and shoot length of plant over control at 60, 120 days of observation. Soil application of FYM combined with *T. viride* application (seed + soil of on 45 DAS) registered

highest root and shoot length of 14.03 and 90.50 cm on 120 DAS followed by soil application of FYM + *T. viride* (seed + soil). Similar observations were made in the field trial experiment (Table 2). These findings are in agreement with the findings of Change *et al.* (1986), Ghisalberti *et al.* (1990) and Ousley *et al.* (1994). Incidence of root rot was recorded at regular intervals and the least root rot incidence of 9.24% was observed with FYM combined with *T. viride* (seed + soil + soil on 45 DAS) application, seed treatment with *T. viride* and carbendazim alone reduced the root rot incidence by 25 and 22.50%, respectively over control (51.45%) (Table 1). All the treatments significantly increased the drymatter production of plants over untreated control. The maximum drymatter of 4.02 g/plant was recorded in pots applied with FYM combined with application of *T. viride* (seed and soil + soil on 45 DAS) followed by soil application of FYM + *T. viride* (seed + soil). The untreated control registered the least drymatter production of 3.14 g/plant. Management of *M. phaseolina* using organic amendments along with bioagents was reported by Jamadar and Desai (1996) and Desai *et al.* (1997). Application of FYM combined with *T. viride* (seed + soil + soil on 45 DAS) recorded the least dry root rot incidence and maximum vigour index of sesame.

Table 1 Effect of bioagents and organic amendments against *M. phaseolina* (Pod culture)

Treatment	Root length (cm)	Shoot length (cm)	% root rot incidence	Drymatter production (g/plant)
<i>T. viride</i> (ST) 4 g/kg	10.94	67.70	25.00	3.52
<i>T. viride</i> (SA) 5 kg/ha	12.74	73.11	18.12	3.55
<i>T. viride</i> 4 g/kg (ST) + <i>T. viride</i> 5 kg/ha on 45 DAS	13.00	75.00	15.00	3.71
FYM (SA) + T ₁	13.11	83.40	12.08	3.86
FYM (SA) + T ₂	13.63	87.41	11.04	3.91
FYM (SA) + T ₃	14.63	90.50	9.24	4.02
Carbendazim 2 g/kg (ST)	11.10	67.70	22.50	3.51
Control	10.03	50.90	51.45	3.14
SED±	1.16	0.32	-	-
CD (P=0.05)	2.55	0.71	-	-

Table 2 Effect of bioagents and organic amendments against *M. phaseolina* (Field trial)

Treatment	Root length (cm)	Shoot length (cm)	% root rot incidence	Drymatter production (g/plant)
<i>T. viride</i> (ST) 4 g/kg	10.89	68.01	24.00	3.02
<i>T. viride</i> (SA) 5 kg/ha	12.68	72.21	16.12	3.46
<i>T. viride</i> 4 g/kg (ST) + <i>T. viride</i> 5 kg/ha on 45 DAS	12.52	77.02	14.52	3.59
FYM (SA) + T ₁	13.17	85.61	10.94	3.72
FYM (SA) + T ₂	13.59	89.01	10.96	3.82
FYM (SA) + T ₃	13.39	85.50	9.12	3.92
Carbendazim 2 g/kg (ST)	11.10	67.69	21.22	3.49
Control	9.99	45.99	51.45	3.14
SED±	1.15	0.30	-	-
CD (P=0.05)	2.54	0.69	-	-

References

- Change, Y.C., Baker, R., Kleifeld, O. and Chet, I. 1986. Increased growth of plants in the presence of the biological control agent, *Trichoderma harzianum*. *Plant Disease*, **70** : 145-148.
- Desai, S.A., Malabasari, T.A., Patil, D.R. and Jamadar, M.M. 1997. Non-chemical management of charcoal rot in rabi sorghum (*Sorghum bicolor* L. Moench). *Advances in Agricultural Research in India*, **8** : 147-151.
- Ghisalberti, E.L., Narbey, M.L., Dewan, M.M. and Sivasithamparam, K. 1990. Variability among *Trichoderma harzianum* in their ability to reduce take-all and to produce pyrones. *Plant Soil*, **129** : 287-291.
- Jamadar, M.M. and Desai, S.A. 1996. Evaluation of certain organic substrates and adjuvants for mass multiplication of *T. harzianum* Rifai. *Journal of Bio-control*, **10** : 129-131.
- Ousley, M.A., Lynch, J.M. and Whipps, J.M. 1994. The effects of addition of *Trichoderma* inocula on flowering and shoot growth of bedding plants. *Scientia Horticulturae*, **59** : 147-155.

Short communication

Multiple resistance sources against major diseases and pests of safflower, *Carthamus tinctorius* L.

D.R. Murumkar, D.V. Indi, V.B. Akashe, A.J. Patil and M.A. Gud

AICRP on Safflower, Zonal Agricultural Research Station, 97, Raviwar Peth, Solapur-413 002, MS

(Received: November, 2007; Revised: November, 2009; Accepted: December, 2009)

Abstract

From the above study, it could be concluded that the three genotypes of safflower viz., GMU-5097, GMU-5133 and GMU-7017 recorded moderately tolerant reaction to *Alternaria* leaf spot, resistant reaction to *Fusarium* wilt and tolerant reaction to aphid.

Keywords: Safflower, multiple resistance, pests

Safflower (*Carthamus tinctorius* L.) is an important rabi oilseed crop, damaged by a number of insect pests and diseases of which aphid, *Uroleucon compositae* and *Fusarium oxysporum* f.sp. *carthami* are key pests (Singh *et al.*, 1999 and Anonymous, 2003). *Fusarium* infects the plants right from seedling stage to flowering and can cause upto 100% yield loss under severe incidence. Most of the earlier workers tried to identify resistant sources separately to *Alternaria* leaf spot of safflower (Lukade and Indi, 1985), safflower wilt (Mehetre *et al.*, 2004) and safflower aphids (Rathore and Pathak, 1982). The present investigations were, therefore, undertaken to screen the safflower genotypes for isolating multiple resistant/tolerant sources to *Alternaria* leaf spot, *Fusarium* wilt and aphid.

The most promising 50 safflower germplasm accessions supplied by Project Director, Directorate of Oilseeds Research, Hyderabad were used for screening against *Alternaria* leaf spot, *Fusarium* wilt and aphid for isolating multiple resistant/tolerant sources. The accessions were grown in single row of 4 m length with the spacing of 45 x 20 cm with randomized block design having two replicates. The variety Nira was used as a susceptible check which was sown after every 5th row of the test material and cultivar Bhima was used as a local check. The inoculum load of *Fusarium oxysporum* f. sp. *carthami* in the wilt sick plot was 3-4 x 10⁴ cfu/g of soil. The wilt incidence was recorded at 15, 30, 45, 60, 75 and 90 days after sowing. The results were again confirmed by screening same set of germplasm during 2004-05 in the wilt sick plot.

Subsequently, the same set of germplasm accessions was screened during rabi season of 2003-04 and 2004-05 against *Alternaria* leaf spot and aphid. For *Alternaria* leaf spot, these accessions were screened under natural field

conditions in a single row of 4 m length replicated twice using HUS-305 as a tolerant check, Manjira as a susceptible check, Bhima and Phule Kusuma as local checks. Crop was sown in second fortnight of August to create natural epiphytotics of the disease. The conidial suspension was sprayed on the crop frequently in the evening hours. On the following day, the crop was sprayed with water to provide favourable humidity for infection. Five randomly selected plants from each plot were scored for the disease reaction at 15 days interval using 1-9 scale (Anonymous, 2006). The per cent disease intensity was calculated. Same germplasm accessions were sown in the first fortnight of November for screening against aphid using cultivar CO-1 as a susceptible check, Bhima and Phule Kusuma as local standard checks. The observations on aphid count were recorded on 5 cm apical twig plant-1 on five randomly selected plants.

Out of 50 germplasm only five genotypes recorded tolerance to the *alternaria* disease (disease intensity below 40%) under such a high disease pressure (77.8% on susceptible check, Manjira) of *Alternaria* leaf spot and tolerant check, HUS-305 had only 33.3% disease incidence. These results are in conformity with Indi *et al.* (2004). Out of 50 genotypes screened against wilt in sick plot, five germplasm accessions viz., GMU-1630, GMU-3955, GMU-5097, GMU-5133 and GMU-7017 found free from wilt successively for two years, whereas six accessions viz., GMU-332, GMU-1633, GMU-2020, GMU-3256, GMU-3944, and GMU-7188 recorded moderately resistant reaction to wilt showing less than 10% wilting. The genotypes viz., GMU-174, GMU-1087 and GMU-2936 recorded tolerant reaction (11-20%) to wilt. Rest of the genotypes along with local checks Bhima and Phule Kusuma fell under susceptible to highly susceptible category. The susceptible check Nira which was sown after every fifth row of test material recorded 95.1% average wilting. These results are in conformity with Mehetre *et al.* (2004).

Among the 50 germplasm screened, only two entries (GMU-5134 and GMU-5136) exhibited highly tolerant (1.0) foliage drying grade against aphids, eight entries viz., GMU-4480, 4484, 5097, 5130, 5133, 5137, 5142 and 7017

Multiple resistance sources against major diseases and pests of safflower

possessed tolerant (2.0) foliage drying grade, 26 genotypes were moderately tolerant (3.0), whereas rest of the entries were categorized as susceptible (4.0) to the aphid reaction. The local check Bhima and Phule Kusuma

recorded moderately tolerant (3.0) foliage drying grade, whereas the susceptible check CO-1 exhibited highly susceptible (5.0) foliage drying grade.

Table 1 Screening of safflower genotypes for multiple resistance against pest and diseases (2003-04 and 2004-05)

Entry	Alternaria leaf spot		Fusarium wilt		Aphid		Entry	Alternaria leaf spot		Fusarium wilt		Aphid	
	DI grade (1-9 scale)	PDI	Incidence (%)	Category	Foliage drying grade	Category		DI grade (1-9 scale)	PDI	Incidence (%)	Category	Foliage drying grade	Category
GMU-174	6.6	73.3	20.0	T	3.0	MT	GMU-5133	3.4	37.8	0.0	I	2.0	T
GMU-178	6.6	73.3	33.3	S	3.0	MT	GMU-5134	6.6	73.3	33.3	S	1.0	HT
GMU-332	6.6	73.3	9.1	MR	3.5	S	GMU-5135	5.4	60.0	60.0	HS	3.0	MT
GMU-589	6.2	68.9	80.0	HS	3.0	MT	GMU-5136	6.2	68.9	100	HS	1.0	HT
GMU-1087	7.0	77.8	11.1	T	3.0	MT	GMU-5137	6.2	68.9	64.3	HS	2.0	T
GMU-1251	5.4	60.0	38.5	S	3.0	MT	GMU-5141	5.4	60.0	100	HS	3.0	MT
GMU-1394	5.8	64.4	87.5	HS	4.0	S	GMU-5142	6.6	73.3	53.3	HS	2.0	T
GMU-1628	5.4	60.0	28.6	S	3.0	MT	GMU-5143	7.0	77.8	33.3	S	3.0	MT
GMU-1630	6.6	73.3	0.0	I	4.0	S	GMU-5144	5.8	64.4	42.9	S	4.0	S
GMU-1633	5.8	64.4	5.3	MR	3.5	S	GMU-5145	5.8	64.4	76.5	HS	4.0	S
GMU-1811	7.0	77.8	35.3	S	4.0	S	GMU-5146	5.8	64.4	33.3	S	3.0	MT
GMU-2020	7.0	77.8	8.3	MR	2.5	MT	GMU-5147	3.4	37.8	94.4	HS	3.0	MT
GMU-2130	5.8	64.4	36.4	S	3.0	MT	GMU-5149	5.0	55.5	25.0	S	4.0	S
GMU-2775	7.4	82.2	100.0	HS	3.0	MT	GMU-6299	5.8	64.4	100	HS	3.0	MT
GMU-2936	6.2	68.9	14.3	T	3.0	MT	GMU-6401	6.6	73.3	69.2	HS	3.0	MT
GMU-3256	5.8	64.4	6.7	MR	3.0	MT	GMU-7017	3.0	33.3	0.0	I	2.0	T
GMU-3944	6.2	68.9	8.3	MR	3.0	MT	GMU-7172	5.0	55.6	90.0	HS	3.0	MT
GMU-3955	5.8	64.4	0.0	I	3.5	S	GMU-7152	5.4	60.0	92.9	HS	4.0	S
GMU-4480	6.6	73.3	80.0	HS	2.0	T	GMU-7186	6.6	73.3	54.5	HS	3.0	MT
GMU-4484	6.6	73.3	83.3	HS	2.0	T	GMU-7187	5.8	64.4	75.0	HS	3.5	S
GMU-4608	5.4	60.0	43.8	S	3.0	MT	GMU-7188	3.0	33.3	8.3	MR	4.0	S
GMU-4625	5.4	60.0	33.3	S	3.0	MT	GMU-7191	7.0	77.8	100	HS	4.0	S
GMU-4609	6.2	68.9	100.0	HS	4.0	S	Manjira (SC)	7.0	77.8	-	-	-	-
GMU-5097	2.6	28.9	0.0	I	2.0	T	Nira (SC)	-	-	95.1	HS	-	-
GMU-4627	5.8	64.4	66.7	HS	3.0	MT	CO-1 (SC)	-	-	-	-	5.0	HS
GMU-5130	5.4	60.0	66.7	HS	2.0	T	HUS-305(TC)	3.0	33.3	8.3	MR	-	-
GMU-5131	5.4	60.0	81.8	HS	3.0	MT	Bhima (LC)	3.0	37.8	57.3	HS	3.0	MT
GMU-5132	5.8	64.4	81.8	HS	3.0	MT	P.Kusuma(LC)	3.4	37.8	57.4	HS	3.0	MT

References

Anonymous. 2003. Annual Progress Report, 2002-03 of Rabi Oilseeds Workers' Group Meeting held at UAS, Dharwad (Karnataka), India, Aug. 26-28, pp. 146.

Anonymous. 2006. AICRP on Safflower: Technical programme (2006-07) and guidelines for implementation, Directorate of Oilseeds Research, Rajendranagar, Hydereabad - 500 030, India, pp. 55.

Indi, D.V., Murumkar, D.R., Patil, A.J. and Akashe, V.B. 2004. Screening of safflower germplasm against Alternaria leaf spot under field conditions. Journal of Maharashtra Agricultural Universities, 29 (3) : 344-346.

Lukade, G.M. and Indi, D.V. 1985. Reaction of some safflower varieties to Alternaria leaf spot. Madras Agricultural Journal, 72(4) : 240.

Mehetre, N.M., Kurundkar, B.P., Chavan, R.A. and Bharose A.A. 2004. Screening of safflower genotypes against wilt in water culture technique. Journal of Soils and Crops, 14(1): 100-104.

Rathore, V.S. and Pathak, S.C. 1982. Field screening of safflower varieties for aphid tolerance. JNKV Research Journal, 16 (3): 217-220.

Singh, V., Prasad, Y.G. and Lakshminarayana, M. 1999. Insect pests of safflower and their management. In: IPM System in Agriculture, Vol.5-Oilseeds (Eds. R. K. Upadhyay, K.G., Mukherjee and R.L. Rajak). Aditya Books Pvt. Ltd., New Delhi, India, pp. 552.

Short communication

Seed mycoflora associated with castor, *Ricinus communis* L. and their effect on germination

O. Nagaraja, M. Krishnappa and A.M. Sathisha

Dept. of P.G. Studies and Research in Applied Botany, Kuvempu University, Jnana Sahyadri, Shankaraghatta-577 451, Shivamogga, Karnataka

(Received: May, 2009; Revised: September, 2009; Accepted: October, 2009)

Abstract

Castor (*Ricinus communis* L.) seeds were analyzed for mycoflora and germination. Forty seven fungal species belonging to seven genera were isolated from 185 seed samples of castor. The seed samples were collected from different agro climatic regions of Karnataka during *kharif*, 2006-07. Seeds were also collected from fields (49) farmers (73), retail shops (16) and APMC markets (47). These seeds were subjected for seed health test by standard blotter method (SBM). During the two years 20 samples showed higher incidence of pathogens, and these samples were tested by potato dextrose agar (PDA), water agar (WA) and 2,4-Dichloro phenoxy acetic acid (2,4-D) methods. Fields and farmers samples showed a higher incidence of pathogens than others. Among the samples screened, seeds collected from Bangalore-rural and Mysore districts showed highest incidence of pathogens than other districts. Important fungal species are *Fusarium oxysporum* f.sp. *ricini* (23-100%), *Alternaria ricini* (2-22%), *A. alternata* (0-10%), *Curvularia lunata* (6-32%), *Macrophomina phaseolina* (10-18%) *Sclerotinia sclerotiorum* (4-20%), *Cladosporium herbarum* (2-10%) and *Chaetomium globosum* (4-29%), *Botryodiplodia acerina* (0-5%), *Stachybotrys chartarum* (6-10%), *Aspergillus ochraceus* (1-31%), *A. niger* (2-20%), *A. flavus* (1-28%), *A. versicolor* (5-12%) and *Rhizopus stolonifer* (1-9%). These 15 fungi were predominant on castor beans and were isolated from normal, abnormal, ungerminated and rotted seeds. Among the seed health test methods, standard blotter method (SBM) proved superior over other methods. Heavily infected seeds did not germinate. The associated fungi reduced the germination.

Keywords: Castor seed, mycoflora, germination, seed health test

Castor is one of the important non edible oilseed crop. Castor is cultivated over on area of 20161 ha with a

production 17493 tones and productivity 193 kg/ha in Karnataka (Anonymous, 2006). The important diseases of castor crop are The important diseases are wilt (*Fusarium oxysporum* f.sp. *ricini*), leaf spot and blight (*Alternaria ricini*), *Cercospora* leaf spot (*Cercospora ricinella*), root rot, stem rot and charcoal rot (*Macrophomina phaseolina*), seedling blight (*Phytophthora parasitica*), capsule rot (*Cladosporium oxysporum*), fruit rot and gray rot (*Botrytis ricini*), rust (*Melampsora ricini*), powdery mildew (*Leveillula taurica*), phyllosticta leaf spot (*Phyllosticta bosensis*), angular leaf spot (*Botrytis* sp.), damping off (*Phythium aphanidermatum*) (Rangaswamy and Mahadevan, 2005). The aim of the present study was to investigate the incidence of fungi associated with seeds of castor their frequency and their effect on germination.

Seed samples of castor (185) were collected from different agro climatic regions of Karnataka during *kharif*, 2006 and 2007 (Table 1) fields, farmers, retail shops and APMC markets. The collected seeds were stored in cloth bags at room temperature $25\pm 2^{\circ}\text{C}$ for further investigation. Selected seed samples were subjected for standard blotter method (SBM), Potato dextrose agar (PDA), water agar (WA) and 2,4-Dichloro phenoxy acetic acid (2,4-D) methods.

The seed mycoflora were analysed by following methods

Standard blotter method (SBM): Four hundred seeds were surface sterilized with 0.2% sodium hypochloride or mercuric chloride (HgCl_2) solution for 2 to 3 minutes, at the rate of 10 seeds in each Petriplate lined with moist blotter papers. The plates were incubated at a room temperature $25\pm 2^{\circ}\text{C}$ under alternating cycles of NUV 12 hrs light and darkness for seven days (ISTA, 1976).

Potato dextrose agar (PDA): Four hundred seeds were surface sterilized with 0.2% sodium hypochloride solution for 2 to 3 minutes and plated on sterile Petri plates containing Potato dextrose agar (PDA). Ten seeds per plate were incubated $25\pm 2^{\circ}\text{C}$ for seven days.

Water agar (WA): Four hundred seeds were surface sterilized with 0.2% sodium hypochloride solution for 2 to

Seed mycoflora associated with castor and their effect on germination

3 minutes and plated on sterile glass Petri plates containing (2.5%: 12.5 g in 1000 ml of distilled water) water agar medium. Ten seeds per plate were incubated at 25±2°C room temperature for seven days (ISTA, 1976).

2,4-D method: The blotter paper discs were dipped in 0.2% of 2,4-Dichloro phenoxy acetic acid solution. Ten

seeds were placed equidistantly on moist blotter discs using sterilized forceps in a laminar air flow. Then the plates were incubated at 25±2°C for room temperature for seven days.

Table 1 Seed mycoflora of castor in different seed health test methods during *kharif*, 2006

Place of collection	Methods	Germ (%)	Seed mycoflora (%)														
			Fo	Ar	Aa	Cl	Mp	Ch	Cg	Sc	Ss	Ba	Ao	Af	An	Av	Rs
Jalahalli	SBM	1.0	97.0	19.0	-	-	11.0	2.0	16.0	6.0	4.0	-	19.0	1.0	2.0	-	-
	PDA	33.0	41.0	15.0	10.0	-	12.0	-	12.0	5.0	-	-	-	-	-	14.0	-
	WA	7.0	22.0	-	-	-	-	-	-	-	-	-	2.0	3.0	-	-	1.0
	2,4-D	-	6.0	1.0	-	-	-	10.0	6.0	-	-	-	-	5.0	-	-	-
Yeshvantha pura	SBM	8.0	22.0	16.0	5.0	6.0	-	12.0	-	-	-	-	10.0	16.0	8.0	4.0	-
	PDA	40.0	9.0	10.0	2.0	-	-	-	-	-	-	-	-	11.0	1.0	-	-
	WA	30.0	2.0	6.0	10.0	11.0	-	-	-	-	-	-	4.0	8.0	-	-	-
	2,4-D	-	11.0	9.0	12.0	5.0	-	13.0	-	-	-	-	17.0	4.0	-	-	-
Nelamagala	SBM	40.0	44.0	22.0	-	6.0	14.0	10.0	22.0	-	13.0	5.0	25.0	-	-	-	-
	PDA	27.0	23.0	10.0	5.0	9.0	11.0	6.0	-	-	10.0	11.0	28.0	21.0	23.0	-	25.0
	WA	76.0	9.0	9.0	4.0	-	-	14.0	-	-	-	-	15.0	10.0	11.0	18.0	9.0
	2,4-D	-	17.0	14.0	10.0	-	10.0	19.0	-	-	-	-	51.0	21.0	22.0	19.0	20.0
Dudda	SBM	55.0	44.0	22.0	-	8.0	-	-	1.0	9.0	-	-	22.0	22.0	20.0	11.0	-
	PDA	29.0	19.0	10.0	-	-	-	-	-	-	-	-	-	4.0	-	1.0	9.0
	WA	81.0	9.0	4.0	1.0	-	-	-	-	-	-	-	30.0	9.0	10.0	2.0	-
	2,4-D	-	22.0	10.0	5.0	-	-	8.0	8.0	-	-	-	-	14.0	15.0	3.0	15.0
Anekai	SBM	53.0	23.0	16.0	-	3.0	18.0	5.0	17.0	-	20.0	-	29.0	03	-	-	-
	PDA	31.0	15.0	9.0	-	-	-	-	8.0	-	14.0	2.0	17.0	5.0	6.0	-	11.0
	WA	41.0	2.0	-	-	-	-	-	-	-	-	5.0	2.0	-	1.0	3.0	-
	2,4-D	-	14.0	-	-	-	-	11.0	-	-	-	-	10.0	5.0	3.0	-	10.0
Bannur	SBM	5.0	50.0	18.0	4.0	9.0	-	-	-	-	-	-	18.0	28.0	2.0	12.0	4.0
	PDA	28.0	28.0	-	-	7.0	-	-	-	-	-	-	12.0	11.0	19.0	-	1.0
	WA	62.0	15.0	10.0	8.0	4.0	-	-	-	-	-	-	-	17.0	10.0	-	11.0
	2,4-D	-	18.0	20.0	5.0	-	-	4.0	-	-	-	-	17.0	23.0	42.0	2.0	-
Thubina kere	SBM	22.0	46.0	28.0	10.0	-	-	-	8.0	10.0	-	-	1.0	3.0	4.0	5.0	1.0
	PDA	27.0	21.0	9.0	11.0	-	-	14.0	-	-	-	-	10.0	14.0	11.0	8.0	13.0
	WA	74.0	12.0	-	9.0	4.0	-	8.0	-	-	-	-	23.0	20.0	10.0	25.0	50.0
	2,4-D	-	26.0	10.0	15.0	-	-	-	-	-	-	-	20.0	11.0	4.0	5.0	10.0
Hebbala	SBM	17.0	65.0	22.0	3.0	22.0	10.0	9.0	29.0	-	3.0	-	23.0	16.0	8.0	-	10.0
	PDA	41.0	51.0	-	-	-	-	2.0	-	-	-	-	13.0	3.0	-	18.0	-
	WA	4.0	20.0	-	-	8.0	11.0	10.0	20.0	-	-	-	9.0	4.0	9.0	-	1.0
	2,4-D	-	21.0	-	-	-	10.0	4.0	-	-	-	-	-	9.0	11.0	10.0	-
Chikka ballapur	SBM	23.0	32.0	11.0	8.0	2.0	-	4.0	4.0	11.0	-	-	2.0	9.0	2.0	4.0	9.0
	PDA	-	10.0	8.0	2.0	1.0	-	-	-	-	-	-	-	14.0	6.0	-	1.0
	WA	2.0	4.0	-	-	-	-	10.0	10.0	-	-	-	1.0	9.0	3.0	5.0	10.0
	2,4-D	-	16.0	11.0	-	-	-	-	-	-	-	-	-	5.0	6.0	4.0	10.0
Mylinali	SBM	2.0	100	21.0	-	8.0	14.0	-	20.0	-	20.0	-	31.0	-	-	-	-
	PDA	17.0	68.0	-	-	-	-	-	-	-	4.0	-	17.0	10.0	6.0	-	14.0
	WA	12.0	11.0	-	-	-	-	-	-	-	-	-	2.0	-	-	-	-
	2,4-D	-	41.0	-	-	-	-	-	-	-	-	-	4.0	12.0	10.0	5.0	-

Table 2 Seed mycoflora of castor in different seed health test methods during kharif, 2007

Place of collection	Methods	Germ %	Seed mycoflora (%)														
			Fo	Ar	Aa	Cl	Mp	Ch	Cg	Ss	Ss	Ba	Ao	Af	An	Av	Rs
Kalledevarapura	SBM	28.0	32.0	17.0	10.0	-	20.0	11.0	22.0	10.0	-	-	29.0	11.0	3.0	13.0	5.0
	PDA	37.0	33.0	15.0	9.0	3.0	17.0	14.0	-	3.0	8.0	-	23.0	15.0	-	12.0	10.0
	WA	11.0	12.0	10.0	9.0	-	-	-	2.0	-	3.0	-	16.0	15.0	-	15.0	9.0
	2,4-D	-	15.0	11.0	15.0	-	-	6.0	17.0	-	-	-	5.0	4.0	4.0	-	-
Donnehalli	SBM	16.0	16.0	11.0	-	-	4.0	-	25.0	-	-	-	19.0	14.0	-	10.0	-
	PDA	41.0	30.0	6.0	-	-	-	-	25.0	-	-	-	25.0	6.0	-	7.0	-
	WA	18.0	22.0	-	13.0	-	12.0	1.0	-	-	-	-	17.0	4.0	6.0	8.0	-
	2,4-D	-	10.0	-	9.0	-	-	-	9.0	-	-	-	12.0	6.0	-	-	10.0
Arasikere	SBM	25.0	37.0	25.0	11.0	3.0	18.0	10.0	22.0	8.0	-	-	19.0	7.0	5.0	13.0	7.0
	PDA	25.0	28.0	-	4.0	-	12.0	-	24.0	1.0	-	4.0	11.0	11.0	6.0	-	1.0
	WA	33.0	21.0	7.0	11.0	-	18.0	-	32.0	-	-	-	8.0	4.0	-	4.0	5.0
	2,4-D	-	-	1.0	7.0	-	-	-	27.0	-	-	-	6.0	12.0	-	-	9.0
Nelamagala	SBM	35.0	31.0	20.0	-	5.0	18.0	-	32.0	14.0	-	-	15.0	10.0	13.0	20.0	15.0
	PDA	34.0	28.0	19.0	9.0	-	-	-	-	3.0	-	-	11.0	-	10.0	-	10.0
	WA	29.0	12.0	31.0	18.0	-	13.0	-	52.0	-	8.0	-	20.0	41.0	18.0	22.0	13.0
	2,4-D	-	12.0	-	17.0	-	10.0	-	-	-	-	-	10.0	24.0	-	8.0	6.0
Jagalur	SBM	15.0	37.0	5.0	11.0	-	-	-	-	-	-	-	15.0	7.0	5.0	13.0	7.0
	PDA	25.0	28.0	-	-	-	12.0	-	24.0	-	-	-	11.0	11.0	6.0	-	2.0
	WA	33.0	21.0	7.0	11.0	-	7.0	-	25.0	-	-	-	8.0	4.0	-	4.0	5.0
	2,4-D	-	-	-	7.0	-	-	-	27.0	-	-	-	6.0	12.0	-	-	9.0
Vajarahalli	SBM	42.0	27.0	15.0	4.0	-	19.0	-	19.0	16.0	-	-	14.0	10.0	-	-	-
	PDA	34.0	42.0	16.0	-	12.0	-	-	-	6.0	-	-	5.0	3.0	6.0	7.0	-
	WA	37.0	18.0	22.0	16.0	-	12.0	22.0	28.0	-	-	-	11.0	21.0	12.0	-	-
	2,4-D	-	9.0	12.0	13.0	-	3.0	16.0	31.0	-	-	-	18.0	7.0	17.0	13.0	-
Nandhipura	SBM	36.0	58.0	19.0	6.0	-	23.0	-	28.0	15.0	-	6.0	16.0	32.0	-	-	8.0
	PDA	19.0	23.0	15.0	10.0	-	-	12.0	3.0	-	-	-	11.0	-	-	-	8.0
	WA	15.0	18.0	0.0	17.0	-	10.0	-	-	-	-	-	-	-	-	-	9.0
	2,4-D	-	-	8.0	13.0	-	11.0	-	-	-	-	-	-	-	8.0	-	-
Kaidale	SBM	28.0	21.0	5.0	16.0	-	28.0	9.0	8.0	-	-	-	16.0	9.0	7.0	12.0	6.0
	PDA	21.0	19.0	-	9.0	-	-	-	22.0	-	-	-	-	3.0	-	-	6.0
	WA	18.0	21.0	-	8.0	-	9.0	-	-	-	-	-	-	-	11.0	-	-
	2,4-D	-	-	-	-	-	-	-	-	-	-	-	-	8.0	18.0	-	8.0
Birur	SBM	21.0	18.0	9.0	-	2.0	12.0	-	17.0	-	-	-	-	16.0	13.0	-	-
	PDA	23.0	22.0	-	-	-	10.0	-	24.0	-	-	-	-	12.0	20.0	8.0	-
	WA	16.0	21.0	-	8.0	-	9.0	-	-	-	-	-	-	-	11.0	-	-
	2,4-D	-	18.0	-	7.0	-	7.0	-	31.0	-	-	-	-	8.0	-	-	-
Sondekolia	SBM	10.0	30.0	-	12.0	-	9.0	7.0	-	-	-	-	-	14.0	-	12.0	-
	PDA	32.0	32.0	11.0	12.0	-	-	-	29.0	-	-	-	-	-	6.0	14.0	14.0
	WA	16.0	21.0	-	8.0	-	9.0	-	-	-	-	-	-	-	11.0	-	-
	2,4-D	-	18.0	-	7.0	-	7.0	-	31.0	-	-	-	-	8.0	-	-	-

Data based on 100 seeds for each sample. Each sample in ten replication
 Fo- *Fusarium oxysporum* f.sp. *ricini*
 Ar- *Alternaria ricini*
 Aa- *Alternaria alternata*
 Ao- *Aspergillus ochraceus*
 Ss- *Sclerotinia sclerotiorum*
 PDA: Potato dextrose agar method

Sc- *Stachybotrys chartarum*
 Af- *Aspergillus flavus*
 Ba- *Batrachomyces acerina*
 WA: Water agar method

Rs- *Rhizopus stolonifer*
 Cg- *Chesterium globosum*
 An- *Aspergillus niger*
 Mp- *Macrophoma phaseolina*
 2,4-D: 2,4-D method

Cl- *Curvularia lunata*
 Ch- *Cliosporium herbarum*
 Av- *Aspergillus versicolor*
 SBM: Standard blotter method

Screening of seeds for associated mycoflora: The incubated seeds were observed on eighth day using stereo binocular microscope. The associated mycoflora were recorded in each method and identified with the help of standard guides and manuals.

The analysis of seed-borne mycoflora of castor bean showed occurrence of forty seven species belonging to seven genera. These fungi were isolated from local variety of castor beans. Among the tested samples, the twenty samples shows a higher incidence of fungi. These

Seed mycoflora associated with castor and their effect on germination

samples subjected for potato dextrose agar (PDA), water agar (WA) and 2, 4-Dichloro phenoxy acetic acid (2,4-D) methods and important diseases of corn seed are given in Table 1 and 2.

Seed infection percentage varied from *Fusarium oxysporum* f.sp. *ricini* (23-100%), *Alternaria ricini* (2-22%), *Alternaria alternata* (0-10%), *Curvularia lunata* (6-32%) *Macrophomina phaseolina* (10-18%), *Cladosporium herbarum* (2-10%), *Sclerotinia sclerotiorum* (4-20%) *Botryodiplodia acerina* (0-5%), *Stachybotrys chartarum* (6-10%), *Aspergillus ochraceus* (1-31%), *Aspergillus niger* (2-20%), *Aspergillus flavus* (1-28%), *Aspergillus versicolor* (5-12%) and *Rhizopus stolonifer* (1-9%). During kharif 2007 the percentage seed infection was more in *Fusarium oxysporum* f.sp. *ricini* (21-58%), *Alternaria ricini* (5-25%), *Alternaria alternata* (4-16%), *Curvularia lunata* (3-5%), *Macrophomina phaseolina* (4-23%), *Cladosporium herbarum* (9-11%), *Chaetomium globosum* (8-28%), *Botryodiplodia acerina* (0-6%), *Stachybotrys chartarum* (3-8%), *Sclerotinia sclerotiorum* (3-8%), *Aspergillus ochraceus* (2-29%), *Aspergillus niger* (3-13%), *Aspergillus flavus* (7-16%), *Aspergillus versicolor* (10-20%) and *Rhizopus stolonifer* (5-15%). These 15 fungi were seen

frequently and predominantly in castor beans.

All the seeds associated fungi reduced the germination of castor seeds. The surface sterilized seeds showed a maximum germination percentage. Naik (1994) reported *Fusarium oxysporum* and *Macrophomina phaseolina* significantly reduced germination of castor bean seedlings.

Acknowledgement: The authors are thankful to the Chairman, Department of Applied Botany, for the providing laboratory facilities.

References

- Anonymous.** 2006. Fully revised estimates of principle crops in Karnataka. Directorate of Economics and Statistics, Seshadri Road, Bangalore.
- ISTA.** 1976. International rules for seed testing. *Seed Science and Technology*, **4**: 3-49.
- Naik, M.K.** 1994. Seed borne nature of *Fusarium oxysporum* in castor. *Indian Journal of Mycology and Plant Pathology*, **24**(1): 62-63.
- Rangaswamy, G. and Mahadevan.** 2005. *Diseases of Crop Plants in India*. 4th Ed., Prentice Hall of India Pvt. Ltd., New Delhi-110001.

Short communication

Screening of cold ethyl alcohol extract of *Pongamia pinnata* for insecticidal properties against *Spodoptera litura* Fabricius

Pratibhav Deshmukhe, Ashok A. Hooli and S.N. Holihosur

Karnataka Science College, Dharwad-580 001, Karnataka

(Received: August, 2009; Revised: November, 2009; Accepted: December, 2009)

Abstract

Studies revealed that, cold ethyl alcohol leaf extract (25%) of *Pongamia pinnata* followed by topical application was highly effective against *Spodoptera litura* Fab. Growth and development of *S. litura* was drastically affected by the plant extract (20%) followed by leaf application. The adult emergence was normal for both treatments. For topical application, LC₅₀ and LC₉₀ was found to be 12.28 and 42.53 and for leaf application, 22.50 and 51.21, respectively.

Keywords: Biological insecticide, *Pongamia pinnata*, *Spodoptera litura*, susceptibility

Chemicals of botanical origin used in pest control management programmes may prevent several adverse effects caused due to synthetic insecticides (Gayatri *et al.*, 2003). Present studies involves application of cold ethyl alcohol extract from the leaves of *Pongamia pinnata* to study its effect on the survival rate of *S. litura* at various stages of development, antifeedant activities and deformities in adults.

Freshly laid batches of eggs of *S. litura* were collected from the castor fields at the University of Agricultural Sciences, Dharwad, Karnataka, India. The eggs were sterilized with 0.05% NaOCl and then with distilled water, transferred to a fresh castor leaf and allowed to hatch under laboratory conditions. The larvae were reared in the laboratory in sterilized earthen pots of 2 l capacity with the mouths of the pots covered securely with a clean sterilized white muslin cloth. The larvae were fed on fresh castor leaves, and maintained at 26±1°C a RH of 65±5% and a photoperiod of L10:D14. The laboratory reared pre-starved fourth instar larvae were used as test insects. 0.5, 1, 5, 10, 15, 20, and 25% cold ethyl alcohol extracts of *Pongamia pinnata* leaves were used for the Topical and Leaf application tests.

The shade dried leaves of *Pongamia* were pulverized in to a fine powder in an electric mixer grinder, and sieved through a muslin cloth. This powder was used for preparing solvent extract within 24 h. 500g of the botanical powder was soaked in 500 ml of absolute alcohol (ethyl) and kept overnight. The mixture was stirred with a

magnetic stirrer frequently. The solution was then filtered through an ordinary filter paper. The alcohol was allowed to evaporate from the filtrate at room temperature. The plant extract paste was dissolved in 50 ml of acetone. Solutions of 0.5, 1.0, 5, 10, 15, 20 and 25% concentrations were prepared from this stock using acetone and used within 24 hours of preparation, for trials.

Under each type of application, the first seven groups were treated with the extract of 0.5, 1.0, 5, 10, 15, 20 and 25%. The eighth group was the carrier (acetone control), the ninth, was the absolute control. Each group comprised 20 larvae, kept in 20 separate sterilized earthen pots of 1 liter capacity each and were fed on fresh castor leaves. Extract (2 ml) to be tested was sprayed on the larvae (topical application) and on similar sized castor leaves (and these treated castor leaves were given as food to larvae in leaf application test group.) with a chromatography sprayer. An ordinary cold-air blow dryer was used to hasten the process of drying of the extract on the leaves. Three replications of the test were maintained. Observations were made to assess the total mortality, mortality at the individual stages of development of the insect, feeding deterrence and deformities in the adults. The per cent mortality was corrected using Tukey's honestly significant test. LC₅₀ and LC₉₀ were calculated using Probit analysis (Finney and David, 1971).

In the topically treated group, feeding was totally stopped by the larvae for the initial 30-45 minutes of application, for 20 and 25% concentrations. Faecal pellets were normal for all the seven treatments. In the leaf application test, this behavior persisted for the initial 65-70 minutes in the 15, 20 and 25% botanical treated subgroups. Feeding, when it was resumed, was slow and intermittent in the next four hours. Pellets were extremely moist and pasty in the 20, and 25% leaf applied larvae. This may be because of loss of peritrophic membrane of the digestive tract in the larvae.

In topical application test and leaf application test, maximum mortality was observed at 25% concentration. This indicated knock-down action of the plant extract. No deformities in adults were observed in the topically treated or the leaf application tests. Deformities and death observed at the larval, pre pupal and pupal stages may be

due to inhibition of chitin synthesis as observed in *S. litura* treated with diflubenzuron, a chitin synthesis inhibitor (Nelson and Venugopal, 2006). The LC₅₀ and LC₉₀ were 12.28 and 42.53, respectively. LC₅₀ and LC₉₀ values for leaf application test were 22.50 and 51.21, respectively. Death in the prepupal stage is ascribed to the slow action of plant products on growth stages of insect or due to the enhanced activity of plant constituents when assimilated in insect tissues. The knock-down effect, that resulted in

mortality in the larvae, may be attributed to the action of the natural product by interfering respiration, leading to asphyxiation, or causing muscle paralysis or affecting the nervous system (Patole *et al.*, 2008).

The present study indicated that both topical and leaf applications of cold alcohol extract of *Pongamia pinnata* leaves are highly effective in controlling *S. litura* at various stages of development.

Table 1 Bioefficacy of cold alcohol leaf extract of *Pongamia pinnata* on development profile of the tobacco cutworm, *Spodoptera litura* Fabricius following topical application on fourth instar larvae

Concentration (%)	Larval (IV, V and VI instar) mortality (%) Mean±SE	Pre-pupal (shrunken stage) mortality (%) Mean±SE	Pupal mortality Mean±SE	Deformed adults Mean±SE	Total mortality (%) Mean ±SE
0.5	18.33±1.66 ^e	5.00±0.00 ^{bc}	0 ^a	-	23.33±1.66 ^e
1	25.00±0.00 ^d	8.33±1.66 ^{ab}	1.66±1.66 ^a	-	35.00±0.00 ^{de}
5	28.33±1.66 ^d	10.00±2.88 ^{ab}	1.66±1.66 ^a	-	40.00±2.88 ^{cd}
10	38.33±1.66 ^c	11.66±1.66 ^{ab}	1.66±1.66 ^a	-	51.66±1.66 ^{bc}
15	48.33±1.66 ^b	13.33±1.66 ^a	1.66±1.66 ^a	-	63.33±4.41 ^{ab}
20	61.66±1.66 ^a	8.33±1.66 ^{bc}	0 ^a	-	70.00±2.88 ^a
25	70.00±2.88 ^a	3.33±1.66 ^{ac}	0 ^a	-	73.33±1.66 ^a
Carrier(acetone control)	1.66±1.66 ^f	0 ^c	3.33±1.66 ^a	-	10.00±2.88 ^f
Absolute control	5.00±0.00 ^f	0 ^c	1.66±1.66 ^a	-	6.66±1.66 ^f
SEm±	1.47	1.19	1.35	-	1.65
"F" test	164.94	7.51	0.80 NS	-	78.44
CD (P=0.05)	4.48	3.63	NS	-	5.01

Means followed by the same letters do not differ significantly from each other at P=0.05 (Tukey's honestly significant test)
LC₅₀ = 12.28; LC₉₀ = 42.53

Table 2 Bioefficacy of cold alcohol leaf extract of *Pongamia pinnata* on development profile of the tobacco cutworm, *Spodoptera litura* Fabricius following leaf application on fourth instar larvae

Concentration (%)	Larval (IV, V and VI instar) mortality (%) Mean±SE	Pre-pupal (shrunken stage) mortality (%) Mean±SE	Pupal mortality Mean±SE	Deformed adults Mean±SE	Total mortality (%) Mean ±SE
0.5	18.33±1.66 ^d	10.00±0.00 ^{ab}	0 ^a	-	28.33±1.66 ^d
1	26.66±1.66 ^{cd}	10.00±0.00 ^{ab}	0 ^a	-	36.66±1.66 ^{cd}
5	31.66±1.66 ^{bc}	10.00±2.88 ^{ab}	1.66±1.66 ^a	-	43.33±1.66 ^c
10	38.33±1.66 ^b	16.66±1.66 ^a	8.33±3.33 ^a	-	58.33±1.66 ^b
15	60.00±2.88 ^a	10.00±2.88 ^{ab}	0 ^a	-	70.00±0.00 ^a
20	68.33±1.66 ^a	6.66±1.66 ^{ab}	0 ^a	-	78.33±1.66 ^a
25	68.33±1.66 ^a	0 ^{bc}	0 ^a	-	75.00±2.88 ^a
Carrier(acetone control)	5.00±1.66 ^e	0 ^c	1.66±1.66 ^a	-	6.66±1.66 ^e
Absolute control	1.66±1.66 ^e	0 ^c	0 ^a	-	1.66±1.66 ^e
SEm±	1.90	1.85	1.35	-	1.47
"F" test	172.34	5.61	0.89 NS	-	224.75
CD (P=0.05)	5.77	5.61	NS	-	4.48

Means followed by the same letters do not differ significantly from each other at P=0.05 (Tukey's honestly significant test)
LC₅₀ = 22.50; LC₉₀ = 51.21

References

- Finney and David, J. 1971. *Statistical Method in Biological Assay*. 1st Edition, Griffin, London.
- Gayatri, G., Jesudasan, A. and Wesley, S.D. 2003. Effects of certain plant derived compounds on feeding and growth regulation of *Spodoptera litura* F. *Journal of Applied Zoological Researches*, 14(1) : 121-124.
- Nelson, Jeyarajan, S. and Venugopal, M.S. 2006. Antifeedant and growth disruptive effects of various plant products on *Spodoptera litura*. *Journal of Entomological Research*, 30(2): 93-102.
- Patole, S.S., Chopada, M.Z. and Mahajan, R.T. 2008. Biocidal activities of a common weed *Sphaeranthus indicus* Linn. *Uttar Pradesh Journal of Zoology*, 28(1) : 67-72.

Short communication

Oil quality characteristics and fatty acid composition of castorbean, *Ricinus communis* L. cultivars

Kamal Kumar Verma, P.S. Kendurkar, Madhu Vajpeyi, Ashish Saini and Neeti Singh

C.S. Azad University of Agriculture and Technology, Kanpur-208 002, UP

(Received: December, 2008; Revised: December, 2009; Accepted: December, 2009)

Abstract

Biochemical analysis of 30 castorbean cultivars including 14 varieties, 9 genotypes and 7 hybrids from C.S. Azad University of Agriculture and Technology, Kanpur and Directorate of Oilseeds Research, Hyderabad, India revealed sizable variability in oil quality characteristics.

Keywords: Castorbean, oil quality, fatty acids, ricinoleic acid

Castor (*Ricinus communis* L.) is one of the ancient oilseed crops of the world with diversified uses (Hegde *et al.*, 2003). Castor oil is industrial grade oil containing high content of single fatty acid and ricinoleic acid. The oil content of the castor seeds have been found to vary due mainly to differences in genetic makeup of the varieties. Variability in oil content and fatty acid profile of seeds of castor varieties indicated a great potential of selecting high oil containing varieties for industrial use (Ramos *et al.*, 1984; Lakshminarayana *et al.*, 1984). To explore the potential of these genotypes or hybrids, the present work was undertaken to select varieties/genotypes/hybrids having high oil content and better oil quality characteristics.

Thirty promising cultivars of castor were obtained from Oilseeds Section, C.S. Azad University of Agriculture and Technology, Kanpur and DOR, Hyderabad. Oil content was estimated by Soxhlet method using n-hexane (AOAC, 1974). Oil quality was determined as per methods. Acid value (Cox and Pearson, 1962), saponification value and hydroxyl value by AOAC (1970), iodine value by Jamieson (1943). Oil samples extracted from seeds of different castor genotypes using n-hexane as solvent were converted to methyl esters. Methyl esters of different fatty acids were separated on Gas Chromatograph, Varian Gas Chromatography, model 3900 with data handling workstation and a data processor. Gas Chromatography was carried out using silica packed WCOT capillary column No. CP 7485 following standard procedures. Standard methyl esters were used as reference to characterized different fatty acids. Percentage of fatty acid was based on peak area and retention time recorded by

automatic computerized GC data processor.

Oil content in different castor genotypes significantly varied from 45.9 to 56.4% (Table 1). Amplitude of variability was maximum (10.5%) in varieties. Moreover, differences between 3 groups were found to be significant. Aruna followed by Bhagya and Chandraprabha were identified as high oil containing varieties while Prabha-9801, KC-123 and KC-63 topped the list of genotypes in having higher levels of oil content whereas in hybrids RHC-1, GCH-5 and DCH-32 were identified to be rich in oil. Similar findings on the variability of oil content have also been reported (Nagaraj, 1990). Saponification and iodine values showed very large and significant variations (Table 1). Our results are in agreement with the findings of Patel *et al.* (2004).

The fatty acids profile of the castor seed oil as revealed by Gas Chromatography which can be grouped under three distinct types depending on their peak area and mean fatty acid levels. Palmitic acid, stearic acid, dihydroxy stearic acid, arachidic acid, linolenic acid and arachidonic acid formed a group of minor fatty acids of castor oil, the mean values of which figured below the level of 3% (Table 2). Castor oil possessed little higher concentration of oleic and linoleic acid. The seed oil samples contained the largest concentration of ricinoleic acid (Table 2). Content of oleic and linoleic acid also showed significant variations. Other fatty acids viz., palmitic, stearic, linolenic, arachidic, arachidonic and dihydroxy stearic acids appeared in small concentrations and showed small range of variation (Table 2). Our results in respect of fatty acid composition of different castor cultivar are in agreement with Ramos *et al.* (1984) and Lakshminarayana *et al.* (1984).

Based on the findings generated during the present investigation, it may be concluded that commercially, hybrids are most economically viable for the production of significantly superior oil yield and industrially valuable ricinoleic acid as compared to varieties and genotypes.

Acknowledgement: The authors are grateful to Dr. C. Lavanya, Sr. Scientist, DOR, Hyderabad and Dr. R.V. Sigh, Head, Germplasm Exchange Division, NBPGR, New Delhi for providing some of the hybrids of castorbean for the present study.

Oil quality characteristics and fatty acid composition of castorbean cultivars

Table 1 Variability in oil characteristics of different castorbean cultivars

Entries	Oil content (%)	AV	SV	IV	HV
Varieties					
Balliachayan	53.8	2.57	184.83	87.50	164.3
Bhagya	54.9	2.12	187.00	87.04	165.1
Chandraprabha	54.4	2.67	184.77	86.91	163.8
Aruna	56.4	2.24	186.25	87.63	164.2
Kalpi-6	52.0	2.77	183.40	86.85	163.1
Tarai-4	54.1	2.74	177.38	87.47	162.5
Type-3	51.4	2.88	181.25	83.19	161.7
SKI-272	46.2	2.85	179.27	89.29	162.6
SKI-281	47.9	2.90	175.84	88.88	163.1
SKI-295	47.1	2.81	178.69	88.08	161.7
SKI-299	45.9	3.25	179.86	89.56	162.4
Kranti	50.8	2.94	176.84	88.92	163.8
DCS-9	49.8	2.86	176.61	87.31	160.8
48-1 (Jwala)	49.2	2.65	183.32	88.14	163.4
Mean	51.0	2.73	181.10	87.63	163.0
SEm ±	0.3	0.10	0.46	0.56	0.44
CD (P=0.05)	0.6	0.19	0.92	1.13	0.90
Genotypes					
Prabha-9801	53.5	2.74	179.09	91.77	163.2
KC-1	50.9	2.78	177.72	88.26	160.2
KC-19	51.5	2.89	179.27	89.29	160.9
KC-39	50.9	3.18	181.38	89.80	161.3
KC-40	52.2	2.66	179.86	89.87	160.7
KC-48	52.8	2.72	176.84	87.07	162.5
KC-56	52.6	2.82	179.86	87.87	161.8
KC-63	52.9	2.63	181.38	88.45	160.5
KC-123	53.1	2.83	176.61	86.62	162.0
Mean	52.2	2.80	179.11	88.78	161.4
SEm ±	0.3	0.10	0.46	0.56	0.44
CD (P=0.05)	0.6	0.19	0.92	1.13	0.90
Hybrids					
GCH-4	49.3	2.88	182.22	87.42	160.0
GCH-5	50.2	3.01	179.70	88.37	159.7
GCH-6	49.1	2.65	176.31	90.34	161.5
RHC-1	50.6	3.13	182.75	86.42	162.6
DCH-32	59.5	2.80	180.32	86.50	163.2
DCH-177	48.0	3.00	184.83	87.50	161.4
DCH-519	48.8	2.91	179.86	87.97	162.3
Mean	49.3	2.91	180.85	87.79	160.8
SEm ±	0.1	0.10	0.46	0.56	0.4
CD (P=0.05)	0.6	0.19	0.92	1.13	0.9
SEm ± between (VxG)	0.1	0.03	0.14	0.17	0.1
CD (P=0.05) between (VxG)	0.2	0.06	0.28	0.34	0.3
SEm ± between (VxH)	0.1	0.03	0.15	0.18	0.1
CD (P=0.05) between (VxH)	0.2	0.06	0.30	NS	0.3
SEm ± between (GxH)	0.1	0.03	0.16	0.20	0.2
CD (P=0.05) between (GxH)	0.2	0.07	0.33	0.40	0.3

AV = Acid value; SV = Saponification value; IV = Iodine value; HV = Hydroxyl value

Table 2 Fatty acid composition of different cultivars of castorbean oil

Entries	Palmitic acid	Stearic acid	Oleic acid	Linoleic acid	Linolenic acid	Arachidic acid	Arachidonic acid	Ricinoleic acid	Dihydroxy stearic acid
Varieties									
Balliachayan	2.33	2.89	9.31	9.82	0.93	0.89	-	73.83	-
Bhagya	2.20	2.62	7.68	8.78	0.96	0.66	-	77.10	-
Chandraprabha	1.89	1.94	6.31	8.40	0.73	0.79	-	79.94	-
Aruna	2.52	2.73	9.95	9.90	0.99	0.92	-	72.94	0.05
Kalpi-6	1.95	2.44	8.70	9.07	0.75	1.19	-	75.85	0.05
Tarai-4	2.33	2.90	11.70	9.82	0.96	1.05	-	71.16	0.08
Type-3	1.89	2.53	9.71	9.07	1.16	6.69	-	68.96	-
SKI-272	3.05	3.19	10.12	12.33	1.32	1.29	-	68.58	0.12
SKI-281	3.00	3.22	8.47	11.92	1.15	0.92	-	71.23	0.09
SKI-295	2.67	2.60	7.19	10.38	0.99	0.66	-	75.40	0.11
SKI-299	3.36	3.42	8.13	13.01	1.21	0.95	-	69.92	-
Kranti	2.40	2.59	7.42	10.96	0.98	0.67	-	74.98	-
DCS-9	2.53	2.66	6.79	9.69	0.79	0.46	-	76.99	0.09
48-1 (Jwala)	2.77	2.92	8.73	10.89	0.86	0.56	-	73.18	0.09
Mean	2.49	2.76	8.59	10.29	0.98	1.26	-	73.58	0.08
SEm ±	0.23	0.30	0.37	0.41	0.17	0.24	-	0.31	0.01
CD (P=0.05)	0.46	0.59	0.74	0.83	0.34	0.48	-	0.62	0.03
Genotypes									
Prabha-9801	3.27	3.33	11.61	15.18	1.47	1.64	-	63.50	-
KC-1	2.56	3.07	9.90	10.90	1.03	1.02	-	71.52	-
KC-19	3.18	3.04	10.29	13.07	1.11	1.81	-	67.50	-
KC-39	3.00	2.96	10.79	12.95	1.20	1.27	0.28	67.56	-
KC-40	2.42	2.17	8.70	11.29	1.57	0.97	0.69	72.20	-
KC-48	1.42	1.32	5.85	7.68	0.67	0.71	-	82.35	-
KC-56	2.43	2.82	8.23	10.57	0.79	0.85	-	74.28	0.03
KC-63	2.51	2.64	8.73	11.21	0.99	0.89	0.54	72.46	0.03
KC-123	2.06	2.71	9.49	8.62	0.96	1.13	-	75.01	0.02
Mean	2.54	2.67	9.29	11.27	1.09	1.14	0.50	71.82	0.03
SEm ±	0.23	0.30	0.37	0.41	0.17	0.24	0.22	0.31	0.01
CD (P=0.05)	0.46	0.59	0.74	0.83	0.34	0.48	NS	0.62	NS
Hybrids									
GCH-4	2.02	2.42	7.07	9.11	0.85	0.52	-	77.87	0.14
GCH-5	2.50	3.05	10.76	11.23	0.96	1.14	-	70.27	0.09
GCH-6	3.25	3.85	13.31	13.72	1.50	1.33	-	62.96	0.09
RHC-1	1.99	2.06	5.92	8.19	0.65	0.58	-	80.45	0.17
DCH-32	1.67	1.84	7.00	7.78	0.65	0.68	-	80.28	0.10
DCH-177	2.25	2.39	8.15	9.59	0.79	0.76	-	75.99	0.09
DCH-519	2.28	2.68	8.24	10.25	0.86	0.75	-	74.85	0.09
Mean	2.28	2.61	8.63	9.98	0.89	0.82	-	74.67	0.11
SEm ±	0.23	0.30	0.37	0.41	0.17	0.24	-	0.31	0.01
CD (P=0.05)	0.46	0.59	0.74	0.83	0.34	0.48	-	0.62	0.03
SEm ± between (VxG)	0.07	0.09	0.11	0.12	0.05	0.07	-	0.09	0.007
CD (P=0.05) between (VxG)	NS	NS	0.22	0.25	0.10	NS	-	0.19	0.015
SEm ± between (VxH)	0.07	0.10	0.12	0.13	0.05	0.08	-	0.10	0.006
CD (P=0.05) between (VxH)	0.15	NS	NS	0.27	NS	0.15	-	0.20	0.011
SEm ± between (GxH)	0.08	0.10	0.13	0.15	0.06	0.08	-	0.11	0.007
CD (P=0.05) between (GxH)	0.16	NS	0.26	0.29	0.12	0.17	-	0.22	0.015

Oil quality characteristics and fatty acid composition of castorbean cultivars

References

- AOAC. 1970. *Official Methods of Analysis of Association of Official Analytical Chemists*, 11th Edition, Washington DC, p.477.
- AOAC. 1974. *Official Methods of Analysis of Association of Official Analytical Chemists*, 12th Edition, Washington DC, p.174.
- Cox, H.E. and Pearson, D. 1962. *The Chemical Analysis of Foods*. Chemical Publishing Co. Inc., New York, p.420.
- Hegde, D.M., Sujatha, M. and Singh, N.B. 2003. *Castor in India*. Directorate of Oilseeds Research, Hyderabad.
- Jamieson, G.S. 1943. *Vegetable Fats and Oils*. Reinhold Publishing Corporation, New York, USA, p.262-280.
- Lakshminarayana, G., Pauose, M.M. and Neeta Kumari, B. 1984. Characteristics and composition of newer varieties of Indian Castor Seed and Oil. *Journal of American Oil Chemists Society*, **61**(12) : 1871-1872.
- Nagaraj, G. 1990. Biochemical quality of oilseeds. *Journal of Oilseeds Research*, **7** : 47.
- Patel, M.K., Pathak, H.C., Raj, A.D. and Desai, K.J. 2004. Quality characteristics of some castor (*Ricinus communis* L.) hybrids and varieties. *Journal of Oilseeds Research*, **21**(1) : 210-211.
- Ramos, L.C., Da Silva, Tango, J.S. and Leal, N.R. 1984. Variability for oil and fatty acid composition in castor-bean varieties. *Journal of American Oil Chemist Society*, **61**(12) : 1841-1843.

Short communication

Development of a bullock drawn groundnut, *Arachis hypogaea* L. digger suitable for coastal Orissa

Jayanarayan Mishra

College of Agril. Engineering and Technology, Orissa University of Agril. and Technology, Bhubaneswar-751 008, Orissa

(Received: March, 2008; Revised: August, 2009; Accepted: September, 2009)

Abstract

A ridger type bullock drawn groundnut digger was developed and tested for power requirement, effective field capacity, field efficiency, labour requirement, pod losses, digging of coastal Orissa. The average draft requirement and power requirement of the digger were observed to be 57.0 kgf of 0.39 h.p., respectively which are within the capacity of an average pair of bullock. The maximum digging efficiency of 91.8% was found under an optimum speed of 1.9 km/hr. The performance index was observed as 0.101 considering the quality and quantity of digging along with power requirement for its operation. The performance of this bullock drawn groundnut digger was satisfactory and economical. The use of the digger is within the reach of small and marginal farmers growing groundnut crop on a commercial basis.

Keywords: Groundnut, digger, power requirement, field efficiency, pod losses, digging efficiency

Groundnut (*Arachi hypogaea* L.) is the major oilseed crop of India. Orissa produces 3.78 lakh tonnes of groundnut per year with a production rate of 15.99 q/ha. Major cultivation of groundnut is in the districts of Cuttack, Khurda, Dhenkanal, Jaipur, Ganjam, Sambalpur and Kalahandi (Anonymous, 2008). The socio-economic conditions of farmers of Orissa do not permit them to use tractor, power tillers and their matching implements. Keeping these problems in view, a ridger type bullock drawn groundnut digger was developed and tested at farmers' field in April, 2004.

A ridger shaped digging blade for groundnut having internal angle of 50° was fabricated from 3 mm thick mild steel sheet (Fig. 1). On the blade there was a mild steel plate frame over which the handle was fixed. A beam made of wood of size 245 x 6 x 4 cm was attached to the frame. The required numbers of holes were made for adjustments.

The equipment was tested in the farmers' field at village Debil of Khurda district, Orissa for digging of AK-12-24 variety groundnut as per standard test procedures (I.S.:

11235-1985). Observations on power requirement, effective field capacity, field efficiency, labour requirement pod losses, digging efficiency, performance index and economics of digging were recorded. The moisture content of pod, vine and soil were determined by air over method by dry weight basis. The draft, power requirement and effective field capacity were measured during the test. The pods left on the surface, within the soil and damaged pods were collected separately from the specified area to determine the losses.

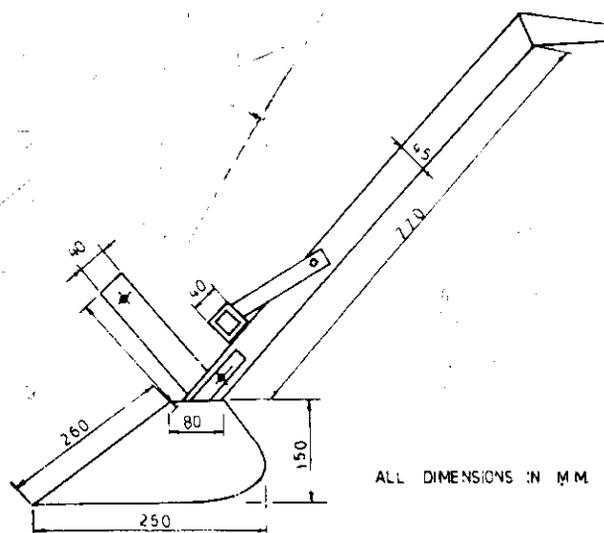


Fig. 1. Groundnut digger (Ridger type)

Percentage of exposed pod loss, $L_a = G/A \times 100$

Percentage of unexposed pod loss, $L_b = H/A \times 100$

Percentage of undug pod loss, $L_c = K/A \times 100$

Where,

G = quantity of detached pods lying exposed on the surface;
H = quantity of detached pods remained inside the soil in the sample area;
K = quantity of pods remained undetached from the undug plants in the sample area
A = total quantity of pods collected from the plants in the sample area.

Development of a bullock drawn groundnut digger suitable for coastal orissa

Total percentage of pod loss = percentage of (exposed + unexposed + undug) pod loss

Digging efficiency = 100 - total percentage of pod loss

Draft, $D = P \cos \theta$

Where,

D = draft in kgf; P = pull in kgf; θ = angle between line of pull and horizontal plane

Power requirement, H.P. = (Draft in kgf x speed in m/sec)/75

Effective field capacity: It is the actual rate of coverage. It includes the time lost making adjustment, unlogging the blades, etc. It is expressed in hectares per hour.

Field efficiency: It is the ratio of effective field capacity to theoretical field capacity and is expressed in percentage.

Labour requirement: It is the number of labours required for one ha area for digging groundnut.

$$\text{Performance index} = \frac{(1-L_a)(1-L_b)(1-L_c) \times W_d \times b_D \times 75}{R_L \times D_D}$$

Where, W_d = total weight of groundnut dug in kg

b_D = width of cut of groundnut digger in meter

R_L = length of run in meter

D_D = draft of groundnut digger in kgf

The draft required to run the groundnut digger varied from 50.2 to 63.9 kgf with an average value of 57.0 kgf and the power requirement from 0.31 to 0.48 hp with an average value of 0.39 hp (Table 1). The average theoretical and effective field capacities were found to be 0.065 ha/hr and 0.058 ha/hr, respectively. The draft and effective field capacity of a riding type bullock drawn groundnut digger developed at TNAU, Coimbatore were reported as 100 kg and 0.09 ha/hr, respectively (Devnani, 1978). The results obtained from the performance evaluation of the above digger corroborated with the results of developed ridger type groundnut digger. The draft and field capacity of the ridger type groundnut digger was also found to be of some range as obtained by Quadri (1988) from the field evaluation of a semicircular blade groundnut digger in Maharashtra.

The effect of speed on effective field capacity of the groundnut digger is shown in Fig. 2. The average field efficiency was found to be 89%. The optimum operating speed to achieve maximum digging efficiency was 1.9 km/hr with a digging efficiency of 91.8%. The average digging efficiency was found to be 90.1%. The variation of digging efficiency with respect to speed of operation of the digger is shown in Fig. 3. The performance index of the groundnut digger was found to be 0.101 considering the quality of digging, quantity of digging and power requirement for the operation. The cost of operation in case of manual digging method was found to be

Rs.1100/ha as compared to Rs. 653.40/ha in this ridger digger, but the effective field capacity of the said digger (0.058 ha/hr) is much higher than manual method (0.0075 ha/hr). The performance of the developed groundnut digger was satisfactory and found to be suitable for harvesting of groundnut in sandy loam soil of coastal Orissa.

Table 1 Average values of power requirement, losses, field efficiency, digging efficiency, performance index and cost of operation of two row riding type groundnut digger

Speed (km/hr)	1.86
Draft (kgf)	57.0
H.P.	0.39
Effective field capacity (ha/hr)	0.058
Theoretical field capacity (ha/hr)	0.065
Field efficiency (%)	89.23
Labour requirement (man hr/ha)	52.18
Exposed pod loss (%)	3.6
Unexposed pod loss (%)	5.5
Undug loss (%)	0.8
Total loss (%)	9.9
Digging efficiency (%)	90.1
Width of cut (cm)	35.0
Performance index	0.101
Cost of operation (Rs/ha)	653.40

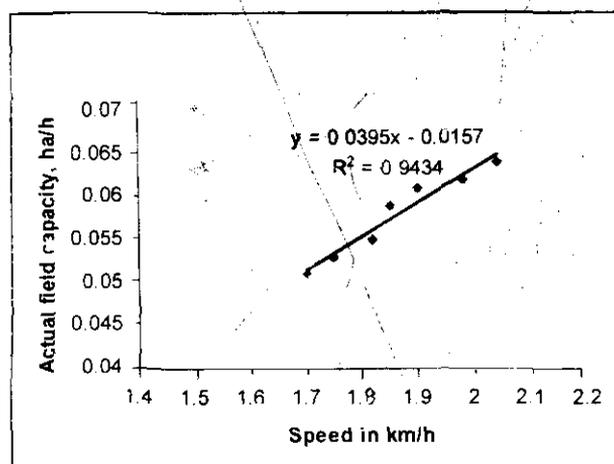


Fig. 2. Effect of speed on actual field capacity

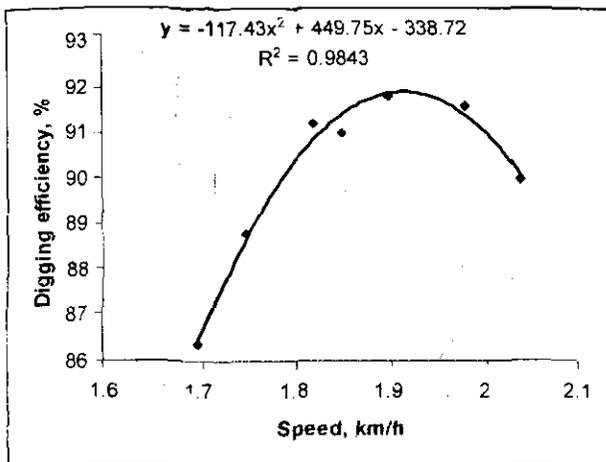


Fig. 3. Effect of speed on digging efficiency

References

- Anonymous.** 2008. Orissa Agricultural Statistics 2006-07. Directorate of Agriculture and Food Production, Government of Orissa, Bhubaneswar, Orissa.
- Devnani, R.S.** 1978. A review of harvesting equipment developed in India. CIAE, Bhopal, India.
- I.S. : 11235-1985.** Indian standard test code for groundnut digger-Animal drawn, BIS, New Delhi.
- Quadri, S.A.** 1988. Development and testing of a groundnut digger. Paper presented at 24th Annual Convention of ISAE held at PDKV, Akola, Maharashtra during 21-23 January.

**Kind Attention to
All the Life Members of
ISOR**

Dear Life Members

The List of Mailing Address of all the Life Members of ISOR is appended along with this issue of Journal of Oilseeds Research. All the Life Members are therefore requested to kindly inform in writing, the change or mistake in their mailing address to

The General Secretary
Indian Society of Oilseeds Research
Directorate of Oilseeds Research
Rajendranagar, Hyderabad-500 030
E-mail: oilseedsociety@gmail.com

so as to enable us to make necessary changes in their mailing address to deliver the things to them at proper address.

An unique ID Number has been allotted to each Life Member and is mentioned just above to their name in the address list. All are requested to kindly note their ID Number and mention the same in their future correspondence.

ID No.1001
Dr. Abha Agnihotri
Fellow, TERI
Darbari Seth Block
Habitat Place, Lodhi Road
New Delhi
New Delhi

ID No.1004
Dr. Adiver, S.S.
Jr. Plant Pathologist (Oilseeds)
Main Research Station
University of Agricultural Sciences
Krishinagar
Dharwad-580 005
Karnataka

ID No.1007
Dr. Agarwal, A.P.
Scientist (Plant Breeding & Genetics)
Agricultural Research Station
Kumhrawan, Bastar Dt.
~~Bastar-494 005~~
Chhattisgarh

ID No.1010
Dr. Agarwal, S.K.
Scientist (Oilseeds)
No. 8/363, Daga Buildings
Byron Bazar
Raipur-492 001

ID No.1013
Dr. Aglawe, B.N.
Jr. Agronomist
Oilseeds Research Station
Marathwada Agricultural University
Latur-413 512
Maharashtra

ID No.1016
Dr. Ajay Kumar Singh
Scientist
Directorate of Rapeseed-Mustard
Research
Sewar Post
Bharatpur-321 303
Rajasthan

ID No.1002
Dr. Abhijit M. Ranaware
Jr. Plant Pathologist
Nimbkar Agril. Research Institute
Survey No. 6471, Vivekanand Nagar
Maharashtra

ID No.1005
Dr. Adkine, S.J.
Jr. Research Assistant
Marathwada Agricultural University
Parbhani-431 402
Maharashtra

ID No.1008
Dr. Agarwal, K.N.
Shri Hari Industries
Hari Oil Mills, P.B. No. 7
Malgodown Road
~~Bharatpur-321 001~~
Rajasthan

ID No.1011
Dr. Agasimani, C.A.
"Somnathkrupa"
Shreenagar Imp. Board Layout
Near KUD Cross
Dharwad-580 003
Karnataka

ID No.1014
Dr. Ahuja, D.B.
Principal Scientist (Entomology)
NCIPM
L&M Wing, LBS Building
IARI Campus, Pusa
New Delhi-110 012
New Delhi

ID No.1017
Dr. Ajay Srivastava
Oilseeds Research Station
CSK H.P. Krishi Vishwa Vidyalaya
Kangra-176 001
Himachal Pradesh

ID No.1003
Dr. Abraham, Z.
Principal Scientist & Head
NBPGR Regional Research Station
Kerala Agricultural University
Thrissur-680 656
Kerala

ID No.1006
Dr. Agarkar, G.D.
Associate Professor
Near Vivekanand Ashram
Sudhir Colony
Akola-444 001
Maharashtra

ID No.1009
Dr. Agarwal, O.P.
Director
Torika Processors Pvt. Ltd.
6th Floor, Nariman Point
Mumbai-400 012
Maharashtra

ID No.1012
Dr. Aginal, M.B.
Training Associate
Krishi Vigyan Kendra
Raichur-584 101
Karnataka

ID No.1015
Dr. Ahuja, K.L.
Bio-Chemist (Oilseeds)
Department of Plant Breeding
Punjab Agricultural University
Ludhiana-141 001
Punjab

ID No.1018
Dr. Ajit Pratap Singh
Dept. of Genetics & Plant Breeding
C.S. Azad University of Agri. &
Technology
Kanpur-208 002
Uttar Pradesh

ID No.1019
Dr. Akashe, V.B.
Jr. Entomologist
AICRP on Safflower
Bramirath, Agricultural **School**
Compound
Solapur-413 002
Maharashtra

ID No.1020
Dr. Akhauri, R.K.
Oilseeds Coordinator (Sunflower)
Rajendra Agricultural University
Tirhut Agricultural College
Dholi-
Bihar

ID No.1021
Ms. Alivelu, K.
Scientist
Directorate of Oilseeds **Research**
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1022
Dr. Alok Jyotishi
Jr. Scientist (Agronomy)
S & N Project,
J.N. Krishi Vishwa Vidyalaya
P.O. Adhartal Krishi Nagar
Jabalpur-482 004
Madhya Pradesh

ID No.1023
Dr. Amaregouda, A.
Asst. Professor (Crop Physiology)
College of Agriculture
Regional Agril. Research Station
University of Agricultural Sciences
Raichur-584 101
Karnataka

ID No.1024
Dr. Amarjit Kaur Atwal
Biochemist
Dept. of Plant Breeding & Biotechnology
Punjab Agricultural University
Ludhiana-141 004
Punjab

ID No.1025
Mr. Ambalal L. Patel
Aravalli Far Services
4, Omiya House
Opp: Bus Station
Himatnagar-383 001
Gujarat

ID No.1026
Mr. Ambidekar Suresh Namdevrao
Dt. Superinting Agril. Officer
Agril. Department, Shivaji Nagar
F.No. 8-9
Nanded
Maharashtra

ID No.1027
Dr. Amitava Datta
Asst. Botanist (Sunflower)
Dept. of Agriculture
Pulses and Oilseeds Research Station
Berhampore-782 101
West Bengal

ID No.1028
Dr. Anand M. Badigannavar
Nuclear Agril. & Biotech Division
Bhabha Atomic Research Centre
Trombay
Mumbai-400 035
Maharashtra

ID No.1029
Dr. Ananthasayana, K.
Associate Professor
MIG-II, 9-Musk Mahal Colony
HUDA Complex
Hyderabad-500 264
Andhra Pradesh

ID No.1030
Dr. Angadi, S.P.
H.No. 582, 6th Cross
"A" Block, Sahakarnagar
Bengaluru-560 092
Karnataka

ID No.1031
Dr. Anil Kumar, P.
Asst. Professor
Agricultural College
ANG Ranga Agricultural University
Bapatla-522 101
Andhra Pradesh

ID No.1032
Dr. Anita B. Chopra
Asst. Professor
Department of Agronomy
Dr. Panjabrao Deshmukh **Vidyapeeth**
Akola-444 104
Maharashtra

ID No.1033
Dr. Anjaneya Reddy, B.
Scientist
AICRP on Sunflower
Regional Agril. Research Station
University of Agricultural Sciences
Raichur-584 101
Karnataka

ID No.1034
Dr. Anjani, K.
Principal Scientist (Plant **breeding**)
Directorate of Oilseeds **Research**
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1035
Dr. Anjay Tripathi
Technical Officer (PBG)
Project Coordinating Unit (S & N)
J.N. Krishi Vishwa Vidyalaya
Jabalpur-482 004
Madhya Pradesh

ID No.1036
Dr. Ankaiah, R.
H.No. 134/3RT, Sirikiran Apartments
Flat No. 201, Vijayanagar Colony
Hyderabad-500 457
Andhra Pradesh

ID No.1037
Dr. Ansari, M.M.
Sr. Scientist (Plant Pathology)
Directorate of Soybean Research
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.1038
Dr. Ansari, N.
Dept. of Genetics and Plant Breeding
College of Agriculture
ANG Ranga Agricultural University
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1039
Dr. Anup K. Mishra
Scientist
Directorate of Rapeseed Mustard
Research
Sewar
Bharatpur-321 303
Rajasthan

ID No. 1040
Ms. Anusha Srinivasan
Jr. Research Fellow
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1041
Dr. Aparna, K.
Flat No. F-2, Plot No. 14 & 19A
Road No. 2, Snehapuri Colony

ID No.1042
Dr. Appa Rao, G.
SRF, Hybrid Sunflower Project
Regional Agril. Research Station
Noonepalli
Nandyal-518 503
Andhra Pradesh

ID No.1043
Dr. Aqil Ahmad
Plant Physiology Section
Department of Botany
Aligarh Muslim University
Aligarh-202 002
Uttar Pradesh

ID No.1044
Dr. Arpita Ray
Department of Apiculture
University of Agricultural Sciences
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1045
Dr. Arun Kumar
Project Coordinating Unit (R&M)
Project Leader, Dept. of Pl. Breeding
Birsra Agricultural University

ID No.1046
Dr. Arunachalam, V.
Flat No. 1, Ram & Bridge View Apts-2
Central Avenue Road
Kodambakkam
Chennai-600 024
Tamil Nadu

ID No.1047
Dr. Arvind Kumar
Dy. Director General (Edn)
Indian Council of Agril. Research
Krishi Bhawan-1
New Delhi-110 012
New Delhi

ID No.1048
Dr. Arvind Kumar Gupta
Jr. Agronomist
AICRP on Linseed
Indira Gandhi Krishi Vishwa Vidyalaya
Raipur-492 006
Chhattisgarh

ID No.1049
Dr. Ashok Anand Rao Pisal
Asst. Professor
Dept. of Agronomy
College of Agriculture
Kolhapur
Maharashtra

ID No.1050
Dr. Ashok Kumar
Scientist (Plant Pathology)
Oilseeds Research Station
Himachal Pradesh Krishi Vidyapeeth
Kangra-176 001
Himachal Pradesh

ID No.1051
Dr. Ashok Kumar
Scientist (Plant Breeding)
ICRISAT, Patancheru
Medak District
Patancheru-502 324
Andhra Pradesh

ID No.1052
Dr. Ashok Kumar Sharma
Scientist
Directorate of Rapeseed Mustard
Research
Sewar Farm
Bharatpur-321 001
Rajasthan

ID No.1053
Dr. Ashok Kumar, A.
Agronomist
Regional Agril. Research Station
ANG Ranga Agricultural University
Nandyal-518 503
Andhra Pradesh

ID No.1054
Dr. Ashok Kumar, K.
Scientist (Agronomy)
Regional Agril. Research Station
S/o K. Venkata Swamy
78-95-3, Krishna Nagar

ID No.1055
Dr. Ashok S. Dhawan
Professor
Dept. of Ag. Chemistry & Social Science
Marathwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1056
Dr. Aurangabadkar, L.P.
Ankur Seeds Pvt. Ltd.
27, New Cotton Market Layout
Nagpur-440 017
Maharashtra

ID No.1057
Dr. Avinash Kumar Gupta
Scientist (Entomology)
AICRP on Linseed
College of Agriculture
Indira Gandhi Krishi Viswa Vidyalaya

ID No.1058
Dr. Awachit Rao T. Bhogle
Jr. Breeder
Asst. Professor of Botany
Dr. Punjabrao Deshmukh Krishi
Vidyapeet
Akola-444 104
Maharashtra

ID No.1059
Dr. Awasthi Alok Kumar
Jr. Scientist (Ento.)
Regional Agril. Research Station
Indira Gandhi Krishi Viswa Vidyalaya
Sarkanda
Bilaspur-495 001
Madhya Pradesh

ID No.1060
Dr. Aziz Qureshi, M.A.
Sr. Scientist
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1061
Dr. Babalad, H.B.
Jr. Agronomist
AICRP on Soybean, Dept. of Agronomy
University of Agricultural Sciences

ID No.1062
Dr. Babubhai M. Patel
Asst. Research Scientist
AICRP on UUC
Regional Research Station
S.D. Agricultural University
S.K. Nagar-385 506
Gujarat

ID No.1063
Ms. Baby Akula
Plot No. 235
Vaidehinagar
Hyderabad-500 070
Andhra Pradesh

ID No.1064
Dr. Badoria, H.S.
Jr. Plant Physiologist
Main Castor & Mustard Research
Station
S.D. Agricultural University
S.K. Nagar-385 506
Gujarat

ID No.1065
Dr. Bajaj, R.K.
Senior Breeder (Sunflower)
Punjab Agricultural University
Ludhiana-141 004
Punjab

ID No.1066
Dr. Bakheta, D.R.C.
MIG Flat No. 16-FF
Block-C, Rishi Nagar
Ludhiana-141 001
Punjab

ID No.1067
Mr. Balakishan, G.
Technical Officer
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1068
Dr. Balakrishna, P.
Associate Professor
Dept. of Seed Science & Technology
University of Agril. Sciences
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1069
Mr. Balakrishnan, R.
Scientist (SG) (Statistics)
Computer Centre
Sugarcane Breeding Institute
Coimbatore-641 003
Tamil Nadu

ID No.1070
Dr. Balode, K.L.
Jr. Pathologist (Sunflower)
Punjabrao Krishi Vidyapeeth
Akola-444 104
Maharashtra

ID No.1071
Mr. Balubhai Vallabhibhai Thumar
Agricultural Research Station
Kerin Road
Gujarat Agricultural University
Amreli-365 601
Gujarat

ID No.1072
Dr. Bandi, A.G.
Professor (Agronomy)
Dept. of Agronomy, College of
Agriculture
University of Agricultural Sciences
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1073
Dr. Bandopadhyay, A.
C-201, NASC Complex
D.P. Shastri Marg
New Delhi-110 012
New Delhi

ID No.1074
Dr. Banga, S.S.
Dept. of Plant Breeding
Punjab Agricultural University
Ludhiana-141 004
Punjab

ID No.1075
Dr. Banga, R.S.
Sr. Scientist (Agronomy)
Dept. of Plant Breeding
CCS Haryana Agril. University
Hisar-125 004
Haryana

ID No.1076
Dr. Bapodra, J.G.
Gurukrupa
Girnar Society
Near Motibagh
Junagadh-362 001
Gujarat

ID No.1077
Dr. Bargale, P.C.
Scientist (Sr. Scale)
Central Institute of Agril. Engineering
Nabi Bagh, Berasia Road
Bhopal-462 038
Madhya Pradesh

ID No.1078
Dr. Basappa, H.
Principal Scientist (Entomology)
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1079
Dr. Basavaraja, G.T.
Associate Professor
AICRP on Soybean
University of Agricultural Sciences
Dharwad-580 005
Karnataka

ID No.1080
Dr. Basavarajappa, M.P.
Asst. Professor
AICRP (Safflower), Agril. Research
Station
University of Agricultural Sciences

ID No.1081
Dr. Basavegoud
Asst. Professor
Regional Research Station
University of Agricultural Sciences
Raichur-584 101
Karnataka

ID No.1082
Dr. Basu, M.S.
ICRISAT
Patancheru
Medak District
Patancheru-

ID No.1083
Mr. Beena Balakrishna Pillai
Sr. Research Asst. (Botany)
College of Agriculture
Nagpur-
Maharashtra

ID No.1084
Dr. Benagi, V.I.
Professor & Head
Dept. of Plant Pathology
University of Agricultural Sciences
Dharwad-580 005
Karnataka

ID No.1085
Dr. Bener Raj
Plant Pathologist
Directorate of Rice Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1086
Dr. Bera, S.K.
Scientist
Directorate of Groundnut Research
P.B. No.5
Ivnagar Road
Junagadh-362 001
Gujarat

ID No.1087
Mr. Bhabani Shankar Bishayi
Programme Assistant (Agronomy)
Krishi Vigyan Kendra
Orissa University of Agril. & Tech.
Subarnayur-767 017
Orissa

ID No.1088
Dr. Bhagat, N.R.
266, Jay Durga, Society No. 11
Extended Narendra Nagar
Nagpur-440 015
Maharashtra

ID No.1089
Dr. Bhalani, P.A.
Research Scientist (Entomology)
Oilseeds Research Station
Gujarat Agriculture University
Junagadh-362 001
Gujarat

ID No.1090
Dr. Bhale Mohan Sadashiv
Dept. of Plant Pathology
College of Agriculture
J.N. Krishi Vishwa Vidyalaya
Jabalpur-482 004
Madhya Pradesh

ID No.1091
Dr. Bhale Vilas Madhukar
Associate Professor
Dept. of Agronomy
Marathwada Agricultural University
Parbhani-431 402
Maharashtra

ID No.1092
Dr. Bhalla, C.S.
Scientist, Dept. of Agronomy
J.N. Krishi Vishwa Vidyalaya
Jabalpur-482 004
Madhya Pradesh

ID No.1093
Dr. Bhanu Rekha, K.
Research Associate
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1094
Dr. Bharaj, G.S.
Sr. Scientist (Entomology)
G-4, Krishi Nagar
Indore-452 001
Madhya Pradesh

ID No.1095
Dr. Bharatha Lakshmi, M.
Sr. Scientist (Agronomy)
Regional Agril. Research Station
Anakapalle-
Andhra Pradesh

ID No.1096
Dr. Bharathi, M.
Asst. Professor
Dept. of Genetics & Pl. Breeding
College of Agriculture, ANGRAU
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1097
Dr. Bharati, S.
A/20, KHB Colony
Puttanahalli, Yelahanka
Bengaluru-560 064
Karnataka

ID No.1098
Dr. Bhaskar R. Wadekar
Mahodaya Hybrid Seeds Pvt. Ltd.
Keshar Complex, P.B. N. 60
Head Post Office Road
Jalna-431 203
Maharashtra

ID No.1099
Dr. Bhaskar, K.
Research Associate
49-1-2, J.D. Laxmi Nagar
Kurnool-518 002
Andhra Pradesh

ID No.1100
Mr. Bhaskara Reddy, M.
Technical Officer
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 040
Andhra Pradesh

ID No.1101
Dr. Bhateria, S.
Sr. Plant Breeder
Dept. of Plant Breeding & Genetics
H.P. Krishi Viswa Vidyalaya
Palampur-176 062
Himachal Pradesh

ID No.1102
Dr. Bhatt, N.S.
Entomologist
Dept. of Apiculture
University of Agril. Sciences, GKVK
Bengaluru-560 065
Karnataka

ID No.1103
Dr. Bhawnick, M.K.
Asst. Agronomist
Dept. of Agriculture
Pulses and Oilseeds Research Station
Barnampore-742 101
West Bengal

ID No.1104
Dr. Bhimsen Naik
Jr. Breeder (Linseed)
Regional Research and Tech. Training
Sub Station
Jashipur-757 091
Orissa

ID No.1105
Dr. Bhoori Singh Sinsinwar
427, Krishna Nagar
Bharatpur-321 001
Rajasthan

ID No.1106
Dr. Bindumadhava, A.N.
Proagro Seed Co. Pvt. Ltd.
8-1-39, Tolichowki
Hyderabad-500 008
Andhra Pradesh

ID No.1107
Dr. Birbal Sahu
Krishi Vigyan Kendra
Near Rajgarhnaka
Jhabua-457 661
Madhya Pradesh

ID No.1108
Dr. Bohra, J.S.
Professor & I/c CSR Project
Dept. of Agronomy
Institute of Agricultural Sciences
Banaras Hindu University

ID No.1109
Dr. Chakrabarthy, S.K.
Sr. Scientist
Seed Technology Division
Indian Agricultural Research Institute
Pusa
New Delhi-110 012
New Delhi

ID No.1110
Dr. Bhat, M.G.
Director
NRC for Cashew
Dakshina Kannada
Puttur
Karnataka

ID No.1111
Dr. Bhatia, V.S.
Sr. Scientist (Crop Physiology)
Directorate of Soybean Research
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.1112
Dr. Bhausaheb D. Jadhav
Associate Research Scientist
Niger Research Station
Gujarat Agricultural University
Vanarasi-396 580
Gujarat

ID No.1113
Dr. Bhillore, S.D.
Sr. Scientist (Agronomy)
Directorate of Soybean Research
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.1114
Dr. Bhogal, N.S.
Sr. Scientist
Directorate of Rapeseed Mustard
Research
Sewar
Bharatpur-321 303
Rajasthan

ID No.1115
Dr. Bikram Singh
Scientist
Regional Research Station
CCS Haryana Agril. University
Bawal-123 501
Haryana

ID No.1116
Dr. Biradar Patil, N.K.
Associate Professor
NSP Seed Unit
University of Agricultural Sciences
Dharwad-580 005
Karnataka

ID No.1117
Dr. Bisht, R.S.
C/o Dr. R.S. Banga
Oilseeds Section, Dept. of Pl. Breeding
Haryana Agricultural University
Hisar-125 004
Haryana

ID No.1118
Dr. Chahal, A.S.
37-A, Shant Park
Ferozepur
Ludhiana-141 012
Punjab

ID No.1119
Dr. Chakresh Kumar
Oilseeds Breeder
Agricultural Research Station
Navgaon, Alwar

ID No.1120
Dr. Chander Rao, S.
Senior Scientist (Pl. Pathology)
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1121
Dr. Chandra Mohan,
C/o S. Chandraiah
H.No. 21-107/8
Syndicate Bank Colony
Kothapet
Hyderabad-500 035
Andhra Pradesh

ID No.1122
Dr. Chandran, K.
Sr. Scientist (Biotechnology)
Sugarcane Breeding Institute
Research Centre, Civil Station (Post)
Coonur-670 002
Tamil Nadu

ID No.1123
Dr. Changa Reddy V.
Cotton Breeder
Regional Agril. Research Station
Lam
Guntur-522 001
Andhra Pradesh

ID No.1124
Dr. Channappa Goudar, B.B.
Physiologist
AICRP on Weed Control
Main Agril. Research Station
University of Agricultural Sciences

ID No.1125
Dr. Chattopadhyay, C.
Principal Scientist (Pl. Pathology)
Division of Crop Protection
Indian Institute of Pulses Research
Kanpur-208 002
Uttar Pradesh

ID No.1126
Dr. Chaudhary, B.D.
121, Mohalla Chaudharian
Hisar-125 004
Haryana

ID No.1127
Dr. Chaudhary, R.K.
Entomologist (Safflower)
G-3, Krishinagar
College of Agriculture
J.N. Krishi Vishwa Vidyalaya
Indore-452 001
Madhya Pradesh

ID No.1128
Dr. Chauhan, Y.S.
Training Organiser
F-11, Krishi Colony
Gwalior-474 002
Madhya Pradesh

ID No.1129
Mr. Chauhan Milichand Hirachand
Oilseeds Specialist
Oilseeds Research Station
Latur-413 512
Maharashtra

ID No.1130
Dr. Chandrakala, R.
Plant Breeder, AICRP (Sunflower)
Tamil Nadu Agricultural University
Coimbatore-641 003
Tamil Nadu

ID No.1131
Dr. Chandrakar, P.K.
Jr. Scientist
Dept. of Plant Breeding
Indira Gandhi Krishi Viswa Vidyalaya
Krishnagar
Raipur-492 012
Madhya Pradesh

ID No.1132
Dr. Chandranath, H.T.
Agril. Tech. Information Centre (ATIC)
University of Agril. Sciences
P.B. No. 24
Dharwad-580 005
Karnataka

ID No.1133
Dr. Channakrishnaiah, K.M.
Professor (Sunflower Breeding)
University of Agril. Sciences
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1134
Dr. Charitha Devi, M.
Plot No. 34, L. Nagar
TTD Employees Colony
Tirupati-517 502
Andhra Pradesh

ID No.1135
Dr. Chaudhari, F.P.
At & PO-Khandusan
Near Dairy, Tq: Visnagar
Mehsana-384 310
Gujarat

ID No.1136
Dr. Chaudhary, J.L.
Meteorologist
Zonal Agril. Research Station
Kumhrawand Farm
Jagdalpur-494 005
Chhattisgadh

ID No.1137
Dr. Chauhan, G.S.
Director
Directorate of Soybean Research
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.1138
Dr. Chauhan, Y.S.
Sr. Oilseeds Breeder
Narendra Dev University of Ag. & Tech.,
Dept. of Genetics & Plant Breeding
Faizabad-224 229
Uttar Pradesh

ID No.1139
Dr. Chauvalia, B.M.
Agril. Supervisor
Agril. Research Station
Gujarat Agricultural University
Amreli-365 601
Gujarat

ID No.1140
Dr. Chavre, K.D.
Technical Officer
AICRP (Safflower)
College of Agriculture, RVSKUU

ID No.1141
Dr. Chetti, M.B.
Professor
Dept. of Crop Physiology
University of Agril. Sciences
Krishinagar
Dharwad-580 005
Karnataka

ID No.1142
Dr. Chinnamuthu
Asst. Professor (Agronomy)
Dept. of Agronomy
Tamil Nadu Agricultural University
Coimbatore-641 003
Tamil Nadu

ID No.1143
Dr. Choubey, N.K.
Sr. Scientist (Agronomy)
Dept. of Agronomy
Indira Gandhi Krishi Vishwa Vidyalaya
Raipur-492 012
Chhattisgadh

ID No.1144
Dr. Chunilal
Scientist (Plant Breeding)
National Research Centre for Groundnut
P.B.No. 5, Ivnagar Road
Junagadh-362 001
Gujarat

ID No.1145
Dr. Dada Rao, S.R.S.
Associate Professor (Entomology)
Marathwada Agricultural University
Parbhani-431 402
Maharashtra

ID No.1146
Dr. Dakhre, S.R.
Sr. Manager (Seeds)
Agril. Production Division
Hindustan Lever Ltd.
No. 1-10-8/6, Begumpet
Hyderabad
Andhra Pradesh

ID No.1147
Dr. Dangaria, C.J.
Research Scientist (Millet)
Air Force Road, Millet Research Station
Junagadh Agricultural University

ID No.1148
Dr. Deokar, A.B.
14-A, Anurekha Society
Near Navsahyadri, Karvenagar
Pune-411 052
Maharashtra

ID No.1149
Dr. Desai, D.B.
Managing Director
Navbharat Seeds Pvt. Ltd.
4, Sarvodaya Community Centre
Ahmedabad-380 001
Gujarat

ID No.1150
Dr. Chellapilla Tara Satyavathi
Scientist (Pl. Breeding)
Directorate of Sorghum Research
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.1151
Dr. Chhaya Atri
Asst. Breeder (R&M)
Dept. of Plant Breeding & Genetics
Punjab Agricultural University
Ludhiana-141 004
Punjab

ID No.1152
Dr. Chiranjeevi, V.
General Manager
C&M Group, C&M House
N.D. Patel Road
Nasik-422 001
Maharashtra

ID No.1153
Dr. Choudhary R. Fujibhai
Training Associate (Pl. Protection)
C/o F.P. Chaudhary
At & PO: Khandosan, Near Dairy
T.A. Visnagar, Dt. Mehasana
Mehasana-384 310
Gujarat

ID No.1154
Dr. Dabre, W.M.
Sr. Research Scientist (Oilseeds)
Sandip Building
Hanuman Nagar
Akola-444 005
Maharashtra

ID No.1155
Dr. Dagaonkar, V.S.
93, Rajvilas Apartments
Narendranagar
Nagpur-440 015
Maharashtra

ID No.1156
Mr. Damodaram, T.
Head (DOIC)
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1157
Dr. Dattatri, K.
Scientist (Ag. Extension)
Zonal Project Directorate
CRIDA Campus, Santoshnagar
Saidabad
Hyderabad-500 059
Andhra Pradesh

ID No.1158
Dr. Desai, A.G.
Asst. Research Scientist
Main Castor-Mustard Research Station
S.D. Agricultural University
S.K. Nagar-385 506
Gujarat

ID No.1159
Dr. Desai, S.A.
Wheat Breeder
University of Agricultural Sciences
Krishi Nagar
Dharwad-580 005
Karnataka

ID No.1160
Dr. Deshmukh, M.P.
Asst. Professor of Plant Breeding
Agril. Research Station
Mahatma Phule Krishi Vidyalaya
K. Digraj, Sangli Dt.
K. Digraj-416 305
Maha

ID No.1161
Dr. Deshmukh, S.B.
Jr. Agronomist
AICRP on Safflower
Agricultural School Compound
Solapur-413 002
Maharashtra

ID No.1162
Dr. Deshmukh, S.R.
Sr. Research Assistant
Nimbkar Agricultural Research Institute
P.B. No. 44
Phaltan-415 502
Maharashtra

ID No.1163
Dr. Deshpande, M.B.
Jr. Agronomist
Nimbkar Agril. Research **Institute**
P.B. No. 44
Phaltan-415 502
Maharashtra

ID No.1164
Mr. Devasenamma, V.
MAO, MDO Office
Kadavalur, Kodvalur **Mandal**
Nellore District
Nellore
Andhra Pradesh

ID No.1165
Dr. Devi Dayal
Principal Scientist & Head
Regional Research Station
Central Arid Zone Research **Institute**
Kukma Bhuj-370 105
Gujarat

ID No.1166
Dr. Dhadge, S.M.
Niger Agronomist
AICRP on Niger, Zonal Agril. Res. Stn.,
Mahatma Phule Krishi Vidyapeeth

ID No.1167
Dr. Dhal, J.N.
Breeder (Groundnut)
Research Wing
Orissa University of Agril. & **Technology**
Bhubaneshwar-751 003
Orissa

ID No.1168
Dr. Dharmendra Sharma
Director of Research
Amber Hybrid Seeds Pvt. Ltd.
Jalna-
Maharashtra

ID No.1169
Dr. Dhillon, A.S.
Sr. Agronomist (Oilseeds)
Dept. of Plant Breeding
Punjab Agricultural University
Ludhiana-141 004
Punjab

ID No.1170
Dr. Deshmukh, M.R.
Tech. Asst. (Agronomy)
O/O Project Coordinating Unit (**S&N**)
J.N. Krishi Vishwa Vidyalyaya
Jabalpur-482 004
Madhya Pradesh

ID No.1171
Dr. Deshmukh, S.N.
Near Dr. Tarinos Hospital
Behind Rajasthanani Chatralaya
Kritinagar
Akola-
Maharashtra

ID No.1172
Dr. Deshpande, K.A.
Sr. Research Fellow
Oilseeds Research **Station**
Latur-413 512
Maharashtra

ID No.1173
Dr. Deshpande, S.L.
Scientist (Agronomy)
32, Telephone Nagar
Kokadia Road
Indore-452 018
Madhya Pradesh

ID No.1174
Dr. Devender Reddy
Principal Scientist (Agronomy)
Cropping System Research
ANG Ranga Agricultural University
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1175
Dr. Devraj Lenka
Jr. Breeder (Linseed)
Regional Agril. Research **Station**
Orissa University of Agril. & **Tech.**,
Judia Farm
Keonjhar-758 002
Orissa

ID No.1176
Dr. Dhaduk, L.K.
Breeder
Junagadh Agricultural **University**
Junagadh-362 001.
Gujarat

ID No.1177
Dr. Dharmendra Kumar, M.
Plant Breeder
Cosmo Plantgene Ltd.
3-52, Gulshan Farm, Medchal Mandal
Kandlakoia, Ranga Reddy Dist.
Kandlakoia-501 403
Andhra Pradesh

ID No.1178
Dr. Dhevagi
Asst. Professor
Dept. of Oilseeds
Tamil Nadu Agril. **University**
Coimbatore-641 003
Tamil Nadu

ID No.1179
Dr. Dhillon, R.S.
Scientist
Dept. of Forestry
CCS Haryana Agril. **University**
Hisar-125 004
Haryana

ID No.1180
Dr. Dhillon, S.S.
Associate Director of Research
Regional Research Station
Punjab Agricultural University
Bhatinda-151 001
Punjab

ID No.1181
Dr. Dhiraj Singh
Sr. Scientist (Plant Breeding)
Oilseeds Section
CCS Haryana Agricultural University
Hisar-125 004
Haryana

ID No.1182
Mr. Dileepkumar, G.
Research Associate
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1183
Dr. Dillon, S.K.
Sunflower Breeder
Department of Plant Breeding
Punjab Agricultural University
Ludhiana-141 004
Punjab

ID No.1184
Dr. Dinesh Chandra, A.T.
Asst. Research Scientist (PB)
3, Ankit Society, B/H Sarodaya Oil Mill
NH-14, Palampur Road
Palampur-380 001
Himachal Pradesh

ID No.1185
Dr. Dinesh Rai
Jr. Pathologist (R&M)
Tirhut College of Agriculture
Dholi-343 121
Bihar

ID No.1186
Dr. Dixit, A.K.
Area Agronomic Research Centre
Amiaha
Sehore-466 113
Madhya Pradesh

ID No.1187
Dr. Alok Kumar Patra
Sr. Scientist
AICRP on Agroforestry of Agriculture
Dept. of Forestry, College & Tech.
Orissa University of Agriculture

ID No.1188
Dr. Biradar, S.A.
Scientist (Agronomy)
AICRP on Linseed
Main Agricultural Research Station
University of Agricultural Sciences

ID No.1189
Dr. Govindappa, M.R.
Scientist (Plant Pathology)
AICRP on Linseed
Main Agricultural Research Station
University of Agricultural Sciences

ID No.1190
Dr. Dhiman Mukherjee
Asst. Professor (Agronomy)
Regional Research Station, Hill Zone
Uttar Banaga, Krishi Viswa Vidyalyaya
Kalinga Dt.,
Darjeeling
West Bengal

ID No.1191
Dr. Dileep Kachrao
Associate Professor
Division of Agronomy
SK University of Agril. & Technology
JRS Pura
Jammu

ID No.1192
Dr. Dilip J. Majumdar
Sagar Laxmi Seeds
206, Adhwamegh Avenue
North Mithakhat Underbridge
Mayur Colony, Navrangapura

ID No.1193
Dr. Dinand Kumar, V.
S/o Sri Jal Singh, SRF
Department of Soil Science
SVBPU&T
Modipuram-250 110
Uttar Pradesh

ID No.1194
Dr. Dinesh Kumar, V.
Senior Scientist (Biotechnology)
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1195
Dr. Divakara, B.N.
Scientist
ICRISAT
Patancheru-502 324
Andhra Pradesh

ID No.1196
Dr. Dixit, R.K.
Scientist (Oilseeds) & Head
Department of Genetics & Plant
Breeding
C.S. Azad University of Agril. & Tech.,
Kanpur-208 002
Uttar Pradesh

ID No.1197
Dr. Avijit Roy
Jr. Agronomist
AICRP on Sunflower
Ramakrishna Ashram, RAKVK
South 24 Parganas
Nimpith-743 338
West Bengal

ID No.1198
Dr. Dandanayake, B.P.
Asst. Pathologist
Regional Agril. Extension Edn. Centre
College of Agriculture
Latur-413 512
Maharashtra

- ID No.1199
Dr. Kulkarni, U.G.
Associate Dean & Principal
College of Agriculture
Marathwada Agril. University
Latur-413 512
Maharashtra
- ID No.1202
Dr. Durga Rani, Ch.V.
Associate Professor
Dept. of Agril. Biotechnology
College of Agriculture, ANGRAU
Rajendranagar
Hyderabad-500 030
Andhra Pradesh
- ID No.1205
Dr. Durga Prasad, M.M.K.
Professor (Retd.)
No.8, Mamatha Apartments
Street No. 4/2, R.K. Nagar
Hyderabad-500 047
Andhra Pradesh
- ID No.1208
Dr. Fateh Singh
Scientist
Directorate of Rapeseed-Mustard
Research
Sewar
Bharatpur-321 303
Rajasthan
- ID No.1211
Dr. Thakur, A.K.
Jr. Agronomist
AICRP (Castor)
S.G. College of Agriculture Res. Station
Kumharwand
- ID No.1214
Mr. Durga Bhavani, K.B.
Ph.D. Student
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh
- ID No.1200
Dr. Patel, I.S.
Research Scientist (Entomology)
AICRP on Pigeonpea
Main Pulses Research Station
S.D. Agricultural University
- ID No.1203
Dr. Dubey, R.N.
45, MIG, Barra-1
Kanpur-208 027
Uttar Pradesh
- ID No.1206
Dr. Dwivedi, R.S.
Principal Scientist (Plant Physiology)
Indian Institute of Sugarcane Research
Lucknow-
Uttar Pradesh
- ID No.1209
Dr. Ganapathy, M.
Reader, Dept. of Agronomy
99, Kanakashbai Nagar
Chidambaram-608 001
Tamil Nadu
- ID No.1212
Dr. Dubey, M.P.
Sr. Scientist (Agronomy)
AICRP on Linseed
J.N. Krishi Viswa Vidyalaya
P.O. Rajana
Sagar-470 002
Madhya Pradesh
- ID No.1215
Dr. Dwivedi, D.C.
Vishal Seeds Pvt. Ltd.
8-2-108/3, Hasthinapuram North
Sagar Road
Hyderabad-500 070
Andhra Pradesh
- ID No.1201
Dr. Lakshman Shyam Sundar
Jr. Breeder
AICRP on Sunflower
Ramakrishna Ashram, RAKVK
South 24 Parganas
Nimpith-743 338
West Bengal
- ID No.1204
Dr. Duhoon, S.S.
Project Coordinator (S&N)
J.N. Krishi Vishwa Vidyalaya
Jabalpur-482 004
Madhya Pradesh
- ID No.1207
Dr. Shinde, E.
Agril. Assistant
Crop Research Unit (Oilseeds)
Dr. Panjabrao Ra Krishividyaapeeth
Akola-444 104
Maharashtra
- ID No.1210
Dr. Shanwad, U.K.
Scientist (Agronomy)
AICRP on Sunflower
Main Agricultural Research Station
University of Agricultural Sciences
- ID No.1213
Dr. Dubey, S.D.
Jr. Biochemist
PC Unit (Linseed)
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh
- ID No.1216
Dr. Dwivedi, S.L.
102, Imperial Manor Apartments
Begumpet
Hyderabad-500 016
Andhra Pradesh

ID No.1217

Mr. Farzana Jabeen
Asst. Professor
Dept. of Genetics and Plant Breeding
College of Agriculture, ANGRAU
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1218

Dr. Gadewadikar, P.N.
Scientist (Pl. Breeding)
Zonal Agril. Research Station
J.N. Krishi Vishwa Vidyalaya
Morena-476 001
Madhya Pradesh

ID No.1219

Dr. Ganesan, K.N.
Asst. Professor (PBG)
Seed Centre
Seed Science & Tech. Building
Tamil Nadu Agricultural University

ID No.1220

Dr. Ganga Prasad, S.
Professor
Dept. of Genetics & Plant Breeding
College of Agriculture, Navile
Shimoga-577 201
Karnataka

ID No.1221

Dr. Ganga Rao, N.V.P.
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1222

Dr. Gangappa, E.
Jr. Breeder
AICRP (Sesame)
University of Agril. Sciences
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1223

Mr. Garu Preeth Car
Oilseeds Section
Punjab Agricultural University
Ludhiana-141 004
Punjab

ID No.1224

Dr. Geetha, K.N.
Jr. Agronomist
AICRP (Sunflower)
University of Agricultural Sciences
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1225

Dr. George Thomas
Dy. Manager (Research)
301 E, Archadya Residency
NCL North Avenue, Kompally Post
Secunderabad
Andhra Pradesh

ID No.1226

Dr. Ghewande, M.P.
Principal Scientist (Retd.)
At: Wakad (Jahagir)
Po: Malkapur Pangra

ID No.1227

Dr. Ghorpade, P.B.
Sr. Linseed Breeder
College of Agriculture
Nagpur-440 001
Maharashtra

ID No.1228

Dr. Ghosh, S.K.
Scientist, NARDI
C-15, Vikrampur
Secunderabad-500 009
Andhra Pradesh

ID No.1229

Dr. Ghuge, S.B.
Safflower Breeder
Dept. of Agronomy
Marathwada Agricultural University
Parbhani-431 407
Maharashtra

ID No.1230

Dr. Ganga Ram, A.
Sr. Scientist (Plant Breeding)
Regional Agril. Research Station
Palem, Mahaboobnagar Dt.
Palem-509 215
Andhra Pradesh

ID No.1231

Dr. Gangadhara Rao, S.V.S.
Sr. Scientist
Agril. Research Station
Yellamanchili-531 055
Andhra Pradesh

ID No.1232

Mr. Garad, L.P.
Asst. Professor
Satephal, Tq. Kalam
Osamanbad-413 405
Maharashtra

ID No.1233

Dr. Gawand, P.B.
Plant Breeder, AICRP
Dryland Agriculture
Near DAU College
Solapur-413 002
Maharashtra

ID No.1234

Dr. Geetha, B.
Research Associate
Tapioca & Castor Research Station
Tamil Nadu Agricultural University
Yethapur, Salem District
Yethapur-636 119
Tamil Nadu

ID No.1235
Dr. George V. Thomas
Director
Central Plantation Crops **Research Inst.**
Kasargod-671 124
Kerala

ID No.1236
Dr. Ghodke, M.K.
Sunflower Breeder
Oilseeds Research **Station**
Latur-413 512
Maharashtra

ID No.1237
Dr. Ghosh, R.K.
Reader
Dept. of Agronomy
Faculty of Agriculture
B.C. Krishi Vishwa Vidyalaya
Mohanpur
West Bengal

ID No.1238
Dr. Ghosh, S.K.
Plant Breeder (Oilseeds)
Dept. of Agriculture
Pulses and Oilseeds **Reserch Station**
Berhampore-742 101
West Bengal

ID No.1239
Dr. Giri Babu, P.
Scientist
Directorate of Oilseeds **Research**
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1240
Dr. Girijesh, G.K.
Asst. Professor of Agronomy
Agricultural Research Station
Bidare Post
Shimoga-577 222
Karnataka

ID No.1241
Dr. Girish M. Kalagudi
Biotechnologist
Block-21, Flat No.32
Ram Raj Nagar, Opp: Suchitra
Medchal Road

ID No.1242
Dr. Girish Puri
Sr. Scientist (Soil Science)
Dept. of Soil Science & Agril. **Chemistry**
J.N. Krishi Vishwa Vidyalaya
Jabalpur-482 004
Madhya Pradesh

ID No.1243
Dr. Godhaya, A.G.
Asst. Research Scientist
Oilseeds Section
Junagadh Agricultural University
Junagadh-362 001
Gujarat

ID No.1244
Dr. Gokhale, D.N.
Department of Agronomy
College of Agriculture
Marathwada Agricultural **University**
Latur-413 512
Maharashtra

ID No.1245
Dr. Gopal Reddy, B.
Sr. Scientist
National Seed Project
ANG Ranga Agricultural **University**
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1246
Dr. Gopal Reddy, P.
Sr. Plant Breeder
No. 14, 5th Main, Tavarekera
Main Road, Chikkadugodi Extension
DPC Post
Bengaluru-560 030
Karnataka

ID No.1247
Dr. Gopalan, A.
Professor & Head
Department of Forages
Tamil Nadu Agricultural University
Coimbatore-641 003
Tamil Nadu

ID No.1248
Dr. Gosh, P.K.
Principal Scientist & Head
Water Management Division
ICAR Research Complex for NEH
Region
Umiam, Umroi Road

ID No.1249
Dr. Giri, S.M.
Technical Assistant
Oilseeds Research **Station**
Marathwada Agricultural **University**
Latur-413 512
Maharashtra

ID No.1250
Dr. Giriraj, K.
Bairambad
Post: Vontiangadi
S.K. Kodagu District
S.K. Kodagu-571 211
Karnataka

ID No.1251
Dr. Girish M.M.
Jr. Biochemist
Marathwada Agril. University
Dept. of Botany, College of **Agriculture**
Parbhani-431 402
Maharashtra

ID No.1252
Dr. Girish, G.
Sr. Research Fellow
AICRP (Sunflower)
University of Agricultural **Sciences**
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1253

Dr. Gohil, V.L.
Sr. Breeder (Sesame)
Agril. Research Station
Gujarat Agricultural University
Amreli-365 601
Gujarat

ID No.1254

Dr. Golasangi, B.S.
Jr. Breeder (Linseed)
Regional Research Station
University of Agricultural Sciences
P.B. No. 24
Raichur-584 101
Karnataka

ID No.1255

Mr. Gopala Krishna Naidu, K.
Research Associate
AICRP on Soybean
University of Agril. Sciences
Dharwad-580 005
Karnataka

ID No.1256

Dr. Gopalakrishnan, C.
Associate Professor (Pathology)
Department of Plant Pathology
Tamil Nadu Agricultural University
Coimbatore-641 003
Tamil Nadu

ID No.1257

Dr. Gosh, P.K.
Head, Division of Crop Production
Indian Institute of Pulses Research

ID No.1258

Dr. Gouda, M.V.C.
Project Coordinator (Minor Millets)
University of Agricultural Sciences
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1259

Dr. Gouri Shanker
Jr. Breeder (Castor)
Regional Agril. Research Station
Palem, Mahaboobnagar District
Palem-509 215
Andhra Pradesh

ID No.1260

Dr. Govinda Rao, B.
Principal Scientist (Plant Breeding)
Redgram, Pulses Section
Regional Agril. Research Station
Lam
Guntur-522 001
Andhra Pradesh

ID No.1261

Mr. Gulganji, G.G.
Asst. Professor
Agricultural Research Station
University of Agricultural Sciences
Annigeri
Karnataka

ID No.1262

Dr. Gupta, M.L.
Sr. Breeder (Oilseeds)
Dept. of Plant Breeding
Punjab Agricultural University
Ludhiana-141 004
Punjab

ID No.1263

Dr. Gupta, P.K.
Assistant Professor
C.S. Azad University of Agril. & Tech.,
Kanpur-208 002
Uttar Pradesh

ID No.1264

Dr. Gupta, S.C.
Chief Manager (R&D)
Harrisons Malayalam Ltd.
6-1-79, First Floor
CEAT Building, Lakadi-ka-pool
Hyderabad
Andhra Pradesh

ID No.1265

Dr. Gururaj Sunkad
Jr. Pathologist (Groundnut)
Regional Research Station
University of Agricultural Sciences
Raichur-584 101
Karnataka

ID No.1266

Dr. Hajare, D.B.
Agronomist
99, Tejasari Vaishnavi Nagar
Vijapur Road
Solapur-413 003
Maharashtra

ID No.1267

Dr. Hanumantha Rao, C.
H.No. 11-6-590
Devi Bagh, Nampally
Hyderabad-500 001
Andhra Pradesh

ID No.1268

Dr. Hanumantharaya, L.
Jr. Entomologist
Agricultural Research Station
University of Agricultural Sciences
Annigeri
Karnataka

ID No.1269

Dr. Govardhan, M.
Jr. Agronomist
Regional Agricultural Research Station
Palem, Mahaboobnagar District
Palem-509 215
Andhra Pradesh

ID No.1270

Dr. Govindan, A.
Asst. Professor (Plant Breeding)
Tapioca Castor Research Station
Tamil Nadu Agricultural University
Yethapur-636 119
Tamil Nadu

ID No.1271
Dr. Gupta, G.K.
Sr. Scientist (Plant Pathology)
Directorate of Soybean Research
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.1272
Dr. Gupta, N.R.
Jr. Breeder (Castor)
Oilseeds Section
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1273
Dr. Gupta, R.R.
EB(O)/Scientist
Dept. of Genetics & Pl. Breeding
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1274
Dr. Gupta, S.K.
368, Shastri Nagar
Jammu-180 004
Jammu & Kashmir

ID No.1275
Dr. Guruswamy, T.
D.I. (Agril. Engineering)
Dept. of Farm Power & Agro Energy
College of Agril. & Engineering
Raichur-584 101
Karnataka

ID No.1276
Mr. Hanumant Babanrao Kenjale
AICRP on Niger
Zonal Agril. Research Station
Igatpuri-422 403
Maharashtra

ID No.1277
Dr. Hanumanthappa, C.
Jr. Agronomist
Regional Agril. Research Station
Hiriyur-512 143
Karnataka

ID No.1278
Ms. Hari Priya, Ch.V.
Technical Officer
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1279
Dr. Hariender Singh
Jr. Agronomist (Sunflower)
Tirhat College of Agriculture
Dholi-843 121
Bihar

ID No.1280
Dr. Harish Babu, B.N.
Breeder (Safflower), AICRP (Safflower)
Agricultural Research Station
University of Agricultural Sciences

ID No.1281
Dr. Harshal E. Patil
Asst. Professor (Niger Breeder)
Mayor Colony, Plot No. 1/A
Deopur, Dhule
Deopur-424 002
Maharashtra

ID No.1282
Dr. Harvir Singh
Principal Scientist & Head
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1283
Dr. Hemalata Sharma
Asst. Professor
41, Near Park, Sector-3
Hiranmagari
Udaipur-313 002
Rajasthan

ID No.1284
Dr. Hemalatha, V.
Scientist
Regional Agril. Research Station
Palem, Mahaboobnagar District
Palem-509 215
Andhra Pradesh

ID No.1285
Mr. Hulihalli, U.K.
Asst. Professor
JSYS, Herbha, Versha Building
Opp: I.B. Road
Haveri
Karnataka

ID No.1286
Dr. Indi, D.V.
Jr. Plant Pathologist
AICRP on Safflower
Agricultural School Compound
Solapur-413 002
Maharashtra

ID No.1287
Dr. Ismail U. Dhruj
Associate Research Scientist
Dept. of Plant Pathology
Main Oilseeds Research Station
Gujarat Agricultural University
Junagadh-362 001
Gujarat

ID No.1288
Dr. Jadhav, K.V.
Associate Research Scientist
C-39, Gunatit Nagar
"Satyam", Opp: Agricultural Campus
Near Motibag
Junagadh
Gujarat

ID No.1289
Dr. Harinath Naidu, P.
H.No. 18-1-504/10
Opp: State Bank of India
Bhavani Nagar
Tirupati-517 501
Andhra Pradesh

ID No.1290
Dr. Harish Chandra Singh
Oilseeds Section
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1291
Dr. Harve, C.D.
Dept. of Plant Breeding & Genetics
Agricultural Research Station
Agricultural College
Gwalior-474 002
Madhya Pradesh

ID No.1292
Dr. Hegde, D.M.
Project Director
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1293
Dr. Hemalatha, S.
Scientist (Agronomy)
A-1, No. 12-2-709/2
Flat No. 506, Laxman Enclave
Karolbagh, Mehdiapatnam
Hyderabad
Andhra Pradesh

ID No.1294
Dr. Hugar, P.S.
Sr. Information Specialist
Directorate of Extension
University of Agricultural Sciences
Krishi Nagar
Dharwad-580 005
Karnataka

ID No.1295
Dr. Hussain, K.
Jr. Agronomist
P.C. Unit (Linseed)
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1296
Dr. Ishwar Singh
Asst. Professor (Agronomy)
AICRP (Castor)
Agricultural Research Station
Mandor
Jodhpur-342 304
Rajasthan

ID No.1297
Dr. Jadhav, G.S.
Head, Dept. of Agronomy
Marathwada Agricultural University
Parbhani-431 402
Madhya Pradesh

ID No.1298
Dr. Jadhav, R.N.
Agril. Assistant
Oilseeds Research Station
Marathwada Agricultural University
Latur-413 512
Maharashtra

ID No.1299
Dr. Jagadeesh, A.H.
Scientist (Agronomy)
AICRP on Soybean
University of Agricultural Sciences
Krishi Nagar
Dharwad-580 065
Karnataka

ID No.1300
Dr. Jagadish Singh
Sr. Scientist (Plant Breeding)
G-I, Krishinagar
College of Agriculture
Indore-452 001
Madhya Pradesh

ID No.1301
Dr. Jaganathan, A.
Principal Scientist (Agronomy)
Agricultural Research Station
Kadiri-515 591
Andhra Pradesh

ID No.1302
Dr. Jagatap, P.K.
Sr. Research Fellow
Oilseeds Research Station
Marathwada Agricultural University
Latur-413 512
Maharashtra

ID No.1303
Dr. Jagmohan Singh
Castor Breeder
AICRP on Castor
Bhavani Patna Centre, RRTTS

ID No.1304
Dr. Jakhmola, S.S.
Professor & Head
Dept. of Entomology
J.N. Krishi Vishwa Vidyalay
College of Agriculture

ID No.1305
Dr. Janila, P.
Scientist (Plant Breeding)
Regional Agril. Research Station
ANGRAU, Palem
Mahaboobnagar District
Palem-509 215
Andhra Pradesh

ID No.1306
Mr. Javed Iqbal A. Mulla
Associate Professor
H.No. 8/1, L.S. Tank Road
Malmaddi
Dharwad-580 007
Karnataka

ID No.1307
Mr. Jay Lal Mahto
C/o Sri S.N. Mahto
Q.No. 3, Type-III (Old), ISM
Dhanbad-826 004
Bihar

ID No.1308
Dr. Jaya Mohan Rao
"Sree Kala", Flat No. 37
Road No. 5, Jubilee Hills
Hyderabad-500 004
Andhra Pradesh

ID No.1309
Dr. Jagadeesh, B.N.
Associate Professor of Genetics
AICRP on Sunflower
University of Agril. Sciences
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1310
Dr. Jagadish, K.
Jr. Entomologist (Sunflower)
No. 30/36, First Cross
Sundarnagar, Gokula
Bengaluru-560 065
Karnataka

ID No.1311
Dr. Jagannatha, D.P.
Sr. Sunflower Breeder
Super Elite Sunflower Seed Prodn.
Scheme, AICRP on Sunflower
University of Agril. Sciences, GKVK
Bengaluru-560 065
Karnataka

ID No.1312
Dr. Jagdish K. Dhanani
Producer, Annadata
ETV (Gujarati)
Ramoji Film City
Hyderabad-501 512
Andhra Pradesh

ID No.1313
Dr. Jain, V.K.
Associate Professor of Agronomy
Inside of Atul Nivas
Kadam-Shahab-Ka-Bada-Mamo-Ka
Bazar
Lashkar Post

ID No.1314
Dr. Jambhulkar, S.J.
NABTD, Bhabha Atomic Research
Centre
Trombay
Mumbai-400 085
Maharashtra

ID No.1315
Dr. Jansi Rani, R.
Asst. Professor of Agril. Extension
Tapica and Castor Research Station
P.G. Palayam (Post)
Yethapur-636 119
Tamil Nadu

ID No.1316
Mr. Jawale Laxman Narayan Rao
JRA, Seed Technology Unit
Marathwada Agril. University
Parbhani-413 402
Maharashtra

ID No.1317
Dr. Jaya Laxmi, V.
Scientist (Plant Breeding)
Regional Agril. Research Station
Nandyal, Kurnool District
Nandyal
Andhra Pradesh

ID No.1318
Dr. Jaya Ram Reddy, G.
Scientist (SG)
Central Research Institute of Dryland
Agril.
Santoshnagar, Saidabad
Hyderabad
Andhra Pradesh

ID No.1319
Dr. Jayappa, A.H.
Asst. Professor (SG)
Department of Entomology
University of Agril. Sciences
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1320
Dr. Jha, S.K.
Scientist (Ag. Extension)
Directorate of Rapeseed Mustard
Research
Sewar Farm
Bharatpur-321 001
Rajasthan

ID No.1321
Dr. Jitendra Bhushan Misra
Principal Scientist (Biochemistry)
P.B. No. 5, NRC for Groundnut
Ivnagar
Junagadh-362 001
Gujarat

ID No.1322
Dr. Jitendra Singh Chauhan
Principal Scientist (Pl. Breeding)
Directorate of Rapeseed Mustard
Research
Sewar Farm
Bharatpur-321 303
Rajasthan

ID No.1323
Dr. John Joel, A.
Associate Professor
U.G. Botany Lab, CPB&G
Tamil Nadu Agril. University
Coimbatore-641 003
Tamil Nadu

ID No.1324
Mr. Joseph Royal
H.No. 26-612, Royal Compound
R.S. Road, Nandyal
Kurnool District
Nandyal
Andhra Pradesh

ID No.1325
Dr. Joshi, H.J.
Research Scientist
Agril. Research Station
Gujarat Agril. University
Amreli-365 601
Gujarat

ID No.1326
Dr. Juyal, S.P.
Director (Retd.)
Directorate of Oilseeds Development
Telhan Bhawan, Himayatnagar
Hyderabad-500 029
Andhra Pradesh

ID No.1327
Dr. Kachhadia Vinodrai Harilal
Agricultural Officer
39-Shridhar Nagar, B/L Lal Building
Near Geta Nagar
Junagadh-362 001
Gujarat

ID No.1328
Dr. Kalarani, M.K.
Associated Professor (C.P.)
Tapioca and Castor Research Station
Yethapur, P.G. Palayam, Salem
Yethapur-636 119
Tamil Nadu

ID No.1329
Dr. Jegathambal, R.
Asst. Professor
Seed Science & Technology
Tapioca & Castor Research Station
Yethapur-636 119
Tamil Nadu

ID No.1330
Dr. Jha, S.K.
Scientist (Agronomy)
AICRP on Linseed, Dept. of Agronomy
College of Agriculture

ID No.1331
Dr. Jitendra H. Akade
Research Associate
Nimbkar Agril. Research Institute
Phaltan, Satara District
Phaltan-415 523
Maharashtra

ID No.1332
Mr. Jivani, L.L.
Agricultural Officer
77-A, Akashdeep, Giriraj Park Society
Near Dipanjali-2, Anandanagar
Timbawadi
Junagadh-362 015
Gujarat

ID No.1333
Dr. John Sudheer, M.
H.No. 13/682, Ramachandra Nagar
Anantapur-515 001
Andhra Pradesh

ID No.1334
Dr. Joshi, B.M.
C/o Dr. V.N. Deshmukh
Eye Specialist, Behind Tarodeka
Veg. Market, Vazirabad
Nanded-431 601
Maharashtra

ID No.1335
Dr. Joshi, O.P.
Principal Scientist (Soil Science)
Directorate of Soybean Research
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.1336
Dr. Jyoti Singh
Plant Pathologist
P.C. Unit (Linseed)
C.S. Azad University of Agril. & Technol.
Kanpur-208 002
Uttar Pradesh

ID No.1337
Mr. Kailash Digambarrao Navgre
Jr. Pathologist
AICRP on Safflower
Marathwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1338
Dr. Kale, U.V.
Moulikrupa
Behind Mantri Mangal Kanyalaya
Shivaram Nagar, Basmathi Road
Parbhani-431 401
Maharashtra

ID No.1339
Dr. Kalegare, N.K.
Associate Professor
Dept. of Agronomy
College of Agriculture
Nanded Road
Latur-413 512
Maharashtra

ID No.1340
Dr. Kaligoud, V.B.
field Officer
Karnataka Oilseed Growers Federation
Raibagh
Belgaum-591 255
Karnataka

ID No.1341
Dr. Kalyana Sundaram, A.
Asst. Professor (Ag. Entomology)
Regional Research Station
Tamil Nadu Agricultural University
Vridhachalam-606 001
Tamil Nadu

ID No.1342
Dr. Kamalkrishna Pal
Scientist
Directorate of Groundnut Research
Ivnagar Road, P.B. No.5
Junagadh-362 001
Gujarat

ID No.1343
Dr. Kamata Prasad
Principal Scientist (Agronomy)
Project Directorate for Cropping System
Research, Modipuram
Meerut-250 110
Uttar Pradesh

ID No.1346
Dr. Kandalkar, A.B.
Jr. Agronomist
Oilseeds Research Unit
Dr. Panjabrao Krishi Viswa Vidyalaya
Akola-444 104
Maharashtra

ID No.1349
Dr. Kalidas, P.
Principal Scientist
Directorate of Oilpalm Research
Pedavegi, West Godavari District
Pedavegi
Andhra Pradesh

ID No.1352
Dr. Kamannavar, P.Y.
Sr. Scientist (Plant Breeding)
Plot No. 12A/2, 1st Main, II Cross
Rajtagiri
Dharwad-580 001
Karnataka

ID No.1355
Dr. Kanbi, V.H.
Biochemist
Regional Research Station
SK Nagar-Dantiwada Agril. University
Dantiwada-385 506
Gujarat

ID No.1358
Dr. Balade, K.L.
Asst. Professor
K.L. Bawade, Hingwa Road
Kaulkhed
Akola
Maharashtra

ID No.1344
Dr. Kamlesh N. Choudhary
Asst. Professor
Dept. of Agricultural Botany
Navsari Agril. University
N.M. College of Agriculture

ID No.1347
Mr. Kanna Babu, N.
Scientist (Seed Technology)
Directorate of Sorghum Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1350
Dr. Kalpana Sastry, F
Principal Scientist
NAARM
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1353
Dr. Kamble, K.R.
Seed Technology Research Centre
Marathwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1356
Dr. Kandhola, S.S.
Asst. Breeder (Oilseeds)
Oilseeds Section, Dept. of Pl. Breeding
Punjab Agricultural University
Ludhiana-141 004
Punjab

ID No.1359
Dr. Kar, U.C.
Breeder, Q.No. Type-IV-27
OUAT Colony
Bhubaneswar-751 003
Orissa

ID No.1345
Dr. Kanade, B.C.
Jr. Plant Physiologist
AICRP (Safflower)
Agricultural School Compound
Solapur-413 002
Maharashtra

ID No.1348
Dr. Kapadia, S.I.
Research Scientist (C&M)
Main Castor & Mustard Research
Station
Gujarat Agricultural University
S.K. Nagar-385 506
Gujarat

ID No.1351
Dr. Kamal Dhawan
Sr. Biochemist
CCS Haryana Agricultural University
Hisar-125 004
Haryana

ID No.1354
Dr. Kamlesh Singh
PC Unit (Linseed)
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1357
Ms. Kanti Meena
Scientist
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1360
Dr. Karkal, V.Y.
Sr. Research Assistant
AICRP (Safflower)
Solapur-413 002
Maharashtra

ID No.1361
Dr. Karuna Sagar, G.
Asst. Professor
Dept. of Agronomy
S.V. Agricultural College
Tirupati-517 502
Andhra Pradesh

ID No.1362
Dr. Kathmale, D.K.
Agronomist, Shivdarshan Apartments
North Shivaji Nagar
Near Dadge Girls High School
Sangli-416 416
Maharashtra

ID No.1363
Dr. Kawthe, D.R.
Jr. Research Asst.
AICRP (Safflower)
Marathwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1364
Dr. Kerkhi, S.A.
Professor, Dept. of Pl. Breeding &
Genet.
S.V. B.P. University of Agril. & Tech.
Modipuram, Meerut
Meerut-250 110
Uttar Pradesh

ID No.1365
Dr. Khadtare, S.V.
Jr. Agronomist
AICRP on Safflower
Agril. School Compound
Solapur-413 003
Maharashtra

ID No.1366
Dr. Khanapara, D.V.
C-7, Academic Quarter
Junagadh Agril. University
Junagadh-362 001
Gujarat

ID No.1367
Dr. Kharat, B.S.
Oilseeds Research Unit
Dr. Panjabrao Krishi Viswa Vidyalyaya
Akola-444 104
Maharashtra

ID No.1368
Dr. Khayum Ahmed
Jr. Pathologist
Regional Agril. Research Station
Palem, Mahaboobnagar District
Palem-509 215
Andhra Pradesh

ID No.1369
Dr. Karamakar, P.G.
Principal Scientist (Pl. Breeding)
Central Research Institute for Jute and
Allied Fibres, Nilganj
Barrackpore-743 101
West Bengal

ID No.1370
Dr. Karle, A.S.
Agronomist (Safflower)
Marathwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1371
Dr. Karuna, K.
Jr. Pathologist
AICRP on Sunflower
Zonal Agril. Research Station
University of Agricultural Sciences
Bengaluru-560 065
Karnataka

ID No.1372
Dr. Katiyar, S.L.
Breeder (Linseed)
Agricultural Research Station
C.S. Azad University of Agril. & Tech
Jhansi
Uttar Pradesh

ID No.1373
Dr. Kenchanagoudar, P.V.
Groundnut Breeder
Main Research Station
University of Agricultural Sciences
Krishi Nagar
Dharwad-580 005
Karnataka

ID No.1374
Dr. Khadam, S.R.
Jr. Research Assistant
Oilseeds Research Scheme (Sunflower)
Zonal Agril. Research Station
Solapur-413 002
Maharashtra

ID No.1375
Dr. Khale, S.D.
Jr. Research Assistant
AICRP on Safflower
Agril. School Compound
Solapur-413 002
Maharashtra

ID No.1376
Dr. Khandhar, R.R.
429/B, Raijinagar Society
Street No. 14, Near Motibaug
Junagadh-362 001
Gujarat

ID No.1377
Dr. Khare, J.P.
C/o Ramesh Patnaik
Rampura Ward
Sagar-470 001
Madhya Pradesh

ID No.1378
Dr. Khire Arun, R.
Breeder (Groundnut)
Zonal Agricultural Research Station
Kharagone-451 001
Madhya Pradesh

ID No.1379
Dr. Kiransingh S. Raghuwanshi
Asst. Prof. of Soil Microbiology,
O/o Associate Director of Research,
97, Raviwar Peth, Near DAV College
Solapur-413 002
Maharashtra

ID No.1380
Dr. Kirasur, V.
Regional Manager,
Sandoz (India) Ltd.,
Shivaji Nagar,
Revenue Colony
Pune-411 001
Maharashtra

ID No.1381
Dr. Kiresur, V.R.
Head, Project Planning & Monitoring
Cell
University of Agril. Sciences
Krishinagar
Dharwad-580 005
Karnataka

ID No.1382
Dr. Kirtimayi Mishra
C/o Dr. C. Kale,
Lab of Molecular Biology and
Biotechnology
Orissa University of Agril. & Technology
Bhubaneshwar-3
Orissa

ID No.1383
Dr. Kishore Varma, R.
Scientist (Pl. Pathology),
Agril. Research Station, **ANGRAU**
Tandur-501 141
Andhra Pradesh

ID No.1384
Dr. Kochu Babu, M.
Director
NRC for Oilpalm,
Near Jawahar Navodaya Vidyalaya
Pedavegi-534 450
Andhra Pradesh

ID No.1385
Dr. Kohale, S.R.
Sr. Research Fellow, **Oilseeds**
Research Station, **Latur, (MS)**
Latur-413 512
Maharashtra

ID No.1386
Dr. Kolekar, N.M.
Research Associate,
Nimbkar Agril. Research Institute,
P.O. Box. No.44
Phaltan-415 523
Maharashtra

ID No.1387
Dr. Kolte, S.J.
Flat No.8, Anuprita Apartments,
S.No. 78, Plot No. 160, Left Bhusri
Colony, Near Telephone Exchange,
Paud Road, Kothrud
Pune-411 038
Maharashtra

ID No.1388
Dr. Koteswar Reddy
H.No. 3-4-456/304,
Venkata Sai Apartments,
Naryanaguda
Hyderabad-500 027
Andhra Pradesh

ID No.1389
Dr. Krishna Gopal Mandal
Scientist (Agronomy),
Indian Institute of Soil Science,
Nabi Bagh, Barasia Road
Bhopal-462 038
Madhya Pradesh

ID No.1390
Dr. Krishna Gouda, K.P.
PC Unit (Millets),
University of Agril. Sciences,
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1391
Dr. Krishna Reddy
Asst. Professor,
Dept. of Agronomy, **S.V. Agril. College,**
Tirupati, Chittoor Dt.
Tirupati-
Andhra Pradesh

ID No.1392
Dr. Krishna Reddy, R.
Scientist, J.N. Krishi Vishwa Vidyalaya
Zonal Agril. Research Station
Chhindwara
Madhya Pradesh

ID No.1393
Mr. Kubsad, V.S.
H.No.28,
Shivambrutha,
Krishi Nagar
Dharwad-580 001
Karnataka

ID No.1394
Dr. Kular, J.S.
Dept. of Entomology
Punjab Agricultural University
Ludhiana-141 001
Punjab

ID No.1395
Dr. Kuldeep Singh Dangi
Dept. of Genetics & Pl. Breeding,
College of Agriculture, **ANGRAU,**
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1396
Dr. Kuldip Raj Chopra
Director (Production),
Mahendra Hybrid Seeds Co. Ltd.,
P.B. No. 52, Mahaveer Marg
Jalna-431 203
Maharashtra

ID No.1397
Dr. Kulkarni, J.H.
Vice-Chancellor,
University of Agril. Sciences,
Krishinagar
Dharwad-580 005
Karnataka

ID No.1398
Dr. Kulkarni, M.S.
Assoc. Professor of Pl. Pathology,
KRC College of Horticulture
Arabhavi-591 310
Karnataka

ID No.1399
Dr. Kulkarni, R.S.
Professor,
Dept. of Plant Breeding,
University of Agril. Sciences,
GKVK campus
Bengaluru-560 065
Karnataka

ID No.1400
Dr. Kulkarni, S.N.
Assoc. Professor (Entomology),
College of Agriculture,
Ambayogai Dist, Beed
Beed
Maharashtra

ID No.1401
Dr. Kumar, K.
Asst. Oilseeds Breeder,
Dept. of Pl. Breeding,
N.D. University of Agril. & Technology,
Narendranagar, Kumarganj
Faizabad-223 229
Uttar Pradesh

ID No.1402
Dr. Kumar, P.R.
H.No. 203,
DDA SFS Flats,
Sector 3, Packet-2, Dwaraka
New Delhi-110 075
New Delhi

ID No.1403
Dr. Kumaran, K.
Associate Professor (Forestry),
Forestry College & Research Institute,
Mettupalayam-641 003
Tamil Nadu

ID No.1404
Mr. Kumaraswamy, H.H.
Scientist,,
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1405
Dr. Kumaresan, D.
Asst. Professor (Plant Breeding),
Regional Research Station,
Vridhachalam-606 001
Tamil Nadu

ID No.1406
Dr. Kunwar Harendra Singh
Scientist,
Directorate of Rapeseed-Mustard
Research
Sewar
Bharatpur-321 303
Rajasthan

ID No.1407
Dr. Kureel, R.S.
Joint Director of Secretary,
NOVOD Board, B6, Sector-18
Institutional Area
Gurgaon-122 015
Haryana

ID No.1408
Dr. Kushwinder Singh Brar
Plant Breeder
Punab Agril. University,
Regional Research Station,
Dabwali Road
Bhatinda-151 001
Punjab

ID No.1409
Dr. Lakshamma, P.
Sr. Scientist (Pl. Physiology),
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1410
Dr. Lakshmana Reddy, D.C.
Scientist (Biotechnology),
Division of Biotechnology,
Indian Institute of Horticultural
Research,
Hessargatta
Bengaluru-560 089
Karnataka

ID No.1411
Dr. Lakshmi Prayaga
Scientist (Plant Physiology),
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1412
Dr. Lakshmi Reddy, P.
Sr. Scientist (Pathology),
DATT Centre,
Anantapur-515 001
Andhra Pradesh

ID No.1413
Dr. Lakshminarayana, M.
Sr. Scientist (Entomology),
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1414
Dr. Laf, U.M.
H.No. 7-1-282-C/50,
Sreeramnagar,
B.K. Guda, Balkampet
Hyderabad-500 038
Andhra Pradesh

ID No.1415
Dr. Lallu
Physiologist (Oilseeds Section),
C.S. Azad University of Agril. & Tech.,
Kanpur-208 002
Uttar Pradesh

ID No.1416
Dr. Lalya Naik, R.
Tech. Assistant,
Castor Research institute
Hiriyur
Karnataka

ID No.1417
Dr. Lashminarayananamma, V.
Scientist (Ent.),
Regional Agril. Research Station,,
Palem, Mahaboobnagar, Dist.
Palem-509 215
Andhra Pradesh

ID No.1418
Dr. Latha, K.R.
Associate Professor (Agronomy),
B-45, Staff Quarters,
Tamil Nadu Agricultural University
Coimbatore-641 003
Tamil Nadu

ID No.1419
Dr. Lavanya, C.
Sr. Scientist (Plant Breeding),
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1420
Dr. Laxman L. Mane
Associate Professor,
Flat No. 6, Parakram Apartments,
1-9-4/1, Jaistnghpura
Aurangabad-431 004
Maharashtra

ID No.1421
Dr. Leela Rani, P.
H.No. 1-9-40, Plot No. 103,
Ravindra Nagar, Street No.8,
Habsiguda
Hyderabad-500 007
Andhra Pradesh

ID No.1422
Dr. Linga Raju, B.S.
Asst. Professor (Agronomy),
AICRP on Soybean,
University of Agril. Sciences
Dharwad-580 005
Karnataka

ID No.1423
Dr. Lingappa, S.
Director of Research (Retd.),
Biological Control Laboratory,
Dept. of Entomology, UAS,
Krishinagar
Dharwad-580 005
Karnataka

ID No.1424
Dr. Loganathan, P.
Jr. Breeder,
Oilseeds Research Station,
Tamil Nadu Agril. University
Tindivanam-604 001
Tamil Nadu

ID No.1425
Mr. Lokesha, K.R.
Research Associate,
Ramanna, P. Kadihall,
Garje Post, Kadur Taluk
Chikmagalur-577 140
Karnataka

ID No.1426
Dr. Lokesha, R.
Associate Professor,
College of Agriculture, P.B. No.24
Raichur-584 101
Karnataka

ID No.1427
Dr. Lukose, C.
Research Scientist (Pl. Pathology),
Oilseeds Section,
Junagadh Agril. University
Junagadh-362 001
Gujarat

ID No.1428
Dr. Rajashekhar M. **Kachapur**
No.49, Krishinagar,,
Kelgeri Road
Dharwad-580 001
Karnataka

ID No.1429
Dr. Madan Mohan, B.
S/o B. Appa Rao,
Palasapuram (Village),
Sompeta Mandal
Srikakulam-532 284
Andhra Pradesh

ID No.1430
Dr. Madan Prasad Srivastava
Asst. Botanist,
Pulses and Oilseeds Research Station
Berhampore-742 101
West Bengal

ID No.1431
Dr. Madaria, R.B.
Sr. Research Fellow,
Vardhman Society,
Zanzarda Road
Junagadh-362 001
Gujarat

ID No.1432
Dr. Madharia, S.K.
Jr. Scientist (Pl. Breeding)
Regional Agril. Research Station
Benori Seed Farm
Sagar-470 002
Madhya Pradesh

ID No.1433
Mr. Madhu, B.
Jr. Research Fellow,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1434
Ms. Madhuri, P.
Scientist,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1435
Mr. Madhusudhan Reddy, J.
Sr. Research Fellow,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1436
Dr. Mahadeva Reddy
Tech. Officer, AAS Unit,
Regional Agril. Research Station,
P.B. No. 24
Raichur-584 101
Karnataka

ID No.1437
Dr. Mahajan, A.M.
Sr. Research Asst.,
SRS (Oilseeds),
Crop Research Unit (Oilseeds),
Panjabrao Deshmukh Krishi Vidyapeeth
Akola-444 104
Maharashtra

ID No.1438
Dr. Maharaj Singh
Scientist (Pl. Physiology),
Directorate of Rapeseed-Mustard
Research
Sewar Farm
Bharatpur-321 001
Rajasthan

ID No.1439
Dr. Mahavishnan, K.
Research Associate,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1440
Dr. Mahendra Krishna Moon
Asst. Professor,
Indira Nagar Housing Society,
Malkapur
Akola-444 104
Maharashtra

ID No.1441
Dr. Mahindra Singh Pal
Associate Professor (Agronomy),
G.B. Pant University of Agril. & Tech.
Pan Nagar-263 145
Uttar Pradesh

ID No.1442
Dr. Maithi, S.
Director,
National Research Centre for Aromatic
& Medicinal Plants
Anand
Gujarat

ID No.1443
Dr. Majumdar, J.C.
General Manager,
Divisional Tech. Services, 7,
Basant Lok, Vasant Vihar
New Delhi
New Delhi

ID No.1444
Dr. Makne, V.G.
Oilseed Specialist,
Engineering College,
Ausa Road
Latur-413 512
Maharashtra

ID No.1445
Dr. Malam Singh
Asst. Professor (Agronomy),
46, Hanwant "A", B.J.S. Colony,
Near Bagawarshop
Jodhpur-342 010
Rajasthan

ID No.1446
Dr. Malani Ramanikbhai, P.
Proprietor,
Ganesh Agri Seeds,
109, Ashwamegh Avenue,
Near Mithakhali, Under Bridge

ID No.1447
Dr. Mali Dnyaneshwar Vishnu
Sr. Research Assistant (Oilseeds),
Dr. P.D. Krishi Viswa Vidyalya
Akola-444 104
Maharashtra

ID No.1448
Dr. Malichand Hirachand Chauhan
Oilseed Specialist,
Oilseed Research Station
Latur-413 512
Maharashtra

ID No.1449
Dr. Malik, G.C.
Lecturer (SS in Agronomy),
Palli Siksha Bhavana
(Institute of Agriculture),
Viswa-Bharati, Sriniketan
Birbhum-731 236
West Bengal

ID No.1450
Dr. Malik, R.S.
Project I/c, Oilseeds,
Dept. of Genetics & Pl. Breeding,
Indian Agril. Research Institute, Pusa
New Delhi-110 001
New Delhi

ID No.1451
Dr. Malik, Y.P.
Sr. Scientist (Ent.),
P.C. Unit (Linseed),
C.S. Azad University of Agril. & Tech.,
Kanpur-208 002
Uttar Pradesh

ID No.1452
Dr. Mallapur, C.P.
Director of Research,
University of Agril. Sciences,
Krishinagar
Dharwad-580 005
Karnataka

ID No.1453
Dr. Malligwad Lokanath, H.
Jr. Agronomist (Groundnut),
Oilseeds Scheme, Main Research
Station
University of Agril. Sciences,
Krishinagar
Dharwad-580 005
Karnataka

ID No.1454
Dr. Manak Singh
Jr. Breeder (Oilseeds),
Section of Oilseeds,
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1455
Dr. Mandal, S.
Biochemist,
National Bureau of Plant Genetic
Research, Pusa Campus
New Delhi-110 012
New Delhi

ID No.1456
Dr. Mangala Rai
Retd. Secretary, DARE & DG, ICAR
Indian Council of Agril. Research
Krishi Bhawan
New Delhi-110 001
New Delhi

ID No.1457
Dr. Mangesh Y. Dudey
Scientist,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1458
Dr. Manickam, S.
Asst. Professor (Agronomy),
Tapica Castor Research Station
Yethapur-636 119
Tamil Nadu

ID No.1459
Dr. Manikantan, M.R.
Scientist,
CIPHET, ICAR,
Punjab Agril. University
Ludhiana-141 004
Punjab

ID No.1460
Dr. Manisegaran, S.
909, Bavi Sundar Illam,
Valar Nagar,
Uthangudi (Post)
Madurai-625 104
Tamil Nadu

ID No.1461
Dr. Manish Yashvantro Ladole
Asst. Professor,
At: P. Sirasgaon Kasha,
Tq: Chandur Bazar, Amaravati District
Maharashtra

ID No.1462
Dr. Manivannan, N.
Associate Professor (Pl. Breeding),
Dept. of Oilseeds,
Tamil Nadu Agril. University
Coimbatore-641 003
Tamil Nadu

ID No.1463
Dr. Manivel, P.
Principal Scientist (Pl. Breeding),
National Research Centre for Medicinal
& Aromatic Plants
Anand
Gujarat

ID No.1464
Dr. Manjaya
Scientific Officer,
Nuclear Agril. & Biotechnology Division,
Bhabha Atomic Research Centre,
Trombay
Mumbai-400 085
Maharashtra

ID No.1465
Dr. Manjeet Kaur Sangha
Asst. Biochemist,
Dept. of Plant Breeding &
Biotechnology,
Punjab Agril. University
Ludhiana-141 001
Punjab

ID No.1466
Dr. Manjunath, G.O.
Asst. Professor,
College of Forestry
Sirsi
Karnataka

ID No.1467
Dr. Manjunatha Hebbar
Soil Scientist,
Agril. Research Station,
University of Agril. Sciences
Gangavathi-583 227
Karnataka

ID No.1468
Dr. Manoharan, V.
Professor (Agril. Botany),
Oilseeds Research Station
Tindivanam-604 001
Tamil Nadu

ID No.1469
Dr. Manoj Kumar **Dhurve**
Scientist,
Oilseeds Section
Raipur-492 006
Chhattisgarh

ID No.1470
Dr. Manoj Kumar
Sr. Scientist,
Central Potato Research Station,
Post: Sahay Nagar
Patna-801 506
Bihar

ID No.1471
Dr. Manoranjitham, S.K.
Asst. Professor,
Dept. of Plant Pathology,
Tamil Nadu Agril. University
Coimbatore-3
Tamil Nadu

ID No.1472
Mr. Maruthi, V.
Scientist,
Central Research Institute for **Dryland**
Agriculture,
Santoshnagar, Saidabad
Hyderabad-500 659
Andhra Pradesh

ID No.1473
Dr. Mary Kamala, P.
SPF, AICRP on Sunflower,
Regional Agril. Research Station,
Noonepalli
Nandyal-518 503
Andhra Pradesh

ID No.1474
Dr. Mathur, R.K.
Sr. Scientist (Pl. Breeding),
Directorate of Oilpalm Research
Pedavegi, W.G. Dist.
Pedavegi
Andhra Pradesh

ID No.1475
Dr. Mathur, Y.K.
Prof. of Entomology,
176, Suryanagar,
Gopal Pura Bypass
Jaipur-302 015
Rajasthan

ID No.1476
Dr. Mathuria, O.P.
Oilseeds Section (**Linseed**),
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1477
Dr. Maurya, C.L.
Asst. Professor,
Dept. of Seed Technology,
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1478
Dr. Mayee, C.D.
Chairman,
Agril. Scientists' Recruitment Board,
Krishi Anusandhan **Bhavan-II**,
Pusa, IARI Campus
New Delhi-110 012
New Delhi

ID No.1479
Dr. Mazumdar, B.
General Manager (R&D),
Safal Seeds & Biotech Ltd.,
D/102, Raj Heights, **Jalna Raod, CIDCO**
Aurangabad-431 003
Maharashtra

ID No.1480
Dr. Meera Srivastava
Section of Economic Botanist,
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1481
Dr. Mehta, K.S.
Director,
Bharat Pulversing Mills P. Ltd.,
Sriniketan, 14, Queens Road,
Church Gate
Mumbai-400 020
Maharashtra

ID No.1482
Ms. Mehtab Yasmeen
Research Associate,
H.No. 16-7-213, **Azampura, Chaderghat**
Hyderabad-500 024
Andhra Pradesh

ID No.1483
Dr. Men, U.B.
Associate Professor (Ent.),
3, Ravi Nagar, Near Santaji Colony
Akola-444 105
Maharashtra

ID No.1484
Dr. Mev Singh
C/o Dr. G.P. Lodhi,
Principal **Brahmanand**
Vedik College
Ameerpur
Uttar Pradesh

ID No.1485
Dr. Michael Pienyii
SMS (Entomology),
Krishi Vigyan Kendra
Kohima-797 001
Nagaland

ID No.1486
Dr. Milal Sopan Rao Supe
Jr. Research Assistant,
42, Radhakrishna Nagar,
Murtizapur Road
akola-444 001
Maharashtra

ID No.1487
Dr. Mishra, P.K.
Regional Agril. Research Station
Chindwara
Maharashtra

ID No.1488
Dr. Misra, J.B.
Director,
National Research Centre for
Groundnut,
Innagar Road, P.B. No.5
Junagadh-362 001
Gujarat

ID No.1489
Dr. Mohammad M. Anwar
11-5-147,
Red Hills
Hyderabad-500 004
Andhra Pradesh

ID No.1490
Dr. Mohan Bentur
C/o G.N. Bentur,
H.No. 6, Saraswaripur,
Gokul Road
Hubli-580 030
Karnataka

ID No.1491
Dr. Mohan Chavan
Sr. Agronomist,
Main Agril. Research Station,
University of Agril. Sciences
Raichur-584 102
Karnataka

ID No.1492
Dr. Mohan Raju, B.
Jr. Plant Physiologist,
Dept. of Crop Physiology,
University of Agril. Sciences,
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1493
Dr. Mohan Rao, A.
Asst. Professor,
Dept. of Plant Breeding & Genetics,
University of Agril. Sciences,
GKVK
Bengaluru-560 065
Karnataka

ID No.1494
Mr. Mohd. Ashraf Achfan
Flat No. 302A,
Akbar Towers,
New Malakpet
Hyderabad-500 038
Andhra Pradesh

ID No.1495
Dr. Mohinder Singh
Asst. Breeder (Oilseeds),
Dept. of Pl. Breeding,
Punjab Agril. University
Ludhiana-141 004
Punjab

ID No.1496
Dr. Momin, S.A.
Reader in Oils,
Nathalal Parekh Marg,
Matunga
Mumbai-400 019
Maharashtra

ID No.1497
Dr. Monpara, B.A.
Associate Professor (Pl. Breeding),
Vallabh Nagar, Anand Nagar,
Main Road, Timbawadi
Junagadh-362 015
Gujarat

ID No.1498
Dr. Moradia, A.M.
Nematologist,
Junagadh Agril. University
Junagadh-362 001
Gujarat

ID No.1499
Dr. Motagi, B.N.
Research Associate,
Seed Research Laboratory,
Seed Unit, University of Agril. Sciences
Dharwad-580 005
Karnataka

ID No.1500
Dr. Mothilal, A.
Asst. Professor (PB)
Regional Research Station
Vridhachalam-606 001
Tamil Nadu

ID No.1501
Dr. Motilal Verma
Retd. Sr. Scientist (Pl. Path)
Vill: Madhi (Near Hotel Shark Inn)
Post: Atariya,
Rewa-486 446
Maharashtra

ID No.1502
Dr. Muddalagiriappa
Asst. Professor (Agronomy)
Krishi Vignana Kendra
University of Agril. Sciences
P.B. No. 24
Raichur-584 101
Karnataka

ID No.1503
Dr. Mukesh Kumar Saxena
Tech. Asst. (Pl. Breeding),
G-10 (OTC),
College of Agril. Campus
Indore
Madhya Pradesh

ID No.1504
Dr. Mukta, N.
Senior Scientist
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1505
Dr. Mukund N. Mohrir
Sr. Breeder,
Nimbkar Seeds P. Ltd.,
Jinti Bridge
Phaltan-415 525
Maharashtra

ID No.1506
Dr. Mundada, R.V.
Karwa Nagar,
Vithal,
SRPF Road
Jalna-431 203
Maharashtra

ID No.1507
Dr. Murali Arthanari, P.
67/62, West Street No. 2.,
Kumarasampatty,
Hasthampatty (Post)
Salem-636 007
Tamil Nadu

ID No.1508
Dr. Muralidharan, V.
Prof. & Head, TNAU,
No. 2, Tauta Nagar,
Vadavalli
Coimbatore-600 041
Tamil Nadu

ID No.1509
Dr. Muralidharudu, Y.
Project Coordinator,
Central Institute for Agril. Engineering
Bhopal-462 038
Madhya Pradesh

ID No.1510
Dr. Murthy, I.Y.L.N.
Principal Scientist,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1511
Dr. Murthy, K.S.R.K.
Agril. Scientist,
H.No. 3-81, NR. Nagar,
Malkajigiri
Hyderabad-500 047
Andhra Pradesh

ID No.1512
Dr. Murthy, T.G.K.
Scientist,
National Research Centre for
Groundnut,
P.B. No.5, Ivnagar Road
Junagadh-362 001
Gujarat

ID No.1513
Mr. Murumkar Dattatraya Ranganath
Jr. Plant Pathologist,
AICRP (Safflower),
Agril. School Compound,
93, Bhavanipeth
Solapur-413 002
Maharashtra

ID No.1514
Dr. Mutalik Desai, S.G.
Head, Emergent Genetics,
Presaria C-8, Dt. Nashangaon,
Phase-I, Keshwarsee Nagar
Aurangabad
Maharashtra

ID No.1515
Dr. Mutkule, D.S.
Jr. Entomologist,
Oilseeds Research Station
Latur
Maharashtra

ID No.1516
Dr. Nadaf, H.L.
Breeder (NSP),
Main Research S
University of Agri.
Krishinagar
Dharwad-580 005
Karnataka

ID No.1517
Dr. Nagabhushanam, G.V.S.
1-68/3, Plot No. 28.,
Prabhat Nagar,
Chaitanyapuri, Dilsukhnagar
Hyderabad-500 660
Andhra Pradesh

ID No.1518
Dr. Nagalikar, V.P.
Sr. Farm Superintendent,
Regional Research Station
Raichur-584 101
Karnataka

ID No.1519
Dr. Nagaraj, G.
210, Revathi Twoers,
Maruthinagar, Kothapet
Hyderabad-500 060
Andhra Pradesh

ID No.1520
Dr. Nagarajaiah, K.M.
Jr. Agronomist,
AICRP (Sunflower),
University of Agril. Sciences, GKVK
Bengaluru-560 065
Karnataka

ID No.1521
Dr. Nagaraju
Professor & Head (Pl. Pathology),
No.237, 7th Main Road,
Kalyan Coop. Housing Layout,
Vijayanagar-II Stage (RPS)
Bengaluru-560 040
Karnataka

ID No.1522
Dr. Nagartna, T.K.
Asst. Professor (SS),
University of Agril. Sciences,
AICRP on Sunflower, GKVK
Bengaluru-560 065
Karnataka

ID No.1523
Dr. Nageswar Rao, T.G.
Principal Scientist (Pl. Pathology),
National Research Centre for Sorghum,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1524
Dr. Nageswara Rao, R.C.
Scientist (Physiology),
ICRISAT, Patancheru
Medak District
Patancheru-502 324
Andhra Pradesh

ID No.1525
Dr. Naik, K.S.S.
Scientist (Pl. Breeding),
Agril. Research Station, ANGRAU,
Kadiri, Dt. Anantapur
Kadiri-515 591
Andhra Pradesh

ID No.1526
Dr. Nalawade, S.K.
Jr. Research Asst.,
Oilseeds Research Station,
Marathwada Agril. University
Latur-413 512
Maharashtra

ID No.1527
Dr. Nalini Tewari
Asst. Professor,
CS Azad University of Agril. & Tech.
Kanpur-208 003
Uttar Pradesh

ID No.1528
Dr. Nallathambi, G.
Professor & Head,
Regional Agril. Research Station,
Tamil Nadu Agril. University
Vridhachalam-606 001
Tamil Nadu

ID No.1529
Mr. Nanda Kumar, R.
SRS
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1530
Dr. Nandagopal, V.
Division of Entomology,
Central Rice Research Institute
Cuttack
Orissa

ID No.1531
Dr. Nandane Anil, S.
Sr. Research Fellow,
C/o G.S. Patil, Omkar Niwas,
Bhagyalaxmi Nagar
Parbhani-431 401
Maharashtra

ID No.1532
Dr. Nandeha, K.L.
Sr. Scientist (Agronomy),
TCB College of Agriculture & Res.
Station,
Srikanda
Bilaspur
Chhattisgarh

ID No.1533
Dr. Nandini Devi, K.
Jr. Agronomist,
Central Agril. University
Imphal-795 004
Manipur

ID No.1534
Dr. Nandkishor V. Rao Larhe
Asst. professor (Entomologist),
Dr. P.D. Krishi Vidyapeeth
Amaravati Road, Wadi
Nagpur-440 023
Maharashtra

ID No.1535
Dr. Nanja Reddy, Y.A.
Associate Professor,
College of Agriculture,
University of Agril. Sciences
Raichur-584 101
Karnataka

ID No.1536
Dr. Nanjappa, H.V.
Professor of Agronomy,
Dept. of Agronomy,
University of Agril. Sciences,
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1537
Mr. Narasimha Rao, N.
S/o N. Subba Rao,
Srigangapuram Post,
Chebrolu Mandal, Guntur District
Chebrolu
Andhra Pradesh

ID No.1538
Dr. Narasimha Rao, S.B.S.
Scientist (Agronomist),
AgroMet Cell,
Agril. Research Institute, ANGRAU,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1539
Dr. Narayanamma
Sr. Scientist (Agronomy),
Agril. Research Institute, ANGRAU,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1540
Dr. Narender Reddy, S.
Plant Physiologist,
Regional Agril. Research Station
Jagtial-505 327
Andhra Pradesh

ID No.1541
Dr. Narender Singh Sachan
Scientist (Pathology),
Oilseeds Section,
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1542
Dr. Narendra, B.
Sunflower Breeder,
Sunflower Project, **Regional Agril.**
Research Station
Nandyal
Andhra Pradesh

ID No.1543
Dr. Nargund, V.B.
Professor (Pl. Pathology),
University of Agril. Sciences,
College of Agriculture
Dharwad-580 005
Karnataka

ID No.1544
Mr. Narinder Kaur
SRF,
H.No. 142-D,
Kitchlu Nagar
Ludhiana-141 004
Punjab

ID No.1545
Dr. Natarajamoorthy, K.
Sunflower Breeder,
Agril. Research Station,
Tamil Nadu Agril. University
Bhavanisagar-638 451
Tamil Nadu

ID No.1546
Dr. Nautiyal, P.C.
Sr. Scientist (Pl. Physiology),
P.B. No.5,
NRC for Groundnut, Ivnagar Road
Junagadh-362 002
Gujarat

ID No.1547
Dr. Navkhat, D.A.
Asst. Professor (Agronomy),
Marathwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1548
Dr. Neelima, S.
Sunflower Breeder,
D/o S. Sankar Reddy,
Royal Public School, Ring Road
Ramavarappadu-521 108
Andhra Pradesh

ID No.1549
Dr. Nekar, M.M.
C/o T.B. Chilakwad Basan Prabhu,
Basavshant Nagar,
Yellakki Shettar Colony,
Near Shankar Math
Dharwad-580 004
Karnataka

ID No.1550
Dr. Nigam, S.N.
Principal Groundnut Breeder,
ICRISAT
Patancheru-502 324
Andhra Pradesh

ID No.1551
Dr. Nikam, R.R.
Agronomist,
Linseed Research Project,
College of Agriculture
Nagpur-440 010
Maharashtra

ID No.1552
Dr. Nikhil Shrivastava
Sr. Scientist (Ent.),
PC Unit (S&N),
J.N. Krishi Vishwa Vidyalyaya
Jabalpur-482 004
Madhya Pradesh

ID No.1553
Dr. Ninganur, B.T.
Sr. Farm Superintendent,
Main Agril. Research Station
University of Agril. Sciences
Dharwad-580 005
Karnataka

ID No.1554
Dr. Niraj Daga
Director,
Betul Oils & Flours Ltd.,
Kosmi Industrial Area
Betul-460 001
Maharashtra

ID No.1555
Dr. Nirmal, S.V.
Jr. Physiologist,
AICRP (Safflower)
Agricultural School Compound
Solapur
Maharashtra

ID No.1556
Dr. Nirmalakumari
Associate Professor (Pl. Breeding)
Dept. of Millets, CPBG,
Tamil Nadu Agril. University
Coimbatore-641 003
Tamil Nadu

ID No.1557
Dr. Nitesh N. Prajapati
SRF, TBO Project,
AICRP on UUC,
S.D. Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1558
Dr. Om Prakash Premi
Scientist,
NRC for Rapeseed-Mustard,
Post: Sewar
Bharatpur-321 303
Rajasthan

ID No.1559
Dr. Om Prakash
PC Unit (Linseed),
C.S. Azad University of Agril. & Tech
Kanpur-208 002
Uttar Pradesh

ID No.1560
Dr. Omkarappa, T.
Jr. Breeder (Castor),
AICRP (Castor),
Agril. Research Station,
University of Agril. Sciences
Hiriyur-572 143
Karnataka

ID No.1561
Dr. Padihar, S.K.
Jr. Scientist, Zonal Agril. Research
Station, Powerkheda, Hoshangabad,
461, 110
Hoshangabad-461 110

ID No.1562
Dr. Padma Raju, A.
Director of Research (Retd.),
B-18, Varsha Society,
Karolbagh Road, Padmanabnagar
Hyderabad-
Andhra Pradesh

ID No.1563
Dr. Padmaiah, M.
Sr. Scientist,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1564
Dr. Padmaja Rao, S.
Principal Scientist
Seed Technology Division,
Sugarcane Breeding Institute
Coimbatore-641 003
Tamil Nadu

ID No.1565
Dr. Padmalatha, Y.
Scientist,
D.No. 14-432,
Kamala Nagar
Anantapur-515 001
Andhra Pradesh

ID No.1566
Dr. Padmavathi, N.
USA

ID No.1567
Ms. Padmavathi, P.
Senior Scientist (Agronomy),
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1568
Dr. Pali, G.P.
Jr. Scientist (Agronomy)
Oilseed Section
Indira Gandhi Krishi Vishwa Vidyalaya
Raipur-492 012
Uttar Pradesh

ID No.1569
Ms. Pallavi, M.
Sr. Research Fellow
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

No.1570
Pandey Anil
Breeder (Oilseeds & R-M),
pt. of Pl. Breeding,
hut College of Agriculture, Dholi
Jazafarpur-843 121

ID No.1571
Mr. Pankaj Kumar
Asst. Commandant,
Kutch B.P.F.,
Gandhidham, Adipur, Dt.
Kutch-370 205
Gujarat

ID No.1572
Dr. Pankaj Reddy
Principal Scientist,
NRC for Sorghum,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1573
Dr. Paramathma, M.
Professor & Head,
Forest College & Research Institute,
Tamil Nadu Agril. University
Mettupalayam-641 301
Tamil Nadu

ID No.1574
Dr. Parameshwarappa, K.G.
Sr. Scientist (Oilseeds),
Main Research Station,
University of Agril. Sciences
Dharwad-580 005
Karnataka

ID No.1575
Dr. Parameswari, C.
Pushpam 30,
Maruti Pandayar Street,
KK Nagar
Madurai-625 020
Tamil Nadu

ID No.1576
Dr. Parashar, R.R.
Breeder (Oilseeds),
Zonal Agril. Research Station,
Powerkheda
Hoshangabad-461 110
Madhya Pradesh

ID No.1577

Dr. Paravinder Sheoran
Asst. Agronomist,
Oilseeds Section,
Punjab Agril. University
Ludhiana-141 004
Punjab

ID No.1578

Dr. Parlekar, G.Y.
Asst. Professor of Entomology,
Avantinagar, F-2 Building,
Near Old Pune Nala
Solapur-413 001
Maharashtra

ID No.1579

Dr. Parmar Pinakin Khedabhai
Sr. Research Fellow,
F-23/4, S.D. Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1580

Dr. Parminder Kaur
Research Fellow (Pl. Path),
Dept. of Plant Breeding,
Punjab Agril. University
Ludhiana-141 004
Punjab

ID No.1581

Dr. Paroda, R.S.

ID No.1582

Dr. Parsana, G.C.
Agril. Supervisor,
Agril. Research Station,
Gujarat Agril. University
Amreli-365 601
Gujarat

ID No.1583

Dr. Parthasarathy, V.A.
Director,
Indian Institute of Spices **Research**,
P.B. No. 1701, Marrikunnu
Calicut-673 012
Kerala

ID No.1584

Dr. Parthiban, K.T.
Asst. Professor,
Tamil Nadu Agril. University
Mettupalayam-641 301
Tamil Nadu

ID No.1585

Dr. Paslawar, A.N.
Jr. Agronomist,
AICRP on Safflower,
Dr. P.D. Krishi Vidyapeeth
Akola-444 104
Maharashtra

ID No.1586

Dr. Patel Babubhai Shankardas
Asst. Research Scientist,
Main Castor & Mustard Research
Station,
Gujarat Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1587

Dr. Patel, B.K.
Main Castor & Mustard,
Research Station,
Gujarat Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1588

Dr. Patel, D.B.
Asst. Research Scientist (**Pathology**),
Main Castor & Mustard **Research**
Station,
S.D. Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1589

Dr. Patel, D.P.
Scientist (Pl. Physiology),
Division of Agronomy,
ICAR Research Complex **for NEH**
Region, Umrol Road
Umlam-793 103

ID No.1590

Dr. Patel, G.A.
Jr. Pathologist,
AICRP (Castor),
Main Castor & Mustard **Research**
Station,
S.D. Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1591

Dr. Patel, J.B.
Research Scientist,
Main Pulse Research Station,
Gujarat Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1592

Dr. Patel, K.N.
Oilseeds Research Unit,
Dr. Punjabrao Krishi Viswa **Vidyalaya**
Akola-444 104
Maharashtra

ID No.1593

Dr. Patel, K.R.
Research Scientist (C&M),
Main Castor & Mustard **Research**
Station,
Gujarat Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1594

Dr. Patel, K.S.
Asst. Research Scientist,
Main Castor-Mustard **Research Station**,
Gujarat Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1595
Dr. Patel, M.K.
Asst. Research Scientist (**Soil Science**),
Main Castor-Mustard Research Station,
Gujarat Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1596
Dr. Patel, P.S.
Asst. Research Scientist,
Regional Research Station,
Gujarat Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1597
Dr. Patel, R.M.
Jr. Agronomist,
Main Castor & Mustard Research
Station,
S.D. Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1598
Dr. Patel, R.S.
Research Scientist,
Oilseeds Research Station,
Gujarat Agril. Univrsity
Junagadh-362 001
Gujarat

ID No.1599
Dr. Patel, S.
Associate Professor (Agril.),
S.D. College of Agriculture ,
Kumharawand Farm
Jagdarpur-494 005
Chhattisgadh

ID No.1600
Dr. Patel, S.R.
Sr. Scientist,
Dept. of Agrometerology,
Regional Agril. Research Station,
I.G. Krishi Vishwa Vidyalaya
Raipur-492 006
Chhattisgadh

ID No.1601
Dr. Pathak, H.C.
Director of Research,
Navsari Agril. University
Navsari-396 450
Gujarat

ID No.1602
Dr. Pati, D.
Technical Information Officer,
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1603
Dr. Patil Chandrakant **Babu Rao**
C/o B.T. Bhogaonkar,
Vrindavana Colony
Parbhani-431 401
Maharashtra

ID No.1604
Dr. Patil M. Ravindra
Manager (R&D),
Agsun Seeds (India) Pvt. Ltd.,
Plot No. 32, IInd Stage,
Mundargi Industrial Area
Bellary-583 102
Karnataka

ID No.1605
Dr. Patil, A.J.
Breeder (Safflower),
AICRP (Safflower),
Agril. School Compound
Solapur-413 002
Maharashtra

ID No.1606
Dr. Patil, B.K.
Jr. Agronomist (**Safflower**),
Marahwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1607
Dr. Patil, B.N.
Sr. Scientist (**Seed Tech. Research**),
Dr. P.D. Krishi Viswa Vidyalaya
Akola-444 001
Maharashtra

ID No.1608
Dr. Patil, B.R.
Associate Professor (**Botany**),
Punjabrao Krshividyaapeeth,
Crop Research Unit (**Oilseeds**)
Akola-444 104
Maharashtra

ID No.1609
Dr. Patil, B.V.
Director of Instructions (Agril.),
College of Agriculture,
Regional Research Station
University of Agril. Sciences
Raichur-584 101
Karnataka

ID No.1610
Dr. Patil, B.V.
Jr. Entomologist (**Sunflower**),
AICRP (Sunflower),
Oilseeds Research Static
Latur-444 104
Maharashtra

ID No.1611
Dr. Patil, H.S.
Asst. Professor of Botany,
College of Agriculture
Dhule-424 004
Maharashtra

ID No.1612
Dr. Patil, M.S.
110, Panchavati Society,
Near Railway Crossing,
Ahmedabad High
Palanpur-385 001
Gujarat

ID No.1613
Dr. Patil, P.V.
Pathologist, AICRP on Soybean,
Main Research Station,
University of Agril. Sciences
Dharwad-580 005
Karnataka

ID No.1616
Dr. Patil, S.A.
Director,
Indian Agril. Research Institute
Pusa Campus
New Delhi-110 012
New Delhi

ID No.1619
Dr. Patil, V.C.
Prof. & Head,
Dept. of Agronomy, College of
Agriculture,
University of Agril. Sciences
Krishi Nagar
Dharwad-580 005
Karnataka

ID No.1622
Dr. Paulkar, K.S.
Associate Professor (Breeding),
Punjabrao Krishi Vidyapeeth,
Crop Research Unit (Oilseeds)
Akola-444 104
Maharashtra

ID No.1625
Dr. Pawankumar
Sr. Scientist (Oilseeds),
Agril. Research Institute,
Lohinagar
Patna-800 020
Bihar

ID No.1628
Dr. Poonguzhalan, R.
Associate Professor (Agronomy),
P.J.N. College of Agriculture &
Research Institute
Karfaikal-609 603
Pondicheri

ID No.1614
Dr. Patil, R.H.
Entomologist,
University of Agril. Sciences,
Main Research Station
Dharwad-580 005
Karnataka

ID No.1617
Dr. Patil, S.H.
Bhabha Atomic Research Centre
Trombay
Mumbai-400 035
Maharashtra

ID No.1620
Dr. Patil, V.D.
Asst. Director General (OP),
Indian Council of Agril. Research,
Krishi Bhawan
New Delhi-110 001
New Delhi

ID No.1623
Mr. Pavanchandra Reddy
Ph.D. Student,
P.G. Hostel,
ANGRAU, Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1626
Dr. Phool Chand
Jr. Scientist-cum-Asst. Professor,
Dept. of Plant Pathology,
Muzaffarpur TCA
Dholi-843 121
Bihar

ID No.1629
Dr. Potdurkhe, N.R.
Associate Professor (Oilseeds),
Sunflower Seed Production,
Oilseeds Research Station,
Dr. P.D. Krishi Viswa Vidyalyaya
Akola-444 104
Maharashtra

ID No.1615
Dr. Patil, R.K.
Prof. of Entomology,
Dept. of Entomology,
University of Agril. Sciences,
Krishi Nagar
Dharwad-580 005
Karnataka

ID No.1618
Dr. Patil, S.N.
Jr. Agronomist,
B-1, Sudhir Colony
Akola-444 104
Maharashtra

ID No.1621
Dr. Patra, G.J.
Dept. of Pl. Breeding and Genetics,
Orissa University of Agril. & Tech.
Bhubaneswar-751 001
Orissa

ID No.1624
Dr. Pawan Kumar
Scientist,
Agril. Research Station
Patna
Bihar

ID No.1627
Dr. Poonam Singh
Associate Professor, NATP,
Dept. of Seed Science & Technology,
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1630
Mr. Prabhakara Rao, N.
Head (FOM),
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1631

Dr. Prabhakara Reddy, G.
Asst. Professor (Agronomy)
S.V. Agril. College
Tirupati-517 002
Andhra Pradesh

ID No.1632

Dr. Prabhakaran, A.J.
Principal Scientist,
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1633

Dr. Prabhijyot Kaur
Asst. Agrometeorologist,
Dept. of Agronomy,
Punjab Agril. University
Ludhiana-141 004
Punjab

ID No.1634

Dr. Prabhjadh Singh Sandhu
Asst. Plant Pathologist,
Oilseeds Section,
Punjab Agril. University
Ludhiana-141 004
Punjab

ID No.1635

Dr. Prabhu Dayal Meena
Scientist (Pl. Pathology),
Directorate of Rapeseed Mustard
Research
Sewar
Bharatpur-321 303
Rajasthan

ID No.1636

Dr. Prabhu, B.
Director, Basaran Biocon (India) Ltd.,
24/7, Dr. Radhakrishnan Salai,
Palanippinagar, Valasaravakkam
Chennai-600 087
Tamil Nadu

ID No.1637

Dr. Pradeep Kumar Singh
Asst. Professor (Pl. Breeding)
ND University of Agril. & Tech,
Narendranagar, Kumarganj
Faizabad-224 229
Uttar Pradesh

ID No.1638

Dr. Pradhan, H.R.
Addl. Director of Agriculture
Lambuchera
Sikkim

ID No.1639

Dr. Prafulla Kumar Das
Breeder (Groundnut),
A-304, Rashmi Vihar,
Buddheswari, PO: Lane, Laxmi Sagar
Bhubaneswar-751 006
Orissa

ID No.1640

Dr. Pramila Rani, B.
Scientist (Agronomy),
Regional Agril. Research Station,
Lam
Guntur-522 034
Andhra Pradesh

ID No.1641

Dr. Pramod Katti
Jr. Entomologist (Sunflower),
Regional Agril. Research Station
University of Agril. Sciences,
P.B. No. 24
Raichur-584 101
Karnataka

ID No.1642

Dr. Prasad, M.V.R.
H.No. 6-3-712/98/B (93 RT),
Sowbhagya Nilayam,
Panjagutta Housing Board Colony
Hyderabad-500 004
Andhra Pradesh

ID No.1643

Dr. Prasad, R.B.N.
Scientist,
Indian Institute of Chemical Technology,
Tarnaka
Hyderabad-500 007
Andhra Pradesh

ID No.1644

Dr. Prasad, R.D.
Sr. Scientist,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1645

Dr. Prasad, Y.G.
Sr. Scientist,
Central Research Institute for, Dryland
Agriculture,
Santoshnagar, Saidabad
Hyderabad-500 659
Andhra Pradesh

ID No.1646

Dr. Prasant N. Mane
Asst. Professor,
Oilseeds Research Unit,
Dr. P.D. Krishi Vidyapeeth
Akola-444 104
Maharashtra

ID No.1647

Dr. Pratibha, G.
Sr. Scientist (Agronomy),
Central Research Institute for
Dryland Agriculture,
Santoshnagar, Saida
Hyderabad-500 659
Andhra Pradesh

ID No.1648

Dr. Praveen Rao, V.
Professor (Agronomy),
College of Agriculture,
ANGRAU, Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1649

Dr. Pravin Kashiram Rathod
Associate Professor (Ent.),
Opp: Oilseeds Research Unit,
Dr. P.D. Krishi Vidyalaya
Akola-444 104
Maharashtra

ID No.1650

Dr. Prithviraj, K.
H.No. 10-32,
Nethaji Road,
Narasam Pet (Post)
Warangal-506 132
Andhra Pradesh

ID No.1651

Dr. Pujar, B.T.
Associate Director of Research
Regional Agril. Research Station
P.B. No. 24
Raichur-584 101
Karnataka

ID No.1652

Dr. Pujar, M.F.
Asst. Professor of Agronomy,
Main Agril. Research Station,
University of Agril. Sciences
Dharwad-580 005
Karnataka

ID No.1653

Dr. Pushp Sharma
Oilseeds Section
Punjab Agril. University
Ludhiana-141 004
Punjab

ID No.1654

Dr. Puttaranga Swamy, K.T.
Associate Professor,
Genetics & Plant Breeding,
AICRP on Sunflower,
University of Agril. Sciences
Bengaluru-560 065
Karnataka

ID No.1655

Dr. Radadia, B.V.
Associate Research Scientist (Ent.),
Agril. Research Station,
Gujarat Agril. University
Amreli-365 601
Gujarat

ID No.1656

Dr. Radha Kishan, T.
Sr. Scientist,
Directorate of Groundnut Research
Innagar Road, P.B. No.5
Junagadh-362 001
Gujarat

ID No.1657

Dr. Radha Krishna Murthy, V.
Asst. Professor,
Dept. of Agronomy,
College of Agriculture, ANGRAU,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1658

Dr. Ragavan, T.
Asst. Professor (Agronomy),
Agril. Research Station
Kovilpatti-628 501
Tamil Nadu

ID No.1659

Dr. Raghavaiah, C.V.
Principal Scientist,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1660

Mr. Raghavendra, M.
Plant Pathologist
Chilli Research Station
University of Agril. Sciences
Devihosur
Haveri
Karnataka

ID No.1661

Mr. Raghunath, G.
Technical Officer,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1662

Dr. Raghupathi Reddy, K.
Plot No. 12,
Ventur 1,
Mallareddi Nagar,
Lothukunta
Secunderabad-500 015
Andhra Pradesh

ID No.1663

Dr. Raghuwanshi, K.M.S.
Tech. Officer,
PC Unit (S&N),
J.N. Krishi vishwa Vidyalaya
Jabalpur-482 004
Madhya Pradesh

ID No.1664

Dr. Raheja, R.K.
Sr. Scientist (Oilseeds),
Dept. of Plant Breeding,
Punjab Agril. University
Ludhiana-141 004
Punjab

ID No.1665

Dr. Rai Bhupendra
Professor,
Dept. of Genetics & Pl. Breeding,
Institute of Agril. Sciences,
Banaras Hindu University
Varanasi-221 005
Uttar Pradesh

ID No.1666

Dr. Raikhel Kar, S.V.
C/o Dr. V. Praveen Rao,
Assoc. Professor, ANGRAU,
Manohara Apartments,
Flat No. 211, Vidyanagar
Hyderabad-500 005
Andhra Pradesh

ID No.1667
Dr. Raj, S.K.
B-2/452,
Kalyani, Dt.
Nadia-741 235
West Bengal

ID No.1668
Dr. Raja Bhaskar
Jr. Entomologist
Seeds Science & Technology,
Tapiaca and Castor Research Station¹
Yethapur-636 119
Tamil Nadu

ID No.1669
Dr. Raja Reddy, K.
Professor (Pl. Breeding),
H.No. C-44(6),
Nadamuni Reddy Building,
S.V. Nagar, Post Office Road
Tirupati-517 502
Andhra Pradesh

ID No.1670
Dr. Rajagopal, V.
303, Pallavi Apartments
Irlangar
Tirupati-517 507
Andhra Pradesh

ID No.1671
Mr. Rajan P. Khedekar
JRA, Adarsha Colony,
Pragya Bungalow
Behind New Power House
Akola-444 104
Maharashtra

ID No.1672
Dr. Rajan, S.S.
Retd. Sr. Advisor to FAO,
Falt No. 21, Abhijit Apartments,
67, Seventh Main Road
Bengaluru-560 003
Karnataka

ID No.1673
Dr. Rajanna
Entomologist,, AICRP on White Grub.,
Department of Entomology.,
University of Agril. Sciences,
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1674
Dr. Rajanna, M.P.
Asst. Professor (Pl. Breeding),
Regional Research Station,
Navile, P.B. No. 126
Shimoga-577 231
Karnataka

ID No.1675
Mr. Rajasekhara Reddy, D.
S/o D. Bakki Reddy,
Syamala Nagar, 3rd Lane.
Near Balakatur School
Guntur-522 006
Andhra Pradesh

ID No.1676
Dr. Rajeev Shrivastava
Asst. Professor, Dept. of Biotechnology,
College of Agriculture,
Indira Gandhi Krishi Vishwavidyalaya
Raipur
Chhattisgarh

ID No.1677
Dr. Rajendra Choudhary
Principal Scientist,
Division of Environmental Sciences,
Indian Agril. Research Institute, Pusa²
New Delhi-110 012
New Delhi

ID No.1678
Dr. Rajendra Pras
Seed Research Officer,
AICRP on Seed Technology, UAS
NSP, GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1679
Dr. Rajendra Prasad
Pl. Pathologist (Oilseeds),
Oilseeds Section,
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1680
Dr. Rajendran, T.P.
Asst. Director General (PP),
Indian Council of Agril. Research,
Krishi Bhawan
New Delhi-110 001
New Delhi

ID No.1681
Dr. Rajesh S. Patil
Cotton Breeder,
Cotton Scheme, Agril. Research
Station,
Hebbal Farm, Hebballi
Dharwad-580 005
Karnataka

ID No.1682
Dr. Rajguru, A.B.
Asst. Professor (Physiology),
Oilseeds Research Scheme
(Sunflower),
Zonal Agril. Research Station
Solapur
Maharashtra

ID No.1683
Dr. Rajkumar Ramteke
Scientist (Genetics),
NRC for Soybean,
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.1684
Dr. Rajkumar, S.
Asst. Professor (Agronomy),
Seed Unit, University of , Agril.
Sciences,
Dharwad-580 005
Karnataka

ID No.1685
Dr. Rajnikant H. Kavani
Sr. Research Asst.,
Sai Shaba Society, Block No.25,
Rajni
Junagadh-362 002
Gujarat

ID No.1686
Dr. Rajpurohit, T.S.
Pathologist, Agril. Research Station,
Rajasthan Agril. University
Mandor-342 304
Rajasthan

ID No.1687
Dr. Rajput, A.S.
Scientist, AICRP (Rice),
Zonal Agril. Research Station,
Kumhrawand Farm
Jagdalpur-494 005
Chhattisgadh

ID No.1688
Dr. Rajput, J.C.
Director of Research,
Nirmal Seeds Pvt. Ltd.,
Bhadgaon Road, Anturli Phata
Jalgaon-404 201
Maharashtra

ID No.1689
Dr. Raju, S.G.
Plant Pathologist (SMS),
KVK, University of Agril. Sciences,
Krishinagar
Dharwad-580 008
Karnataka

ID No.1690
Dr. Ram Ashish Yadav
Jr. Agronomist (Sesame)
Oilseeds Section,
C.S. Azad Univ. of , Agril. &
Technology,
Nawabganj
Kanpur-208 002
Uttar Pradesh

ID No.1691
Dr. Ram Bhajan
Jr. Breeder (R&M),
Dept. of Plant Breeding,
NDUA&T, Narendranagar,
Kumarganj
Faizabad-224 229
Uttar Pradesh

ID No.1692
Dr. Ram Kishor Gupta
Sr. Scientist ,
CIPHET, Punjab Agril. University,
Ludhiana-141 001
Punjab

ID No.1693
Dr. Ram Shanker Singh
Jr. Agronomist,
Rapeseed - Mustard,
Tirhut College of Agriculture
Dholi-343 121
Bihar

ID No.1694
Dr. Ram Wamanrao Gawane
Jr. Entomologist,
College of Agriculture
Nagpur-440 001
Maharashtra

ID No.1695
Dr. Rama Subba Reddy, R.
Sr. Scientist (Pl. Breeding),
Head, Agril. Research Station
Tandur-501 141.
Andhra Pradesh

ID No.1696
Dr. Ramachandra, K.S.
155/A, 6th Cross,
II Block, Jayanagar
Bengaluru-560 011
Karnataka

ID No.1697
Dr. Ramachandram, M.
Ex-Principal Scientist
H.No. 1-8-21/1B, Municipal Colony
New Dilsukhnagar
Hyderabad-500 036
Andhra Pradesh

ID No.1698
Dr. Ramachandrappa, B.K.
~~Associate Director of Research,~~
Agril. Research Station,
University of Agril. Sciences
Hiriyur-572 143
Karnataka

ID No.1699
Dr. Ramana Rao, S.V.
~~26-75,~~
Chanakyapuri,
Safilguda
Secunderabad-500 047
Andhra Pradesh

ID No.1700
Dr. Ramana, J.V.
Asst. Professor,
Dept. of Plant Breeding & Genetics,
Agril. College
Bapatla-522 201
Andhra Pradesh

ID No.1701
Dr. Ramana, M.V.
Sr. Scientist (Pl. Breeding),
AICRP on Soybean,
Lam
Guntur-522 001
Andhra Pradesh

ID No.1702
Dr. Ramanath Rao, V.
C/o V. Padmaja Rao,
Head of the Division of **Physiology**,
IGFRI, Gwalior Road
Jhansi-284 003
Uttar Pradesh

ID No.1703
Dr. Ramanjaneyulu, A.V.
Jr. Agronomist, AICRP (Castor),
Regional Agril. Research Station,
Palem., Mahaboobnagar Dt.
Palem-509 215
Andhra Pradesh

ID No.1704
Dr. Ramanjaneyulu, G.V.
Executive Director,
Centre for Sustainable Agriculture,
12-13-445, Street No.1,
Tarnaka
Secunderabad-500 017
Andhra Pradesh

ID No.1705
Dr. Ramappa, H.K.
Jr. Pathologist,
AICRP on Sunflower,
University of Agril. Sciences,
GKVK
Bengaluru-560 065
Karnataka

ID No.1706
Dr. Ramashray Yadav
H.No. 103,
B rindavan Enclave,
Railway Niwas,
Opp: Railway Station, Bolaram
Secunderabad-500 014
Andhra Pradesh

ID No.1707
Dr. Ramegouda, G.K.
S/o Krishna Gouda,
A/P Mandya
Goravale-571 405
Karnataka

ID No.1708
Dr. Ramesh Babu, V.
Pl. Physiologist,
NARP, ANGRAU
Tirupati
Andhra Pradesh

ID No.1709
Dr. Ramesh Kumar, S.C.
Sr. Scientist (Ag. Econ.),
NBSSLUP Regional Centre,
Hebbal
Bengaluru-560 024
Karnataka

ID No.1710
Dr. Ramesh
Principal Scientist,
Directorate of Soybean Research
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.1711
Dr. Ramesh, T.
Asst. Professor,
Environmental Science & Technology,
College of Agriculture, ANGRAU
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1712
Dr. Ramprakash, T.
Scientist (Soil Science),
Regional Research Station,
Palem, Mahaboobnagar Dt.
Palem-509 215
Andhra Pradesh

ID No.1713
Dr. Rana, D.S.
Principal Scientist (Agronomy),
Division of Agronomy,
Indian Agril. Research Institute, Pusa
New Delhi-110 012
New Delhi

ID No.1714
Dr. Rana, J.S.
Professor,
Nanotechnology,
Guru Jainbeshwar University
Hisar-124 004
Haryana

ID No.1715
Dr. Ranbir Singh
Principal Scientist,
National Bureau of Plant Genetic
Resources
Pusa Campus
New Delhi-110 012
New Delhi

ID No.1716
Dr. Ranga Rao, V.
Plot No. 128-A, MLA Colony,
Road No. 12 (Old), Avenue No.1,
Street No. 12 (New)
Banjara Hills
Hyderabad-500 003
Andhra Pradesh

ID No.1717
Dr. Ranga Reddy, R.
H.No. 2-2-1147/1,
Sri Sai Saketh Apartments,
New Nallakunta
Hyderabad-500 044
Andhra Pradesh

ID No.1718
Dr. Ranganatha, A.R.G.
Project Coordinator (S&N)
PC Unit (Sesame & Niger)
Jawaharlal Nehru Krishi Viswavidyalaya
Jabalpur-482 004
Madhya Pradesh

ID No.1719
Dr. Rao, D.S.
Asst. Professor,
Agril. Research Station, (
Rajasthan Agril. University
Mandor
Jodhpur-342 304
Rajasthan

ID No.1720
Dr. Rao, J.V.
Principal Scientist, (Retd.)
CRIDA, Santoshnagar
Saidabad
Hyderabad
Andhra Pradesh

ID No.1721
Dr. Rao, M.V.H.
Associate Professor,
P.B. No 24,
University of Agril. Sciences
Raichur-584 101
Karnataka

ID No.1722
Dr. Rao, S.S.
Scientist (Oilseeds),
Indira Gandhi Agril. University,
Krishi Nagar, P.B. No. 94
Raipur-492 012
Chhattisgarh

ID No.1723
Dr. Rao, S.V.
Scientist, AICRP on Pigeonpea,
Agril. Research Station,
Ratanpur, Post: Krupasindhupur,
Via: Nimakhandi, Berhampur
Gajam-761 001
Orissa

ID No.1724
Dr. Rao, V.S.N.
Sr. Scientist (Pl. Breeding),
134, Keshanpura,
ITIRoad
Hoshangabad
Madhya Pradesh

ID No.1725
Dr. Rao, Y.R.
401, Sai Vipula Apartments,
6-1-192/A/2, Padmarao Nagar
Secunderabad-500 025
Andhra Pradesh

ID No.1726
Dr. Raoof, M.A.
Principal Scientist ,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1727
Dr. Rathnaparkí, R.D.
Sr. Research Asst.,
Oilseeds Unit,
Dr. Punjabrao Deshmukh Krishi
Vidyapeeth
Akola-444 104
Maharashtra

ID No.1728
Dr. Rathod Darasing Ramsingh
Research Associate,
Nimbkar Agril. Research Institute,
P.Bo. No.44
Phaltan-415 523
Maharashtra

ID No.1729
Dr. Rathore, M.S.
Asst. Professor,
Agril. Research Station
Mandor
Jodhpur-342 304
Rajasthan

ID No.1730
Mr. Ratna Sree, R.
SRF,
Directorate of Oilseeds, Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1731
Dr. Ratnakumar, A.L.
Sr. Scientist (Pl. Breeding),
Directorate of Groundnut Research
Ivnagar Road, P.B. No.5
Junagadh-362 001
Gujarat

ID No.1732
Dr. Raut Vinod Kishan Rao
Asst. Research Scientist,
Plot 463/B-4, Mauli Nagar,
Datta Nagar, Ambad Road
Jalna-431 203
Maharashtra

ID No.1733
Dr. Raval, C.M.
Associate Research Scientist,
Gujarat Agril. University
Junagadh-362 001
Gujarat

ID No.1734
Dr. Raveendran, T.S.
Director,
Centre for Plant Breeding & Genetics,
Tamil Nadu Agricultural University
Coimbatore-641 003
Tamil Nadu

ID No.1735
Dr. Ravi Hunje
Associate Professor,
Seed Science & Technology Unit,
University of Agril. Sciences,
Krishinagar
Dharwad-580 005
Karnataka

ID No.1736
Mr. Ravi Kanta
121,
Mohalla Chaudharian
Hisar-124 004
Haryana

ID No.1737
Dr. Ravi Kishan, P.
B-50/F-6,
Vijayanagar Colony
Hyderabad-500 457
Andhra Pradesh

ID No.1738
Dr. Ravi Kumar, G.H.
Sr. Farm Superintendent, College of
Agriculture, Navile, P.B. No. 126,
Shimoga-577, 204
Shimoga-577 204

ID No.1739
Dr. Ravi Kumar, R.L.
Dept. of Plant Breeding & Genetics,
University of Agril. Sciences,
Krishinagar
Dharwad-580 005
Karnataka

ID No.1740
Dr. Ravi, G.M.
207, Orange Block,
My Home Rainbow Complex,
Tolichowki
Hyderabad-500 008
Andhra Pradesh

ID No.1741
Dr. Ravichandran, V.K.
Professor & Head,
Oilseeds Research Station,
Tamil Nadu Agril. University
Tindivanam-604 001
Tamil Nadu

ID No.1742
Dr. Ravindra Babu, Y.
Associate Reserach Scientist,
AICRP on Under Utilized Crops,
RAS, S.D. Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1743
Dr. Ravindra, V.
Scientist (Pl. Physiology),
Indian Institute of Horticultural
Research,
Hasargattlake Post
Bengaluru
Karnataka

ID No.1744
Dr. Ravindrachary, G.
Sr. Scientist,
Central Research Institute for, Dryland
Agriculture,
Santoshnagar, Saidabad
Hyderabad-500 659
Andhra Pradesh

ID No.1745
Dr. Ravishankar, G.
Jr. Agronomist,
Salinity Scheme,
Agril. Research Station
Gangavati-583 227
Karnataka

ID No.1746
Dr. Reddy Sekhar, M.
Asst. Professor,
Dept. of Genetics & Pl. Breeding,
S.V. Agril. College
Tirupati-517 502
Andhra Pradesh

ID No.1747
Dr. Reddy, D.V.R.
Principal Virologist,
ICRISAT
Patancheru-502 324
Andhra Pradesh

ID No.1748
Dr. Reddy, K.R.K.
Chief Executive VGBF,
302, Manas, 5-36/36,
Prasanthnagar, Kukatpally
Hyderabad-500 037
Andhra Pradesh

ID No.1749
Dr. Reddy, L.J.
Groundnut Breeder,
ICRISAT
Patancheru-502 324
Andhra Pradesh

D No.1750
Jr. Reddy, P.S.
Flat No. 201,
Block "C", Amruta Enclave,
Road No. 14, Banjara Hills
Hyderabad-500 034
Andhra Pradesh

ID No.1751
Mr. Reddy, Y.R.G.
Technical Officer,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1752
Dr. Rekha Mittal
Scientist,
Indian National Scientist Doc. Centre
New Delhi-110 067
New Delhi

ID No.1753
Dr. Renukadevi, P.
Jr. Pathologist,
Oilseeds Research Station,
Tamil Nadu Agril. University
Tindivanam-604 001
Tamil Nadu

ID No.1754
Dr. Riazuddin Ahmed, S.
Sr. Scientist (Soil Science),
STCR, Agril. Research Institute,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1755
Dr. Rinku Dey
Scientist,
Directorate of Groundnut Research
Innagar Road, P.B. No.5
Junagadh-362 001
Gujarat

ID No.1756
Dr. Rudra Naik, V.
Associate Professor of Plant Breeding,
University of Agril. Sciences,
Krishinagar
Dharwad-580 005
Karnataka

ID No.1757
Dr. Rudra Samy, P.
Sr. Scientist (Breeder),
AICRP on Sunflower,
University of Agril. Science
BKVK Campus
Bengaluru-560 065
Karnataka

ID No.1758
Dr. Ruopeiiselhou Kehie
BPO-Lalmaji,
SMS (Ent.),
Krishi Vigyan Kendra,
Peducha
Kohima-979 001
Nagaland

ID No.1759
Dr. Rustum Kishan Rao, B.
Safflower Breeder,
C/o Dr. R.K. Bhalerao,
A-12, Sector-3, MAU Quarters
Parbhani-431 402
Maharashtra

ID No.1760
Dr. Ubale. S.S.

ID No.1761
Dr. Sah, J.N.
Sr. Sunflower Breeder,
AICRP on Sunflower,
Tirhut College of Agriculture,
Rajendra Agril. University
Dholi-843 121
Bihar

ID No.1762
Dr. Saha, P.K.
Agronomist,
Dept. of Agriculture,
Pulses and Oilseeds Research Station
Berhampore-742 101
West Bengal

ID No.1763
Dr. Sahadeva Reddy, B.
Sr. Scientist (Agronomy),
Agril. Research Station,
DCMS Buildings, Kamala Nagar
Anantapur-515 001
Andhra Pradesh

ID No.1764
Mr. Sai Kumar, K.
S/o K. Ananta Ramulu,
H.No. 1-6-141/29/3/A/3,
Balaji Nagar, Suryapet, Nalgonda Dt.
Suryapet-508 213
Andhra Pradesh

ID No.1765
Dr. Sailaja, M.
Research Associate,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1766
Dr. Sakale, B.K.
Sr. Research Fellow (ROPS-1),
College of Food Technology,
Marathwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1767
Dr. Salaimalai, A.
6/116, Melavaragunatama puram,
Rathur (Post), Muhavur
Rajapalayam-626 111
Tamil Nadu

ID No.1768
Dr. Salimath, P.M.
No. 39, Jayanagar
(Vidyanagar)
Hubli-580 021
Karnataka

ID No.1769
Dr. Samal, P.
Principal Scientist (Ag. Economics),
Central Rice Research Institute
Cuttack
Orissa

ID No.1770
Dr. Samanta, S.K.
Jr. Plant Breeder,
Q.No. B-14/170
Kalyani-741 235
West Bengal

ID No.1771
Dr. Samdur, M.Y.
Scientist (Plant Breeding),
Directorate of Groundnut Research
P.B.No.5, Ivnagar Road
Junagadh-362 001
Gujarat

ID No.1772
Dr. Sameer Padmakar Mulay
Executive Director,
Ajeet Seeds Ltd., Cint No. 233,
Vill: Chitagaon, Tq: Fathan
Aurangabad-431 105
Maharashtra

ID No.1773
Dr. Sampathkumar
Sr. Scientist,,
Agril. Research Station,,
Kadiri, Anantapur Dist.
Kadiri-515 591
Andhra Pradesh

ID No.1774
Dr. Samui, R.C.
Professor in Agronomy,
B-6/57, Post: Kalyani
Nadia-741 235
West Bengal

ID No.1775
Dr. Sandeep Bhandarkar
Scientist (Plant Breeding),
S.G. College of Agril. & **Research**
Station
Jagdapur-494 005
Chhattisgadh

ID No.1776
Dr. Sandhyarani S. Nirhari
Research Associate,
Seed Unit,
University of Agricultural Sciences
Dharwad-580 005
Karnataka

ID No.1777
Dr. Sanjay Kumar Dwivedi
Scientist, Dept. of Agronomy,
College of Agriculture,
Indira Gandhi Krishi Viswa Vidyalaya
Raipur

ID No.1778
Dr. Sanjeev Kumar Jha
Scientist,
NRC for Rapeseed-Mustard
Post: Sewar
Bharatpur-321 303
Rajasthan

ID No.1779
Dr. Sanjeev Kumar
Sr. Scientist (Agronomy),
ICAR Res. Complex for NEH Region,
Walmi Complex, Phulwari Sharit
Patna-801 505
Bihar

ID No.1780
Dr. Sanjiv S. Parhatay
Jr. Pathologist,
College of Agriculture
Nagpur-440 001
Maharashtra

ID No.1781
Dr. Santha Laxmi Prasad
Scientist (SS),
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1782
Dr. Santosh Kumar
Scientist (Pl. Breeding),
S.G. College of Agriculture & **Research**,
I.G. Krishi Vishwa Vidyalaya
Jagdapur-494 005

ID No.1783
Dr. Sarada, Ch.
Sr. Scientist,
Directorate of Oilseeds **Research**,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1784
Dr. Sarala, B.S.
SRF, Regional Agril. **Research Station**,
University of Agril. **Sciences**
Raichur-584 101
Andhra Pradesh

ID No.1785
Dr. Saralamma, S.
H.No. 26/338,
Maddi Sudarsan Nagar, **R.S. Road**
Nandyal-518 503
Andhra Pradesh

ID No.1786
Dr. Sarbjeet Kaur
Asst. Pl. Pathologist,
Oilseeds Section,
Dept. of Pl. Breeding,
Punjab Agril. University
Ludhiana-141 004
Punjab

ID No.1787
Dr. Sarode, S.V.
Director of Research,
Dr. P.D. Krishi Vishwa Vidyalaya
Akola-444 104
Maharashtra

ID No.1788
Dr. Sarojini, G.
Associate Professor,
Dept. of Food & Nutrition,
ANGRAU, Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1789
Dr. Sarvendra Kumar
Breeder (Linseed),
C.S. Azad University of Agril. & **Te^{ch}**.
Kanpur-208 002
Uttar Pradesh

ID No.1790
Dr. Sarwan Kumar
Asst. Entomologist,
Oilseeds Section,
Dept. of Pl. Breeding & Genetics,
Punjab Agril. University
Ludhiana-141 004
Punjab

ID No.1791
Dr. Sasidharan, N.
Associate Professor,
Dept. of Agril. Botany & **Biotechnology**,
BA College of Agriculture,
Anand Agril. University
Anand-388 110
Gujarat

ID No.1792
Mr. Sastry, J.A.
H.No. 23/B, Bansilalpet,
Beside Bible House,
R.P. Road
Secunderabad-500 003
Andhra Pradesh

ID No.1793
Dr. Sastry, K.S.
Sr. Scientist (Retd.),
NRCS,
Dilsukhnagar
Hyderabad-500 036
Andhra Pradesh

ID No.1796
Dr. Satpathy, P.C.
Sesame Breeder,
Dept. of Pl. Breeding,
Orissa University of Agril. & Tech.
Bhubaneshwar-751 003
Orissa

ID No.1799
Dr. Satya Prasad, M.
Scientist (Biotechnology), **1-10-177**,
4th Floor, Varun Towers,
Begumpet
Hyderabad-500 016
Andhra Pradesh

ID No.1802
Dr. Satyanarayana, V.
Assoc. Professor (Agronomy),
H.No. 3-8-132/1, Plot No. 61, **Road**
No.4,
Chandranagar, L.B. Nagar
Hyderabad-500 074
Andhra Pradesh

ID No.1805
Dr. Savasaiya, C.D.
Agril. Supervisor,
Agril. Research Station,
Gujarat Agril. University
Amreli-365 601
Gujarat

ID No.1808
Dr. Seetharam, A.
PC, Emeritus Scientist (Millets),
University of Agril. Sciences,
GKVK Campus
Bengaluru-560 065
Karnataka

ID No.1794
Dr. Satish Kumar, G.D.
Scientist,
Directorate of Groundnut **Research**
P.B.No.5, Ivnagar Road
Junagadh
Gujarat

ID No.1797
Dr. Satpute, G.N.
Jr. Breeder,
Crop Research Unit (Oilseeds),
Dr. PDKV, Near Ram Mandir
Akola-444 104
Maharashtra

ID No.1800
Mr. Satyanarayana Rao, V.
Scientist (Breeding) & Head,
AICRP on Forage Crops,
Live Stock Research Institute,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1803
Dr. Satyanshu Kumar
Scientist,
Directorate of Rapeseed-**Mustard**
Research
Sewar Farm
Bharatpur-321 001
Rajasthan

ID No.1806
Dr. Sawant, R.H.
Jr. Research Assistant,
O/o ADR, ZARS, 97,
Raviwarpet
Solapur-413 003
Maharashtra

ID No.1809
Dr. Sekhar, I.
Associate Professor (Forestry),
Forestry College & Research **Institute**
Mettupalayam-641 003
Tamil Nadu

ID No.1795
Dr. Satishchandra Chandulal Sawa
Sr. Scientist,
A-2, Samar,
MAHYCo Colony
Jalna-431 203
Maharashtra

ID No.1798
Dr. Satwadhar, P.N.
Associate Professor,
Faculty of Food Technology,
Marathwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1801
Dr. Satyanarayana, J.
Associate Professor,
Dept. of Entomology,
College of Agriculture, ANGRAU,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1804
Dr. Savalia, R.L.
Research Scientist (Pl. Path),
Pushparaj, Shridhar Nagar,
Behind Lal Bang, Shashikunj Road
Junagadh-362 001
Gujarat

ID No.1807
Dr. Sawarkar, S.D.
Soil Scientist,
Zonal Agril. Research **Station**
J.N. Krishi Vishwa **Vidyalyaya**
Chindwara
Madhya Pradesh

ID No.1810
Dr. Sekhon, B.S.
Entomologist, Regional Research
Station,
Punjab Agril. University
Bhatinda-151 001
Punjab

ID No.1811
Dr. Selvanarayanan, V.
Reader in Entomology,
17, Thyagn Periyar Street
Chidambaram-608 001
Tamil Nadu

ID No.1812
Dr. Sessa Saila Sree, P.
D/o Dr. D. Lakshman Das,
H.No. H2/75, Gowli Street
Kurnool-518 001
Andhra Pradesh

ID No.1813
Mr. Seshadri Reddy S.
Post Doctoral Research Associate,,
H.No.5-3-319/5
Ventaka Rao Nagar Colony
Kukatpally
Hyderabad-500 072
Andhra Pradesh

ID No.1814
Dr. Shadakshari, Y.G.
Geneticist, AICRP (Sunflower),
University of Agril. Sciences,
GKVK campus
Bengaluru-560 065
Karnataka

ID No.1815
Dr. Shaik Ameen Basha
Jr. Pathologist,
Regional Agril. Research Station,
Palem, Mahaboobnagar Dt.
Palem-509 215
Andhra Pradesh

ID No.1816
Dr. Shaik Mohammed
H.No. 18-7-198-A-285,
Moghalpur Road
Hyderabad-500 002
Andhra Pradesh

ID No.1817
Shakti Vardhak Hybrid Seeds P. Ltd.
Tilak Bazar
Hisar-125 001
Haryana

ID No.1818
Dr. Shalini D. Sagar
Jr. Pathologist,
AICRP (Sunflower),
Regional Agril. Research Station
Raichur-584 101
Karnataka

ID No.1819
Dr. Shambuling C. Shahapur
Dy. Manager
Vibha Agrotech Ltd., Plot No.21
Sector-I, HUDA Techno Enclave
Hightech City Road

ID No.1820
Dr. Shankar Goud, I.
Sunflower Breeder,
Regional Research Station,
University of Agril. Sciences,
P.B. No. 24
Raichur-584 101
Karnataka

ID No.1821
Dr. Shanker, M.A.
Chief Scientist,
AICRP for Dryland Agriculture,
University of Agril. Sciences,
GKVK
Bengaluru-560 065
Karnataka

ID No.1822
Dr. Shanta Hiremath
Breeder, Elite Sunflower,
Seed Production Scheme,
University of Agril. Sciences,
GKVK
Bengaluru-560 065
Karnataka

ID No.1823
Dr. Shanthala, J.
Asst. Seed Research Officer,
No. 59, A.G. Colony,
Anandnagar Extension, Hebbal
Bengaluru-560 024
Karnataka

ID No.1824
Dr. Sharma, A.K.
Documentation Officer,
Indain Institute of Soil Science,
Nabi Bagh, Berasaia Road
Bhopal-462 038
Madhya Pradesh

ID No.1825
Dr. Sharma, A.N.
Sr. Scientist (Entomology),
Directorate of Soybean Research
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.1826
Mr. Sharma, D.J.
Scientist,,
Indiragandhi Krishi Vidyalaya,,
CARS
Bilaspur-495 001
Chhatisgadh

ID No.1827
Dr. Sharma, D.K.
Principal Scientist,
Indira Gandhi Krishi Viswa Vidyalaya
Raipur-492 006
Chhatisgadh

ID No.1828
Dr. Sharma, K.N.
Agricultural Research Station
Durgapura-302 018
Rajasthan

ID No.1829
Dr. Sharma, P.B.
Jr. Scientist (Agronomy),
Zonal Agril. Research Station
J.N. Krishi Vishwa Vidyalaya
Hoshangabad-461 110
Madhya Pradesh

ID No.1830
Dr. Sharma, S.M.
Ex-Project Coordinator *S&N)
53, Alok Nagar, Post Adarthal
Jabalpur-482 004
Madhya Pradesh

ID No.1831
Dr. Sharma, T.V.R.S.
Head, Division of Field Crops,
Central Agril. Research Institute,
101 P.B.No. 181
Portblair-744 181

ID No.1832
Dr. Shashi Banga
Sr. Breeder (Oilseeds),
Oilseeds Unit, Dept. of Pl. , Breeding &
Genetics,
Punjab Agricultural University
Ludhiana-141 004
Punjab

ID No.1833
Dr. Shashikant Sadashiv Nahire
Sr. Pathologist,
Syngenta India Ltd.,
1170/27, Revenue Colony, Shivajinagar
Pune-411 005
Maharashtra

ID No.1834
Dr. Sheoran, R.K.
Sunflower Breeder, Oilseeds Section,
CCS Haryana Agril. University
Hisar-125 004
Haryana

ID No.1835
Dr. Sheshidhar Viraktmath
Associate Professor,
Dept. of Entomology,
University of Agril. Sciences
Dharwad-580 005
Karnataka

ID No.1836
Dr. Shete
Seed Research Officer,
Seed Technology, Research Unit,
Marathwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1837
Dr. Shinde Shahaji Kakasaheb
Asst. Professor of Botany,
97, Raviwar Pet, P.B. No.207,
Zonal Agril. Research Station
Solapur-413 002
Maharashtra

ID No.1838
Dr. Shinde, Y.M.
Head, Dept. of Botany,
Mahatma Phule Krishi Vidyapeeth,
Dt. Ahmednagar
Rahuri-413 722
Maharashtra

ID No.1839
Dr. Shirshikar, S.P.
Pathologist (Sunflower),
Oilseeds Research Station
Latur-413 512
Maharashtra

ID No.1840
Dr. Shiva Kumar, B.G.
Sr. Scientist,
Division of Agronomy,
Indian Agril. Research Institute
Pusa
New Delhi-110 012
New Delhi

ID No.1841
Dr. Shivaji Pandurang Mehtre
Asst. Professor of Botany,
Tissue Culture Lab,
Marathwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1842
Dr. Shivam, D.
Scientist (Breeding),
Agril. Research Station,
ANG Ranga Agril. University
Tandur-501 141
Andhra Pradesh

ID No.1843
Dr. Shivshanker Dash
C/o Dr. R.K. Akharui,
Oilseeds Coordinator (Sunflower),
Rajendra Agril. University,
Tirhut Agril. College
Dholi
Bihar

ID No.1844
Dr. Shukla, A.K.
Sr. Scientist (Pl. Pathology),
Directorate of Rapeseed-Mustard
Research
Sewar Farm, P.B. No. 41
Bharatpur-341 001
Rajasthan

ID No.1845
Dr. Shyam Prakash
Sr. Scientist,
Directorate of Research on Plant
Biotechnology
IARI Campus, Pusa
New Delhi-110 012
New Delhi

ID No.1846
Dr. Singh Bhupan
Assoc., Professor,
Dept. of Agronomy,
Tirhut College of Agriculture
Dholi-843 121
Bihar

ID No.1847
Dr. Singh Lallu
Jr. Oil Chemist (Sunflower),
C.S. Azad University of Agril. & Tech
Kanpur-208 002
Uttar Pradesh

ID No.1848
Dr. Singh, A.L.
Plant Physiologist,
Directorate of Groundnut Research
P.B. No.5, Ivnagar Road
Junagadh-362 001
Gujarat

ID No.1849
Dr. Singh, B.R.
Breeder (S&N),
PC Unit (S&N),
J.N. Krishi Vishwa Vidyalaya
Jabalpur-482 004
Madhya Pradesh

ID No.1850
Dr. Singh, C.P.
Principal Scientist (Agronomy),
Crop Science Division,
Directorate of Agril. Research
Krishi Bhawan
New Delhi-110 001
New Delhi

ID No.1851
Dr. Singh, C.P.
Associate Professor (Ent.),
G.B. Pant University of Agril. & Tech
Pantnagar-263 145
Uttar Pradesh

ID No.1852
Dr. Singh, D.V.
Sr. Scientist (Farm Machinery),
Directorate of Soybean Research,
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.1853
Dr. Singh, J.P.
Directorate of Oilseeds Development,
Telhan Bhawan, Himayatnagar
Hyderabad-500 029
Andhra Pradesh

ID No.1854
Dr. Singh, K.D.
Director (Retd.),
Central Research Institute for Tobacco
Rajahmundry
Andhra Pradesh

ID No.1855
Mr. Singh, M.K.
Sr. Scientist (Oilseeds),
Dept. of Plant Breeding & Genetics,
Indira Gandhi Krishi Viswa Vidyalaya
Raipur-492 006
Chhattisgarh

ID No.1856
Dr. Singh, M.V.
Project Coordinator,
AICRP on Micro Nutrients,
Indian Institute of Soil Science,
Nabibagh, Berasia Road
Bhopal-462 038
Madhya Pradesh

ID No.1857
Dr. Singh, N.B.
Agriculture Commissioner,
Govt. of India,
Krishi Bhavan
New Delhi-110 001
New Delhi

ID No.1858
Dr. Singh, N.P.
ICAR Research Complex for NEH
Region,
Tripura Centre
Lembucherra-799 210
Tripura

ID No.1859
Dr. Singh, O.P.
Jr. Physiologist,
AICRP on Groundnut,
C.S. Azad University of Agril. & Tech.,
Research Station
Manipur
Uttar Pradesh

ID No.1860
Dr. Singh, P.K.
Jr. Breeder (Linseed),
AICRP (Linseed),
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1861
Dr. Singh, R.B.
Sr. Scientist (Plant Path.),
Dept of Plant Breeding,
N.D. University of Agril. & Tech.,
Kumarganj
Faizabad-224 229
Uttar Pradesh

ID No.1862
Dr. Singh, R.K.
C/o Harish Gupta,
G-159, Pushkar Enclave,
Paschim Vihar
New Delhi-110 063
New Delhi

ID No.1863
Dr. Singh, R.P.
Director (Retd.),
61, Sardar Club Scheme
Jodhpur-342 001
Rajasthan

ID No.1864
Dr. Singh, R.P.
Flat No. 502, Block 9,
Hill Ridge Spring, IVRCL
Gachibowli
Hyderabad-500 032
Andhra Pradesh

ID No.1865
Dr. Singh, S.N.
Sr. Pl. Pathologist (Oilseeds),
Tirhut College of Agriculture,
Dholi
Muzaffarpur-843 121
Bihar

ID No.1866
Dr. Singh, S.S.
Sr. Scientist (Agronomy),
ICAR Research **Complex for Castor**
Region,
Phulwarisharif
Patna-801 505
Bihar

ID No.1867
Dr. Singh, T.V.K.
Professor, ANGRAU,
23-5-747,
Laldarwaza
Hyderabad-500 065
Andhra Pradesh

ID No.1868
Dr. Singh, V.P.
Sr. Scientist (P&O),
CCS Haryana Agril. University,
Regional Research Station
Bawal
Haryana

ID No.1869
Dr. Singh, Y.P.
Sr. Scientist (Entomology),
Directorate of Rapeseed-Mustard
Research
Sewar Farm
Bharatpur-321 303
Rajasthan

ID No.1870
Dr. Sitaram Kumhar
Asst. Professor,
Agril. Research Station,
Rajasthan Agril. University
Mandor
Rajasthan

ID No.1871
Dr. Siva Prasad, V.
Asst. Manager,
J.K. Agri Genetics,
1-10-177, 4th Floor,
Varun Towers, Begumpet
Hyderabad-500 016
Andhra Pradesh

ID No.1872
Dr. Siva Reddy, V.
H.No. 11-156,
S.V. Nagar
Tirupati-517 502
Andhra Pradesh

ID No.1873
Dr. Sivaraj Natarajan
Scientist,
NBPGR Regional Research Station,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1874
Dr. Snehalatha Reddy, N.
Professor (Food & Nutrition),
Home Science College,
Marathwada Agril. University
Parbhani-431 402
Maharashtra

ID No.1875
Dr. Sohan Ram
Niger Breeder,
Dept. of Pl. Breeding & **Genetics**,
Birsa Agril. University
Ranchi-834 006
Jharkhand

ID No.1876
Dr. Sokka Reddy, S.
Associate Professor,
Dept. of Genetics & Pl. Breeding,
College of Agriculture, ANGRAU,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1877
Dr. Solanki, R.M.
Agril. Officer,
Giriraj Part-2, Block-70,
Near Rajnandan Apartments,
Deepanjali-2, Timbawadi
Junagadh-362 015
Gujarat

ID No.1878
Dr. Solanki, S.S.
Associate Professor,
Agril. Research Station,
Rajasthan Agril. University, **Mandor**
Jodhpur-342 304
Rajasthan

ID No.1879
Dr. Solanki, Z.S.
Professor,
Agril. Research Station,
Rajasthan Agril. University, **Mandor**
Jodhpur-342 304
Rajasthan

ID No.1880
Mr. Soma Sekhar Reddy, P.
Sr. Research Fellow,
S/o P. Chenna Reddy,
H.No. 1-10-85/1, Shashabgutta

ID No.1881
Dr. Somanagouda, G.
Jr. Agronomist (Safflower),
Agril. Research Station
Annigeri-582 201
Karnataka

ID No.1882
Dr. Somasekhar
Entomologist (Groundnut),
AICRP on Groundnut,
Regional Research Station, UAS
Raichur-584 101
Karnataka

ID No.1883
Dr. Somashekhar
C/o G.S.Akki,,
1st Main,
Kumareswar Nagar
Dharwad-580 007
Karnataka

ID No.1886
Dr. Sri Devi, S.
Plot No. 781,
Prasanthnagar,
Vanasthalipuram
Hyderabad-500 070
Andhra Pradesh

ID No.1889
Dr. Sridevi, O.
Asst. Professor,
College of Agriculture,
University of Agril. Sciences
Dharwad-580 005
Karnataka

ID No.1892
Dr. Srikant Kulkarni
Prof. & Head,
Dept. of Pl. Pathology,
University of Agril. Sciences
Dharwad-580 005
Karnataka

ID No.1895
Dr. Srinivas, T.
Scientist (Crop Protection),
DAATT Centre,
Gudimalkapur Market Yard,
Near Mehdiapatnam
Hyderabad
Andhra Pradesh

ID No.1898
Dr. Srinivasulu, K.
SRF,
Directorate of Oilseeds **Research**
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1884
Mr. Sonnad, J.S.
Asst. Professor,
Dept. of Ag. Mktg. Coop. & Agri-
Business Mng., University of Agril.
Sciences,
Krishi Nagar
Dharwad-580 005
Karnataka

ID No.1887
Mr. Sri Hari Reddy, N.
Plant Breeder,
D.No. 201,
Saraswathi House, Saraswathi Colony,
Bapujinagar, Nacharam
Hyderabad-500 076
Andhra Pradesh

ID No.1890
Dr. Sridhar, P.
Asst. Professor,
Tamil Nadu Agril. **University**,
Tapioca & Castor **Research Station**
Yethapur-636 119
Tamil Nadu

ID No.1893
Mr. Srikant, V.
Jr. Research Fellow,
Directorate of Oilseeds **Research**,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1896
Dr. Srinivasa Reddy, M.
H.No. 29-178-10,
SBI Colony
Nandyal-518 501
Andhra Pradesh

ID No.1899
Dr. Srivastava, A.N.
117-1/182,
Naveen Nagar,
Kanpur-208 025
Uttar Pradesh

ID No.1885
Dr. Sreedhar, N.
Asst. Professor,
Dept. of Pl. Breeding & Genetics,
College of Agriculture, ANGRAU,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1888
Mr. Suryavanshi, S.B.
Jr. Research Asst. (*Agronomy*),
Oilseeds Research Station,
Marathwada Agril. University
Latur-413 512
Maharashtra

ID No.1891
Dr. Sridhar, V.
Associate Professor (*Agronomy*),
S.V. Agriculture College,
ANGRAU
Tirupati-517 502
Andhra Pradesh

ID No.1894
Dr. Srinivas, M.
Scientist (*Agronomy*),
Agril. Research Station
Anantapur-525 001
Andhra Pradesh

ID No.1897
Dr. Srinivasan, K.
Professor (Agronomy),
Dept. of Oilseeds,
Tamil Nadu Agril. University
Coimbatore-641 003
Tamil Nadu

ID No.1900
Dr. Srivastava, M.P.
Reader,
P.G. Dept. of Ag. Botany,
S.M.M. Town,
PG College
Ballia-277 001
Uttar Pradesh

ID No.1901
Dr. Srivastava, R.K.
Jr. Breeder, (Linseed),
Dept. of Pl. Breeding & Genetics,
N.D. University of Agril. & Tech.,
Narendranagar, Kumarganj
Faizabad-224 229
Uttar Pradesh

ID No.1902
Dr. Srivastava, R.K.
Dept. of Genetics & Pl. Breeding,
ND University of Agril. & Tech.
Kumarganj
Faizabad-224 229
Uttar Pradesh

ID No.1903
Dr. Srivastava, R.L.
Project Coordinator (Linseed),
C.S. Azad University of Agril. & Tech.
Kanpur-208 003
Uttar Pradesh

ID No.1904
Dr. Srivastava, S.C.
Chief Scientist,
Oilseeds Improvement Project,
Rajendra Agril. University
Pusa-848 125
Bihar

ID No.1905
Dr. Subash Chandra Mohapatra
Programme Coordinator,
Krishi Vidyan Kendra,
Orissa University of Agril. & Tech.,
Sonepur
Subampur-767 017
Orissa

ID No.1906
Dr. Subba Reddy, G.
Pr. Scientist & Head (Crop Science),
CRIDA, Santoshnagar, (AP) Saidabad
Hyderabad-500 659
Andhra Pradesh

ID No.1907
Dr. Subbakrishna, M.L.
Zuari Seeds Ltd.,
805, 13th Main Road,
Yelahanka New Town
Bengaluru-560 064
Karnataka

ID No.1908
Dr. Subbalakshmi, B.
Geneticist (Sunflower),
D.No.13A, St. No. 8, (TN)
P.N. Pudur
Coimbatore-641 041
Tamil Nadu

ID No.1909
Dr. Subbarayudu, B.C.
Breeder (Oilseeds),
Agril. Research Station
Darsi-523 247
Andhra Pradesh

ID No.1910
Dr. Subbarayudu, V.C.
Associate Professor,
Regional Agril. Research Station,
Nandyal, Kurnool D
Nandyal-518 503
Andhra Pradesh

ID No.1911
Dr. Subramanian, A.N.
Secretary,
IOPEA, 78/79,
Bajaj Bhawan, Nariman
Mumbai-400 021
Maharashtra

ID No.1912
Dr. Sudha Rani
Scientist (Agronomy),
AICRP on Sorghum,
Agril. Research Station
Tandur-501 141
Andhra Pradesh

ID No.1913
Dr. Sudha, B.G.
Scientist (Pl. Phy.),
Indian Institute of Horticulture
Research,
Hessargatta
Bengaluru-560 089
Karnataka

ID No.1914
Mr. Sudha, T.
Asst. Professor (Agronomy),
University of Agril. Sciences,
Dharwad-580 005
Karnataka

ID No.1915
Dr. Sudhakar Babu, S.N.
Sr. Scientist (Agronomy),
Directorate of Oilseeds Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1916
Dr. Sudhakar Nagrao Kulkarni
Associate Professor (Ent.),
488, Swasti Shree,
Ramkrishnagar
Parbhani-431 401
Maharashtra

ID No.1917
Mr. Sudhakar, C.
Jr. Agronomist,
AICRP on Safflower,
Agril. Research Station
Tandur-501 141
Andhra Pradesh

ID No.1918
Dr. Sudhakar, R.
Sr. Scientist (Pl. Path),
Biotechnology Centre,
Agril. Research Institute
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1919
Dr. Sudheer Kumar, S.
Asst. Professor,
27-B, Vengalrao Nagar Colony
Hyderabad-500 840
Andhra Pradesh

ID No.1920
Dr. Suganthy, M.
Asst. Professor, (Entomology),
Dept. of Oilseeds,,
Tamilnadu Agril. University
Coimbatore-641 003
Tamil Nadu

ID No.1921
Dr. Suganya
Scientist (Econ. Botany),
Div of Crop Improvement,
Sugarcane Breeding Institute
Coimbatore-641 003
Tamil Nadu

ID No.1922
Dr. Sugunakara Rao. B.
Scientist (Retd.),
1-2-56/56, Domalguda,
Advocates Colony
Hyderabad-500 029
Andhra Pradesh

ID No.1923
Suhas S. Lande, Dr.
Sr. Research Asst.,
A-1, Athvan Apartments,
Besides Seeta Geeta Apartments,
Sudhir Colony
Akola
Maharashtra

ID No.1924
Dr. Sujatha, M.
Principal Scientist,
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1925
Dr. Sujatha, M.
Scientist (Pl. Breeding),
Regional Agril. Research Station,
Palem, Mahaboobnagar Dt.
Palem-509 215
Andhra Pradesh

ID No.1926
Dr. Suma Biradar
Breeder (Safflower),
IX Cross Kusum Nagar,
Opp: BAIF
Dharwad-580 008
Karnataka

ID No.1927
Dr. Sumathi, P.
Dept. of Oilseeds,
Centre for Pl. Breeding & Genetics,
Tamil Nadu Agril. University
Coimbatore-641 003
Tamil Nadu

ID No.1928
Dr. Sumitha. P.
Jr. Pathologist,
Nimbkar Agril. Research Institute,
P.B. No 44
Phaltan-415 502
Maharashtra

ID No.1929
Dr. Sunil Ramchandra Ghorpade
Jr. Research Asst.,
AICRP (Sunflower),
Agril. School Compound,
93, Bhavani Peth
solapur-413 002
Maharashtra

ID No.1930
Dr. Supravo Gupta
Retd. Jt. Director of Agriculture,
Chhaya Neer,
Pragatinagar,
Post: Chinsurah R.S.
Hoghly-712 102
West Bengal

ID No.1931
Mr. Surendar Lal M. Seth
Block No.2,
Indian Marchantile Mansion,
24, Madam Cama Road,
Museum
Mumbai-400 039
Maharashtra

ID No.1932
Dr. Surender Kumar
Scientist (Ent.),
Pocket, A-2, Sector-8,
Rohini, H.No. 185
New Delhi-110 085
New Delhi

ID No.1933
Dr. Surendra Kumar Srivastava
Asst. Professor (Castor),
Oilseeds Section,
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.1934
Dr. Suresh G. Mantur
Pathologist,
AICRP on Sunflower,
University of Agril. Sciences,
GKVK
Bengaluru-560 065
Karnataka

ID No.1935
Dr. Suresh, M.
Scientist (Pathology),
AICRP on Safflower,
Agril. Research Station
Tandur-501 141
Andhra Pradesh

ID No.1936
Dr. Suresh, G.
Sr. Scientist (Agronomy),
Directorate of Oilseeds Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1937
Mr. Suryanarayana, L.
Ex-SRF,
Directorate of Oilseeds **Research**,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1938
Dr. Suseelendra Desai
Principal Scientist (Pl. Path),
CRIDA, Santoshnagar, Saidabad
Hyderabad-500 659
Andhra Pradesh

ID No.1939
Dr. Sushil Kumar Swarnakar
Dept. of Seed Technology,
C.S. Azad University of Agril. & Tech.,
Kanpur-208 002
Uttar Pradesh

ID No.1940
Dr. Sushma Kumari, P.
Associate Professor,
Agril. Research Station
Kayamkulam-690 502
Kerala

ID No.1941
Dr. Sushma Neema
Jr. Pathologist
PC Unit (S&N)
J.N. Krishi Vishwa **Vidyalaya**
Jabalpur-482 004
Madhya Pradesh

ID No.1942
Dr. Sverup John
Associate Professor,
Rice Research Station
Kayamkulam-690 502
Kerala

ID No.1943
Mr. Swamy, R.V.
19-Abhiyantha Colony,
Rukmini Nagar Post
Amaravati-444 606
Maharashtra

ID No.1944
Dr. Syam Kumar, D.
SRF
Directorate of Oilseeds **Research**,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1945
Dr. Syamsunder Joshi
23, RBI Colony,
Anandnagar
Bengaluru-560 024
Karnataka

ID No.1946
Dr. Syamsunder Meena
Scientist (Pl. Breeding),
Directorate of Rapeseed-**Mustard**
Research
Sewar Farm
Bharatpur-321 303
Rajasthan

ID No.1947
Dr. Talwar, H.S.
Principal Scientist (Pl. Breeding),
Directorate of Sorghum Research
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1948
Dr. Tambe Sachidanand Irappa
Sr. Research Asst.,
Vardhaman Nagar,
Plot No. 25/B, Shelgi, Dt.
Solapur-413 006
Maharashtra

ID No.1949
Mr. Tarakeshwari, M.
Jr. Research Fellow,
Directorate of Oilseeds Research
Rajendranagar
Hyderabad
Hyderabad

ID No.1950
Mr. Tasneem Naheedni Khan
Associate Professor,
Dept. of Food & Nutrition,
College of Home Science,
Marathwada Agril. Univrsity
Parbhani-431 402
Maharashtra

ID No.1951
Dr. Tej Bahadur
Brindavan,
73 HIG, Sector "E"
Shyamnagar
Kanpur-208 013
Uttar Pradesh

ID No.1952
Dr. Tejavathi, G.
Scientist, 108,
Gayatri Vihar,
Near New Darpan Colony,
Thatipur
Gwalior-474 011

ID No.1953
Dr. Tejbir Singh
Head, Department of Agril. **Botany**,
Kisan Post Graduate College
Sumbhaoli
Ghaziabad
Uttar Pradesh

ID No.1954
Dr. Thakare, A.Y.
Linseed Research Unit,
College of Agriculture
Nagpur
Maharashtra

ID No.1955

Dr. Thakore Anil, V.
Managing Director,
FORSBERG Agri-tech India Pvt. Ltd.,
315, Race Course Circle
Baroda-390 025
Gujarat

ID No.1956

Dr. Thakral, N.K.
Scientist (Oilseeds Section),
Dept. of Pl. Breeding,
CCS Haryana Agril. University
Hisar-125 004
Haryana

ID No.1957

Dr. Thakur, B.S.
Scientist (Ent.),
Indira Gandhi Krishi Vidyapeeth
Raipur-492 012
Chhattisgarh

ID No.1958

Dr. Thakur, D.S.
Scientist (Soil Science),
S.G College of Agriculture &
Research Station, Kumhrawad
Jagdalpur-494 005

ID No.1959

Dr. Thakur, K.S.
Scientist (Agronomy),
CSK H.P. Krishi Vidyapeeth,
Oilseeds Research Station
Kangra-176 001
Himachal Pradesh

ID No.1960

Dr. Thakur, N.S.
Jr. Scientist (Agronomy),
Zonal Agril. Research Station
Chindwara-482 001
Madhya Pradesh

ID No.1961

Dr. Thirugnana Kumar
Lecturer, Dept. of Genetics & Pl.
Breeding
Faculty of Agriculture,
Annamalai University, Annamalai^{naga},
Madras-608 002
Tamil Nadu

ID No.1962

Mr. Thirumala Rao
Student, ANGRAU,
College of Agriculture
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1963

Dr. Thiyagarajan, K.
Director,
Centre for Plant Breeding & ^{Genetics,}
Tamil Nadu Agril. University
Coimbatore-641 003
Tamil Nadu

ID No.1964

Dr. Thulasi Ram, K.
Training Associate (Pl. Protection),
Krishi Vigyan Kendra,
P.B.No. 24, University of Agril. Sciences^{es}
Raichur-584 101
Karnataka

ID No.1965

Dr. Thyagaraju, H.
Sunflower Breeder,
Suntech Seeds Pvt. Ltd.,
City Press Compound,
G-7, Near Sangam Talkies
Bellary-583 101
Karnataka

ID No.1966

Dr. Tikendra T. Patel
Asst. Research Station,
Anand Agril. University,
Gulab Residency,
Sola Village, Science City Road

ID No.1967

Dr. Tiwari, S.P.
Dy. Director General (Edn), (Retd.)
Indian Council of Agril. Research,
Krishi Anusandhan Bhawan-I, Pusa
New Delhi-110 012
New Delhi

ID No.1968

Dr. Tiwari, V.K.
Jr. Scientist (Oilseeds),
S/o S.N. Thiwari,
63, Bhariyaghat
Sagar-47D 002
Madhya Pradesh

ID No.1969

Dr. Toprope, V.M.
Asst. Breeder (G'Nut),
Oilseeds Research Station,
Marathwada Agril. University
Latur-413 512
Maharashtra

ID No.1970

Dr. Tushar Pimpale, D.
General Manager,
Ecomax Agro Systems Ltd.,
Satya Bhavan Building,
C. Raghunathnagar, Wagalay ^{Estate}
Thana West
Mumbai-400 064
Maharashtra

ID No.1971

Dr. Ubale, S.S.
Jr. Research Assistant,
Cotton Project,,
Mahatma Phule krishi Vidyallaya,
Dist. Ahmednagar
Rahuri-413 722
Maharashtra

ID No.1972

Dr. Uday Deshmankar
C/o Dr. Jagadeesh Singh,
Safflower Breeder,
AICRP (Safflower),
College of Agriculture
Indore-452 071
Madhya Pradesh

ID No.1973
Dr. Uke, P.C.
Jr. Agronomist (Safflower),
Oilseeds Research Unit,
Dr. P.D. Krishi Viswa Vidyalaya
Akola-444 104
Maharashtra

ID No.1974
Dr. Ulemale, R.B.
Jr. Agronomist,
Dr. P.D. Krishi Viswa Vidyalaya
Akola-444 104
Maharashtra

ID No.1975
Dr. Uma, D.
Jr. Biochemist,
Dept. of Oilseeds,
Tamil Nadu Agril. University,
Chettair Thottam, Thindiyalur
Coimbatore-641 034
Tamil Nadu

ID No.1976
Dr. Umapathy, G.
Associate Professor,
Dept. of Entomology,
Tamil Nadu Agril. University
Coimbatore-641 003
Tamil Nadu

ID No.1977
Dr. Umapathy, P.N.
Professor (Farm Management),
Main Agril. Research Station,
University of Agril. Sciences
Dharwad-580 005
Karnataka

ID No.1978
Dr. Umashankara Sharma
Associate Professor (Ent.),
Rajasthan Agril. University, MPUA
Udaipur-313 001
Rajasthan

ID No.1979
Dr. Umate, S.M.
Asst. Professor,
Genetics & Pl. Breeding,
Lokmanyagar, Karegon Road
Parbhani-431 402
Maharashtra

ID No.1980
Dr. Umesh Kumar Singh
Jr. Scientist, Q. No. Old C-14,
Near Rajendra Shisu Sadar School,
Rajendra Agril. University, Pusa
Samastipur-348 125
Bihar

ID No.1981
Mr. Usha Kiran, B.
A1-101, Tirtha Apartments,
Hyderguda, Near Eshwar Theatre
Hyderabad-500 027
Andhra Pradesh

ID No.1982
Dr. Usha Kumari, P.
Associate Professor (Pl. Breeding),
Tapioca and Castor Research Station
Tamil Nadu Agril. University,
Yethapur-636 119
Tamil Nadu

ID No.1983
Dr. Vaghani, J.J.
Agril. Research Station,
Gujarat Agril. University
Amreli-365 601
Gujarat

ID No.1984
Dr. Vaghasia, D.R.
Jr. Agronomist,
Agril. Research Station,
Gujarat Agril. University
Amreli-365 601
Gujarat

ID No.1985
Dr. Vaghasia, P.M.
Agril. Officer,
"Shyam", 19, Patelnagar,
Zanzarda Road
Junagadh-362 001
Gujarat

ID No.1986
Dr. Vajpeyi Madhu
Chemist (Linseed),
Dept. of Biochemistry,
C.S. Azad University of Agril. & Tech,
Kanpur-208 002
Uttar Pradesh

ID No.1987
Dr. Vakade, M.M.
Oilseeds Research Unit,
Dr. Punjabrao Krishi Viswa Vidyalaya,
Akola-444 104
Maharashtra

ID No.1988
Dr. Vala, G.S.
Jr. Agronomist,
Agril. Research Station,
Gujarat Agril. University
Amreli-365 601
Gujarat

ID No.1989
Dr. Vanaja, M.
Scientist (Pl. Physiology),
CRIDA, Santoshnagar, Saidabad
Hyderabad-500 659
Andhra Pradesh

ID No.1990
Dr. Vani Sekhar
Proagro Seeds Co. Ltd., 8-1-39,
Qutub Shahi Tombs Road,
Tolichowki
Hyderabad-500 008
Andhra Pradesh

ID No.1991
Dr. Varadarajan, P.V.
Principal Scientist,
Central Institute for Res. on Cotton
Tech.,
Adenwala Road, Matunga
Mumbai-400 019
Maharashtra

ID No.1992
Dr. Varaprasad, K.S.
Officer Incharge,
NBPGR Regional Research Station
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1993
Mr. Varsha C. Joshi
SRF, NOVOD Board Project,
AICRP on UUC,
S.D. Agril. University
S.K. Nagar-385 506
Gujarat

ID No.1994
Dr. Varshney, S.K.
Sr. Scientist,
Dept. of Seed Technology,
Tirhut College of Agriculture, Dholi
Muzaffarpur-843 121
Bihar

ID No.1995
Dr. Vasanthi, R.P.
Sr. Scientist (Pl. Breeding),
S.V. Agril. College,
Regional Agril. Research Station
Tirupati
Andhra Pradesh

ID No.1996
Ms. Vasavi, S.
Jr. Research Fellow,
Directorate of Olive & Sesame Research,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.1997
Dr. Vasudevan, S.N.
Associate Professor,
College of Agriculture,
University of Agril. Sciences,
Raichur-584 101
Karnataka

ID No.1998
Dr. Vavadia, L.D.
Entomologist,
Junagadh Agril. University
Junagadh-362 001
Gujarat

ID No.1999
Dr. Veena Bhatnagar
Associate Professor (Linseed),
AICRP (Linseed),
Rajasthan Agril. University
Kota-324 001
Rajasthan

ID No.2000
Dr. Veerabhadra Rao, K.
Principal Scientist (Soil Science),
DCMS Buildings,
Agril. Research Station
Anantapur-1
Andhra Pradesh

ID No.2001
Dr. Vekaria, K.D.
Asst. Research Scientist,
99, Shriji Krupa, Behind Lalbang,
Gitanagar
Junagadh-362 001
Gujarat

ID No.2002
Dr. Veladhadi, P.V.
Jr. Bio-Chemist,
AICRP on Sunflower,
Regional Research Station
Raichur-505 101
Karnataka

ID No.2003
Dr. Vemana, K.
Scientist,
Agril. Research Station,
Kadiri, Anantapur Dt.
Kadiri-515 591
Andhra Pradesh

ID No.2004
Mr. Venkata Ramana Rao
D.No. 11-11-6-3,
Nukalamma Temple Street,
Ramraopet,
Kakinada-533 004
Andhra Pradesh

ID No.2005
Dr. Venkatachalam, S.R.
Asst. Professor,
Tapocia-Castor Research Station,
Tamil Nadu Agril. University
Yethapur-636 119
Tamil Nadu

ID No.2006
Mr. Venkatesh, H.
Research Manager (Sunflower),
Monsanto Tech. India Ltd.,
Moka Road, Srivara Village
Bellary-583 103
Karnataka

ID No.2007
Dr. Venkateshan, S.
Professor & Head,
Tapocia Castor Research Station
Yethapur-636 119
Tamil Nadu

ID No.2008
Dr. Venkateswarlu, B.
Director,
Central Research Institute for Dryland
Agriculture
Santoshnagar, Saidabad
Hyderabad-500 659
Andhra Pradesh

ID No.2009
Dr. Venkattkumar, R.
Sr. Scientist,
Directorate of Oilseeds **Research**,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.2010
Dr. Venugopal, N.
Professor of Agronomy,
Dept.I of Agronomy,
College of Agriculture, GKVK Campus
Bengaluru-560 065
Karnataka

ID No.2011
Dr. Venugopal, R.
Chief Scientist (Oilseeds),
Dept. of Plant Breeding & Genetics,
GKVK
Bengaluru-560 065
Karnataka

ID No.2012
Dr. Verma, K.P.
Jr. Scientist,
Dept. of Pl. Pathology,
Indira Gandhi Agril. **University**,
Krishak Nagar
Raipur-492 012
Madhya Pradesh

ID No.2013
Dr. Verma, V.S.
Professor of Agronomy,
NATP/ROPS-16,
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.2014
Dr. Vidya Sagar, G.E.Ch.
Sr. Scientist (Agronomy),
Regional Agril. Research Station,
Palem, Mahaboobnagar Dt.
Palem-509 215
Andhra Pradesh

ID No.2015
Dr. Vijay K. Paradkar
Scientist (Agronomy),
Zonal Agril. Research **Station**,
Chandangaon
Chhindwara
Madhya Pradesh

ID No.2016
Dr. Vijay Kagliwal
Managing Director,
Vijay Seeds Co. Pvt. **Ltd.**,
A/9/7, Addl. MIDC
Jalna
Maharashtra

ID No.2017
Dr. Vijay Kumar, B.
Associate Professor,
H.No. 1-1-230/15/1
Chikkadapally
Hyderabad-500 020
Andhra Pradesh

ID No.2018
Dr. Vijay Kumar, S.
3-18, Fort
Raichur-584 101
Karnataka

ID No.2019
Dr. Vijay Laxmi, P.
Indian Institute of **Chemical Technology**,
Tarnaka
Hyderabad-500 007
Andhra Pradesh

ID No.2020
Dr. Vijay Singh
H.No. 174/22/412, Plot **No. 412**
IV Floor, KRK Golden **Enclave**
Attapur, Hyderguda
Hyderabad-500 030
Andhra Pradesh

ID No.2021
Mr. Vijay Yadavrao Kankal
Jr. Research Asst.,
Q.No. C-6, 97,
Ravivar Peth, Near DAU **College**
Solapur-413 002
Maharashtra

ID No.2022
Mr. Vijay, S.
Jr. Research Fellow,
Directorate of Oilseeds **Research**,
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.2023
Mr. Vijaya Bhaskar, A.
Scientist (Pathology),
H.No 3-10-206,
Reddy Colony
Hanumkonda-506 001
Andhra Pradesh

ID No.2024
Dr. Vijayalakshmi, K.
Consultant (Agri.),
Canadian Coop. Association,
Ground Floor, NCU Building, 4,
Siri Institutional Area, Khelgaon **Marg**
New Delhi-110 016
New Delhi

ID No.2025
Dr. Vijendra Das, L.D.
Professor (Plant Breeding),
Ag College & Research Institute,
Kullikulam
Vallanad-627 252
Tamil Nadu

ID No.2026
Dr. Vimala Devi, P.S.
Principal Scientist
Directorate of Oilseeds **Research**
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.2027

Dr. Vindyavarman, P.
Professor & Head (Oilseeds),
Dept. of Pl. Breeding,
Tamil Nadu Agril. University
Coimbatore-641 003
Tamil Nadu

ID No.2028

Dr. Vineet Kumar
Research Associate,
NATP-ROPS-16,
C.S. Azad University of Agril. & Tech.
Kanpur-208 002
Uttar Pradesh

ID No.2029

Dr. Vinod B. Bhallu
Agronomist,
Junagadh Agril. University
Junagadh-362 001
Gujarat

ID No.2030

Dr. Vinod Kumar
Scientist (CA),
NRC for Rapeseed-Mustard
Sewar Farm
Bharatpur-321 303
Rajasthan

ID No.2031

Dr. Vinod Kumar
Sr. Research Fellow,
Dept. of Soil Science, SVBPUA&T
Modipuram, Meerut
Modipuram
Uttar Pradesh

ID No.2032

Dr. Violet Kerkette
HOD,
Dept. of Plant Breeding,
Birsa Agril. University
Kanke
Ranchi
Jharkhand

ID No.2033

Dr. Virender Sardana
Agronomist,
Oilseeds Section,
Dept. of Pl. Breeding & Genetics,
Punjab Agril. University
Ludhiana-141 004
Punjab

ID No.2034

Dr. Virender Singh Lather
Professor (Pl. Breeding),
10, Type House,
CCS Haryana Agril. University
Hisar-125 004
Haryana

ID No.2035

Dr. Virupakshappa, K.
National Manager,
Advanta India Ltd., 405, 4th Floor,
"A" Wing, Carltan Towers,
1, Airport Road
Bengaluru-560 008
Karnataka

ID No.2036

Dr. Vishnuvardhan Reddy, A.
Professor (Oilseeds),
Regional Agril. Research Station,
ANG Ranga Agril. University,
Palem, Mahaboobnagar Dt.
Palem=509 215
Andhra Pradesh

ID No.2037

Dr. Vivek Prakash Nagaiah
Jr. Breeder,
C.S. Azad University of Agril. & Tech.,
Crop Research Farm,
Mauranipur
Jhansi-210 432
Uttar Pradesh

ID No.2038

Dr. Vrijendra Singh
Safflower Breeder,
Nimbkar Agril. Research Institute,
P.B. No.44
Phaltan-415 502
Maharashtra

ID No.2039

Dr. Vyas, A.K.
Principal Scientist,
Directorate of Soybean Research
Khandwa Road
Indore-452 017
Madhya Pradesh

ID No.2040

Dr. Wankhade, R.R.
Breeder,
Bioseed Research India Pvt.Ltd.,
Plot No.206, H.No.8-2-293/82/206,
Road No.14, Jubilee Hills
Hyderabad-500 033
Andhra Pradesh

ID No.2041

Dr. Weginwar, D.G.
Sunflower Breeder,
A/11, Trimuti Apartments,
Opp: Dr. Tople, Tatharpeth
Akola-444 104
Maharashtra

ID No.2042

Dr. Yadav, C.K.
Sr. Scientist (Pl. Breeding),
Regional Research Station,
CCS Haryana Agril. University
Bawal-123 501
Haryana

ID No.2043

Dr. Yadav, D.K.
Sr. Scientist
Division of Genetics
Indian Agril. Research Institute, Fusa
New Delhi-110 012
New Delhi

ID No.2044

Dr. Yadav, R.A.
Crop Research Farm,
Mauranipur
Jhansi-284 204
Uttar Pradesh

ID No.2045
Dr. Yadava, J.S.
Regional Director,
Regional Research Station,
CCS Haryana Agril. University
Bawal-123 501
Haryana

ID No.2046
Dr. Yadu, Y.K.
Sr. Scientist (Ent),
Dept. of Entomology,
College of Agriculture
Raipur-492 001
Chhattisgarh

ID No.2047
Dr. Yakadri, M.
Associate Professor (Agronomy),
Department of Agronomy,
College of Agriculture, ANGRAU
Rajendranagar
Hyderabad-500 030
Andhra Pradesh

ID No.2048
Dr. Yamini, K.N.
, Ph.D. Scholar
22, Upstairs,
Brindavanam
Pondicherry-605 013
Kerala

ID No.2049
Dr. Yashpal Yadav
Scientist (Pl. Breeding),
Regional Research Station,
CCS Haryana Agril. University
Bawal-123 501
Haryana

ID No.2050
Dr. Yassin Mohd.
Associate Professor,
Dept. of Oilseeds,
T.N. Agril. University
Coimbatore-600 003
Tamil Nadu

ID No.2051
Dr. Yogendra Kumar
Asst. Professor,
143, Keshav Vihar,
Saipath Gopalpura bypass
Jaipur-302 018
Rajasthan

ID No.2052
Dr. Zade, G.J.
Dr.P.D. Krishi Vidyapeeth,
Oilseeds Section
Akola-444 104
Maharashtra

ID No.2053
Mr. Zaveri, P.P.
H-96, Ratilal Park Society,
Darpan Six Roads,
Navrangapura
Ahmedabad-380 014
Gujarat

ID No.2054
Dr. Zirpe, A.G.
A/10, Hari Zmruthi Apartments,
Mandale Garage Road,
27/21, Erandawana
Pune-411 064
Maharashtra

GUIDELINES TO THE CONTRIBUTORS

The contributions in the form of full papers and short communications, based on original research relating to basic and applied aspects of oilseed crops in the disciplines of Genetics and Plant Breeding, Biotechnology, Agronomy, Entomology, Plant Pathology, Crop Physiology, Soil Sciences, Chemistry, Biochemistry, Economics and Extension including post-harvest technology will be considered for publication in the **Journal of Oilseeds Research** only from members of the ISOR. The reviews on current topics and recent books will also be published. The articles submitted for publication must not contain data older than 5 years on the date of receipt of the article in the society office. The period shall be reckoned from the following January and July after the completion of the field experimentation in *kharif* and *rabi* seasons, respectively.

Manuscripts, in triplicate, neatly typed in double space on one side of the white paper (A4 size) can be submitted through the Registered Post to the **Chief Editor, Journal of Oilseeds Research, Directorate of Oilseeds Research, Rajendranagar, Hyderabad-500 030, Andhra Pradesh (India)**. The revised manuscript must accompany a CD (only CD is allowed) having article typed and saved in MS Word. Chief Editor can be contacted at e-mail: harvirn@gmail.com

The Title of the paper should be concise but self explanatory. A short running title should also be given. It should be followed by a list of authors (names and addresses). The manuscript of paper should clearly define aims and objectives of the study and include the relevant review of literature. **Material and methods** should be clear and to the point. In case of well known methods, only the reference will suffice. **Results and discussion** should preferably be combined to avoid repetition. Results should be written concisely. The data should be given only in metric system. Tables should be numbered in arabic numerals, typed on separate sheets with brief and self-explanatory titles. The data given in tables should not be repeated in figures. This should be followed by **Acknowledgements**, if any. The **References** should be arranged alphabetically by the name of the first author and then, if required, by the second and the third author and so on. The names of the journals must be full and in italics according to 'World List of Scientific Periodicals'. The number of references should be kept at minimum possible. These may be cited as below:

- Paper** **Vani, K.P. and Bheemaiah, G. 2004.** Alley cropping and green leaf manures – effective means of integrated nutrient management for sustained returns of rainfed castor, *Ricinus communis* L. *Journal of Oilseeds Research*, **21**(1):73-77.
- Book** **Trenbath, T. 1986.** Resource use by intercrops. In *Multiple Cropping Systems* (ed. Charles A. Francis). Macmillan Publishing Company, New York.
- Chapter** **Hanumantha Rao, C. and Chakrabarthy, S.K. 1997.** Castor. In *Efficient management of dryland crops: Oilseeds* pp.257-272 (eds. R.P. Singh, P.S. Reddy and V. Kiresur) Indian Society of Oilseeds Research, Hyderabad.
- Thesis** **Satyanarayana, K.V. 2000.** Genetic analysis of elite inbred lines using L x T design and modified TTC model in sunflower (*Helianthus annuus* L.). M.Sc. (Ag.) Thesis, Acharya N.G. Ranga Agricultural University, Hyderabad.

The citation of reference in the text should be as Prasad and Nath (1985) or (Prasad and Nath, 1985), depending upon the composition of the sentence. Two or more than two references cited jointly should be arranged alphabetically in ascending order of years of publication and distinguished from each other by semi-colon. More than two authors should be referred to by using *et al.* with the name of the first author. Complete scientific name of crop/organism with its authority must be given on its first mention.

Illustrations: Figures and photographs should be submitted in duplicate along with typewritten titles on separate sheet. Photographs should be on high quality glazed paper with good contrasts. The figures and photographs should fit in A4 size paper and must be included in the softcopy CD submitted along with the revised article.

Certificate that the papers submitted to the **Journal of Oilseeds Research** have not been submitted to any other journal for publication. The responsibility for duplication in publishing, a full paper or part of it in any other journal, lies entirely with the author(s). A certificate from Head of Department along with signature of all the authors indicating the years of work done and their consent to publish in Journal of Oilseeds Research should be sent along with the article.

The Editorial Board assumes no responsibility for the views and statements of the authors published in the Journal.

----> For style of papers, consult the recent issue <----

Note: CD containing the manuscript is a must along with revised article.

Printed & Published by **Dr. H. Basappa**, General Secretary on behalf of The Indian Society of Oilseeds Research,
Directorate of Oilseeds Research, Rajendranagar, Hyderabad-500 030, India,
Ph : (040) 24016141/24015345; Fax : (040) 24017969

Web site: <http://www.isor.org.in>; E-mail: oilseedsociety@gmail.com

Computer Typesetting : **Sasi Graphics**, Rajendranagar, Hyderabad Printed by **M/s Progressive Press Pvt. Ltd**, Hyderabad